

Comparative Study between Conventional and Thermoreversible Mucoadhesive Nasal Gels of Midazolam Hydrochloride

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Abstract The aim of the present study was to prepare conventional and thermoreversible or *in situ* mucoadhesive nasal formulations of midazolam hydrochloride (HCl) prepared from a natural mucilage isolated from *Linum usitatissimum* L. seeds and to compare the various parameters like texture profile analysis, viscosity, mucoadhesive strength, *ex vivo* permeation study and *in vivo* drug absorption study. The formulations were prepared using three different concentrations of the isolated mucilage and sodium taurocholate was used as a penetration enhancer to obtain rapid delivery of the drug into systemic circulation.

Keywords Mucoadhesive, Nasal, Mucilage

1. Introduction

Seizures are a common medical emergency, accounting for 1%–2% of all emergency department (ED) visits, and status epilepticus (SE) exists in approximately 6% of these encounters[1]. SE is associated with high morbidity and mortality. Common complications of SE include aspiration, anoxic brain injury, cardiac instability, metabolic and autonomic dysfunction, and direct neuronal damage[2-9].

Midazolam is administered via intranasal (IN) and buccal routes but has not been developed for rectal administration[10-28]. Intranasal midazolam is an effective medication that can be given safely. Midazolam is also effective after buccal administration but has not been studied in the pre-hospital setting[24-28]. Midazolam is water soluble but becomes fat-soluble at physiological pH, allowing it to cross the nasal mucosa into adjacent tissues including the cerebrospinal fluid, resulting in rapid onset of action[29]. Moreover, the convenience of IN administration and the social acceptability may make IN midazolam the preferred treatment of seizures in the prehospital setting.

A few studies have compared diazepam and midazolam in the prehospital or emergency department setting and have shown midazolam to be equally or more effective in treating seizures, sometimes with fewer adverse effects[10,11,15-20, 29-31]. Fisgin et al compared intranasal midazolam with

rectal diazepam (RD) in the ED setting[19]. Intranasal midazolam was more successful in controlling seizure activity. Bhattacharyya et al[20] compared intranasal midazolam or RD in children and found seizure control to be faster with intranasal midazolam, with fewer adverse effects. In both studies, study medication was given by a physician in an ED setting. Holsti et al conducted a randomized trial comparing intranasal midazolam with RD and reported that Intranasal Midazolam Mucosal Atomization Device (IN-MMAD) controlled seizures better than RD in the pre-hospital setting, due to ease of administration and more cost effectiveness[10].

However, the above studies were conducted with either solution or spray dosage forms of midazolam[32]. Mucoadhesive nasal gels provide a firm platform from which drug diffuses into systemic circulation through nasal mucosa[33]. But conventional gels suffer from the disadvantage of high viscosity and hence cannot be administered properly to the nasal cavity and accurate dose of drug cannot be delivered[34]. This will affect the underlying bioavailability of the drug adversely. To overcome this problem, idea of thermoreversible or *in situ* gels was conceived.

In situ gels are basically liquid at room temperature and convert to a firm gel when instilled into the nasal cavity[34-37]. Hence, accurate dose is delivered via this route. In our previous works we have shown that mucoadhesive gels of midazolam hydrochloride prepared from *Linum usitatissimum* L. mucilage showed better rheological and mechanical properties as well as better drug release profiles and *in vivo* drug absorption pattern than those prepared with

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synthetic polymers.

The aim of the present study is focused on comparison between mucoadhesive conventional and *in situ* nasal gels containing midazolam hydrochloride prepared from *Linum usitatissimum* L. mucilage.

2. Materials and methods

2.1. Materials

Midazolam hydrochloride was obtained as a gift from Sun Pharmaceutical Industries Ltd., Gujarat, India. Flaxseeds were purchased from local market. Pluronic F127 was purchased from Sigma Aldrich Pvt. Ltd., India. Sodium taurocholate was purchased from Loba Chemie Pvt. Ltd., Mumbai, India. All other reagents and chemicals used were of analytical grade.

2.2. Preparation of Mucoadhesive Nasal Gel

Conventional mucoadhesive nasal gels were prepared by dissolving midazolam hydrochloride in nasal solution (0.65% NaCl, 0.04% KH_2PO_4 , 0.09% K_2HPO_4 and 0.02% benzalkonium chloride) (pH 6)[38] in a constant stirring condition. Required amounts of LUM was added to the solution and stirred on a magnetic stirrer until a uniform solution was obtained which was kept at 4°C overnight to allow complete swelling so that a homogenous gel was formed. Penetration enhancers (sodium taurocholate) were also added to the formulations at a concentration of 0.50 % (w/v).

Table 1a. Composition of Conventional Nasal Gels

Formulation code	Midazolam HCl (%w/v)	<i>Linum usitatissimum</i> L. mucilage (LUM) (%w/v)	Nasal solution
LUM 1	5.0	3.0	q.s.
LUM 2	5.0	4.0	q.s.
LUM 3	5.0	5.0	q.s.

Table 1b. Composition of *In situ* Nasal Gels

Formulation code	Midazolam HCl (%w/v)	Pluronic F127 (%w/v)	<i>Linum usitatissimum</i> L. mucilage (%w/v)	Nasal solution
LUM 1 (P)	5.0	16.0	0.5	q.s.
LUM 2 (P)	5.0	16.0	1.0	q.s.
LUM 3 (P)	5.0	16.0	1.5	q.s.

In situ gels are prepared by cold method[39]. A 16% w/v Pluronic F127 was prepared in a mixture of propylene glycol/ water (45:55), and then LUM was added to it in three different concentrations of 0.5%, 1.0% and 1.5% w/v, respectively. Midazolam was then added to the dispersion, which was kept at 4°C overnight for complete swelling to form a homogenous gel. Formulations containing enhancer (0.5% w/v sodium taurocholate) were also prepared to study the effect of enhancer on drug release pattern. Formulation codes of the gels are provided in Table 1.

2.3. Determination of Viscosity

Viscosities of different formulations were measured with

TV-10 viscometer (Toki Sangyo Co. Ltd., Japan) at five different speeds of 10, 20, 30, 60 and 100 rpm, respectively, using spindle M4 and cord no. 23 at $37 \pm 1^\circ\text{C}$ [8]. The corresponding viscosities were plotted against spindle rpm as depicted in Figure 1.

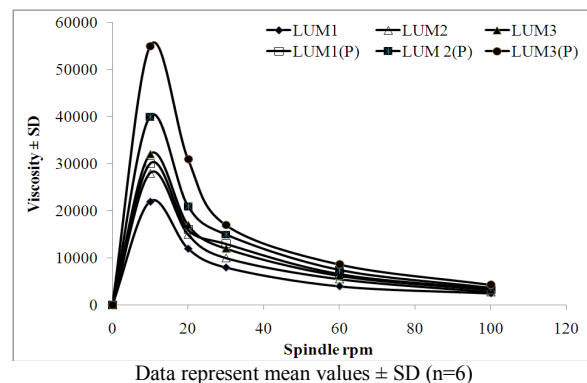


Figure1. Comparative study of viscosities of formulations at $37 \pm 1^\circ\text{C}$

2.4. Determination of Mechanical Properties

Texture profile analysis of the formulations determines mechanical properties of a gel like hardness, adhesiveness and cohesiveness. These parameters were determined using QTS-25 Texture Analyser (Brookfield Engineering Labs, USA). The method involves depression of an analytical probe of diameter 1.2 cm twice into each sample to a defined depth (15 mm), at a defined rate (30 mm/min), with a defined recovery period (15 s), between the end of the first compression and the beginning of the second. A trigger force of 4 g was applied. At least six analyses of each sample were performed at $37 \pm 1^\circ\text{C}$. Data collection and calculation were done by Texture pro software, version 2.1[38]. The analysis was done using a 5-kg load cell instrument.

2.5. Determination of Mucoadhesive Strength

Mucoadhesive strength of each formulation was determined by measuring force required to detach nasal mucous membrane from the formulation using the same texture analyser. The goat nose was collected from local slaughterhouse within 15 min after the goat was sacrificed. After removing the skin, the nose was stored on ice cold phosphate buffer (pH 7.4, 0.05 M). The septum was fully exposed, and nasal mucosa was carefully removed using forceps and surgical scissors. The mucosal tissues were immediately immersed in Ringer's solution[33,37,38]. Freshly excised goat nasal membrane was attached to the upper probe of the instrument, and fixed amount of formulation was kept below that. The upper probe was then lowered at a speed of 10 mm/min to touch the surface of the gel. A force of 0.1 N was applied for 5 min to ensure intimate contact between the membrane and the gel. The surface area of exposed mucous membrane was 1.13 cm^2 .

2.6. Ex vivo Drug Permeation Study

Based on texture profile analysis and drug release pattern of the formulations, LUM1 and LUM1 (P) containing 0.5%

sodium taurocholate were selected for *ex vivo* study.

Ex vivo permeation study was conducted using a Franz diffusion cell containing 100 ml of phosphate buffer (pH 6, 0.1 M) using an excised goat nasal mucosa. The freshly excised nasal mucosa was mounted on the diffusion cell. 100 μ l of gel containing 5 mg midazolam was placed on the membrane dispersed in 100 ml phosphate buffer and stirred at a constant rate by a PTFE-coated magnetic bar at 600 rpm. Cells were kept under constant oxygen flow (95% O₂, 5% CO₂). Throughout the study, the buffer solution in the chamber was maintained at 37 \pm 1°C by connecting the Franz diffusion cell with water bath. At predetermined time intervals, 1 ml of the sample was withdrawn at a time and replenished with an equal amount of phosphate buffer. The samples were diluted appropriately and filtered. Absorbances of the samples were measured spectrophotometrically at 218 nm using Jasco V-550 UV/Vis Spectrophotometer (Tokyo, Japan), taking phosphate buffer (pH 6) as the blank.

The amount of drug permeated was calculated from the calibration curve (linearity range =2.4 to 120 μ g/ml; r^2 =0.9996). The mean cumulative percentage of drug permeated was plotted against time (Figure 2). Permeation area was 2.54 sq cm[38].

2.7. *In vivo* Drug Absorption Study

In vivo studies were conducted on 12 New Zealand albino male rabbits weighing between 1.5 and 2 kg. Based on texture profile analysis and drug release pattern of the formulations, LUM1 and LUM1 (P) containing 0.5% sodium taurocholate were selected for *in vivo* study. Animals were kept in individual metal cages and acclimated at 25°C for 10 days prior to the experiment. They were provided with standard diet and water *ad libitum*. The approval of the Institutional Animal Ethics Committee was obtained before starting the study, and it was conducted according to the institutional guidelines. The rabbits were kept in fasting condition for 24 h before the experiment commenced. The rabbits were grouped into three (group I, II and III), each group containing three rabbits. Group I was administered intravenous bolus injection of midazolam. Groups II and III were administered nasal gels of LUM1 and LUM1 (P) containing 0.5% sodium taurocholate. Single dose of midazolam (2mg/kg body weight of rabbit) was administered intravenously to compare the pharmacokinetic parameters. No anaesthesia was used for the intravenous study. Midazolam was injected through cannulated marginal ear vein. Two millilitres of blood samples each time were collected before intravenous injection and then at 5-, 10-, 15-, 20-, 30-, 45-, 60-, 90-, 120-, 180-, 240- and 300 min intervals in eppendorfs containing heparin sodium (100 U/ml). In case of nasal gels, the dose of midazolam that was administered was also 2 mg/kg body weight of rabbit. Before application of gel, each rabbit was lightly anaesthetized by intramuscular injection of a mixture of xylazine (3 mg/kg) and ketamine (35 mg/kg). Following induction of anaesthesia, a catheter was fixed into the central artery for blood sample collection. About 2 ml blood sample was collected prior to the application of gel and

then at 5, 10, 15, 20, 30, 45, 60, 90, 120, 180, 240 and 300 min intervals in eppendorfs as above.

After every 20 min, each rabbit was administered one-third of the initial dose of xylazine and ketamine intramuscularly to maintain a light plane of anaesthesia. The blood samples were kept on ice and centrifuged at 3,000 rpm for 10 min immediately after collection to separate the plasma and stored at -20°C until the time of analysis. Immediately after each blood sample collection, the catheter was flushed with 0.2 ml of a 10% (v/v) heparin/normal saline solution to prevent blood clotting inside the catheter[40].

All animal experiments are performed as per the standard norms and guidelines of the Animal Ethics Committee of Dr. B. C. Roy College of Pharmacy and Allied Health Sciences, West Bengal University of Technology, as recognized by the Committee for the Purpose of Control and Supervision on Experiments on Animals, India.

2.8. HPLC Analysis of Midazolam in Plasma

Reverse phase HPLC was used to quantitate Midazolam in plasma samples. Midazolam was extracted with 3 ml of cyclohexane/diethyl ether (3:7) after the addition of 10 μ l of 2% sodium hydroxide⁴¹. The organic phase was removed and evaporated to dryness under nitrogen, and the residue was reconstituted in 200 μ l of the mobile phase (10 mM phosphate buffer, pH 6.0)/acetonitrile, 80:20). Of the mixture, 100 μ l was injected for chromatographic analysis. The mobile phase was delivered into the HPLC apparatus at a flow rate of 1 ml/min (isocratic pump, Model LC-10AS, Jasco, Japan). The detection wavelength was 218 nm (ultraviolet variable wavelength detector, Model SPD-10A), and a C18 column was used.

All assays were performed at ambient temperature. Calibration curve of midazolam hydrochloride prepared in rabbit plasma was found to be linear over the concentration range of 10–1,000 ng/ml (r^2 =0.9999). Pharmacokinetic parameters like peak plasma concentration (C_{max}), time to reach peak plasma concentration (T_{max}) and area under the concentration–time curve (AUC), Mean residence time (MRT), $T_{1/2}$ and Clearance (Cl) were calculated following non-compartment model by Kinetic 4.4, PK/PD Analysis, Thermoelectron Corporation. All the parameters were calculated for i.v. bolus injection of midazolam and nasal formulations.

Fraction of dose absorbed (F) was calculated by the following equation:

$$F = \frac{AUC_{(nasal)} \times Dose_{(iv)}}{Dose_{(nasal)} \times AUC_{(iv)}} \quad (1)$$

where $Dose_{(iv)}$ =dose of midazolam given as i.v. solution, $Dose_{(nasal)}$ =dose of midazolam in nasal gels, $AUC_{(i.v.)}$ =AUC after i.v. administration of midazolam and $AUC_{(nasal)}$ =AUC after nasal administration of midazolam.

2.9. Statistics

Data were analysed by one-way ANOVA followed by Tukey's HSD test using Vassar stat software (USA). $p < 0.001$ has been considered to be significant statistically.

3. Results

3.1. Determination of Viscosity

Viscosities of different formulations at different spindle rpm at $37 \pm 1^\circ\text{C}$ are shown in Figure 1. With increase in spindle rpm, viscosity of formulations decreased. It was observed that all the LUM gels showed higher viscosity than the gels prepared from synthetic polymers.

3.2. Determination of Mechanical Properties

Texture profile analyses of the gels were studied at $37 \pm 1^\circ\text{C}$ using QTS-25 texture Analyser. The results of texture profile analysis were displayed in Figure 2. Minimum hardness was shown by LUM1 gel, and maximum was shown by LUM3(P) gel (Figure 2a). Adhesiveness of the formulations displayed in Figure 2b, showed that with increase in concentration of LUM from 3% to 5%, adhesiveness increased by 1.26 times (from -89.00 ± 7.12 to -112.08 ± 5.89 gs). And, for *in situ* gels, with increase in concentration of LUM from 0.5% to 1.5%, adhesiveness increased by 2.1 times (from -1300.75 ± 15.26 to -2940.87 ± 20.77 gs) (Figure 2b). Values of cohesiveness, shown in Figure 2c, report that for conventional gels, cohesiveness decreased from 1.01 ± 0.05 to 0.97 ± 0.03 . And for *in situ* formulations, it decreased from 1.16 ± 0.03 to 0.98 ± 0.05 . However, the formulations containing sodium taurocholate did not show any change in the values of hardness, adhesiveness and cohesiveness.

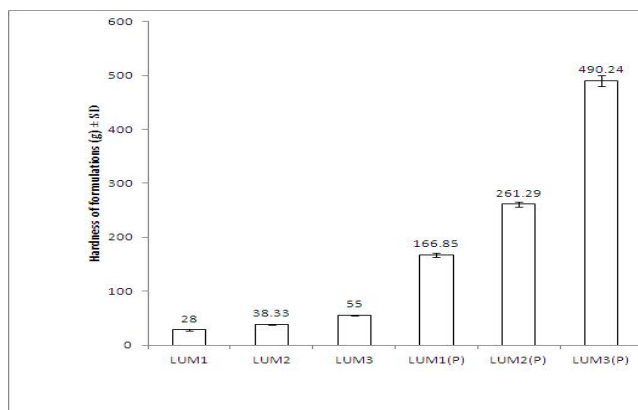


Figure 2a. Comparative study of hardness of different formulations at $37 \pm 1^\circ\text{C}$. Mean values \pm SD (n=6)

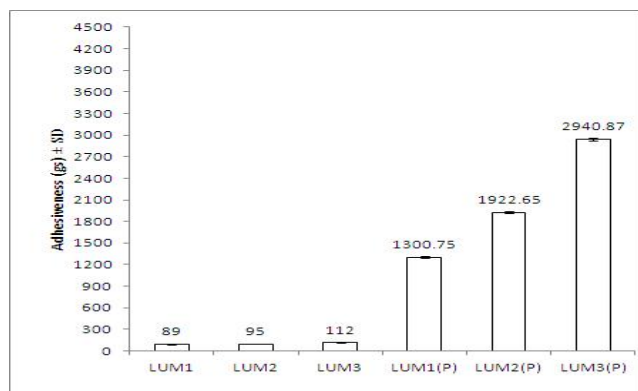


Figure 2b. Comparative study of adhesiveness of different formulations at $37 \pm 1^\circ\text{C}$. Mean values \pm SD (n=6)

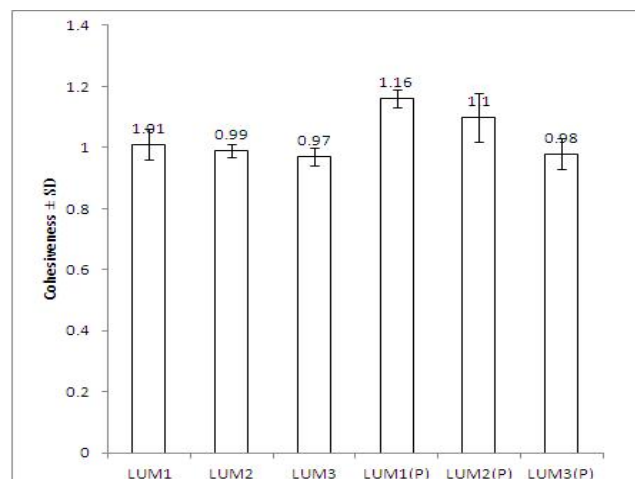


Figure 2c. Comparative study of cohesiveness of different formulations at $37 \pm 1^\circ\text{C}$. Mean values \pm SD (n=6)

3.3. Determination of Mucoadhesive Strength

Mucoadhesive strengths of the gels were also measured in texture analyzer. Results are shown in Figure 3. In case of conventional gels, with increase in concentration of LUM from 3 to 5% mucoadhesive strength increased from 16.26 ± 0.98 g to 22.45 ± 1.02 g. For *in situ* gels, the mucoadhesive strength increased from 72.26 ± 2.25 g to 92.33 ± 4.11 g, with increase in concentration of LUM from 0.5 to 1.5%.

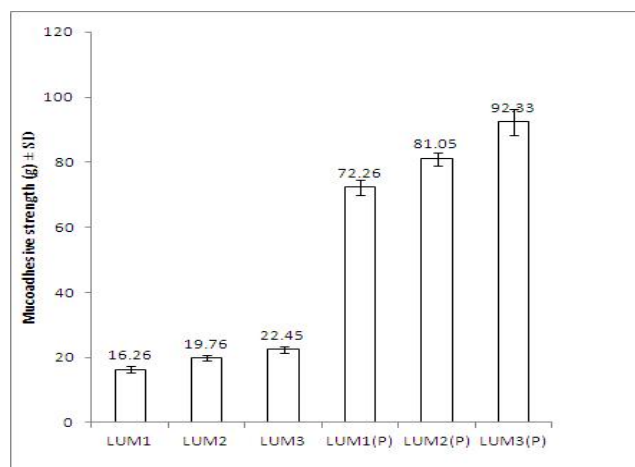


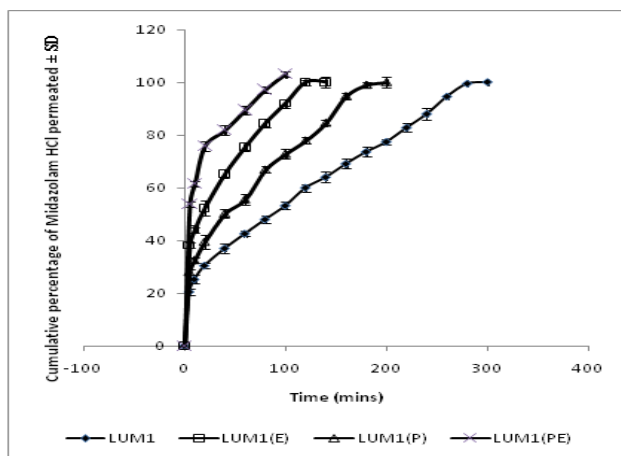
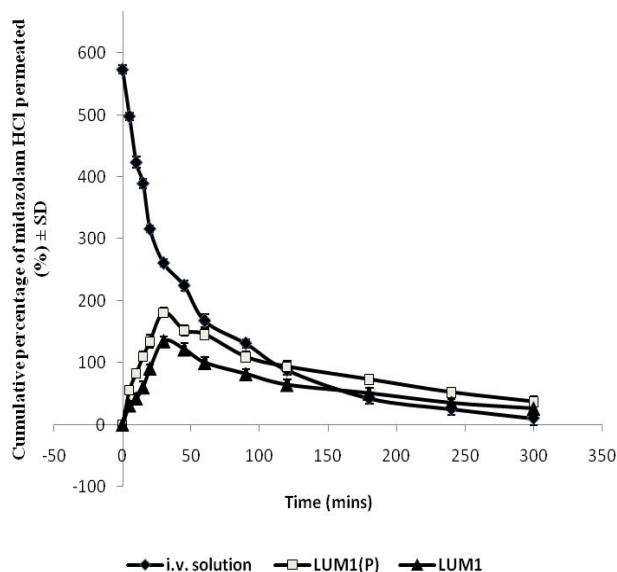
Figure 3. Comparative study of mucoadhesive strength values of formulations at $37 \pm 1^\circ\text{C}$. Data represent mean values \pm SD (n=6)

3.4. Ex vivo Drug Permeation Study

Ex vivo release profiles of midazolam from various gels were studied in phosphate buffer (pH 6.0). Considering the values of hardness, adhesiveness and cohesiveness, LUM1 and LUM1(P) were selected for *ex vivo* drug permeation and *in vivo* drug absorption studies. The formulations containing 0.5% sodium taurocholate as permeation enhancer, were named as LUM1(E) and LUM1(PE). Figure 4 depicts the drug permeation profiles of the formulations with and without enhancer. Results show that cumulative percentage of drug permeated from LUM1 and LUM1(P) were 300 and 200 min, respectively and from LUM1(E) and LUM1(PE) were 120 and 80 min, respectively.

Table 2. Comparative pharmacokinetic parameters of midazolam hydrochloride following administration of intravenous, conventional and *in situ* nasal gels in rabbits (dose = 2mg/kg). Data represent mean \pm SD (n=3)

Pharmacokinetic Parameters	Intravenous solution	LUM 1 (E)	LUM 1 (PE)
C_{max} (ng/ml)	573.64 \pm 5.23	181.12 \pm 6.21	195.03 \pm 9.25
T_{max} (min)		30.00 \pm 4.75	30.00 \pm 9.76
AUC _{last}	32671.50 \pm 90.56	26168.80 \pm 123.67	28160.80 \pm 93.67
AUC _{extra}	865.87 \pm 37.25	5546.92 \pm 48.76	7758.20 \pm 51.23
AUC _{total}	33537.40 \pm 101.25	31715.72 \pm 8.77	35919.00 \pm 250.67
MRT (min)	71.61 \pm 5.05	190.07 \pm 8.05	220.07 \pm 8.55
$T_{1/2}$ (min)	58.78 \pm 6.32	122.55 \pm 5.87	163.76 \pm 7.05
Clearance $\times 10^{-5}$ (mg/ Kg*min/ (ng/ml))	5.79 \pm 6.95	6.67 \pm 4.22	5.85 \pm 5.33

**Figure 4.** Comparative result of *ex vivo* release profiles of midazolam hydrochloride from the both *in situ* and conventional nasal gels prepared with 0.5 % *Linum usitatissimum* L. mucilage with and without 0.5% w/v of sodium taurocholate (E) in phosphate buffer (pH 6) at 37 \pm 1 $^{\circ}$ C. Data represent mean values \pm SD (n=6).**Figure 5.** *In vivo* drug plasma absorption profiles. *In vivo* drug absorption study

In vivo drug absorption study results were shown in Table 2 and Figure 5. In case of conventional formulations, C_{max} was 181.12 \pm 6.21 ng/ml, T_{max} was 30.00 \pm 4.75 min, AUC was 32715.72 \pm 98.77, MRT was 190.07 \pm 8.05 min, $T_{1/2}$ was 122.55 \pm 5.87 min and clearance was 6.67 \pm 4.22 $\times 10^{-5}$ mg/

Kg*min/(ng/ml). For *in situ* gels, C_{max} was 195.03 \pm 9.25 ng/ml, T_{max} was 30.00 \pm 9.76min, AUC was 35919.00 \pm 250.67, MRT was 220.07 \pm 8.55min, $T_{1/2}$ was 163.76 \pm 7.05 min and clearance was 5.85 \pm 5.33 $\times 10^{-5}$ mg/Kg*min/(ng/ml).

4. Discussion

Viscosity of the formulations was determined at 37 \pm 1 $^{\circ}$ C. At this temperature, LUM formulations containing Pluronic F 127 got converted to gels. Figure 1 shows viscosity profile of all formulations and they exhibit pseudoplastic flow. Moreover, with increase in concentration of mucoadhesive agent (LUM), viscosity is found to increase. However, viscosity of *in situ* gels was higher than those of conventional ones. This may be due to the presence of Pluronic F 127 in thermoreversible gels which led to the formation of a firm gel when formulation was administered into nasal cavity.

Mechanical properties of the formulations like hardness, adhesiveness and cohesiveness were evaluated by texture profile analyser. Results are shown in Figure 2. Hardness was defined as the force required to attain a given deformation or as the maximum peak force during the first compression cycle. Adhesiveness is the negative force area for the first compression cycle and represented the work required to overcome the attractive forces between the surface of the gel and the surface of the probe. Cohesiveness is the ratio of the area under the force–time curve produced on the second compression cycle to that produced on the first compression cycle, where successive compressions were separated by a defined recovery period.

It was reported that the hardness of gel formulations were significantly affected by the molecular weight and concentration of the polymer[42]. Low gel hardness ensures that the minimum work is required for removal of gels from the container and the applicability onto the desired site. However, less value decreases the retention time of gel formulation on the site of application and hence a gel should have optimum hardness value for obtaining proper therapeutic effect[43,44]. From Figure 2a, it is observed that hardness values of conventional gels are lesser than *in situ* gels. And, with increase in concentration of mucoadhesive agent, value of hardness increases for both types of gels.

For better release it is important for the gel to spread

uniformly on the nasal mucosa. So values of adhesiveness of formulations should be better. Values of adhesiveness as in Figure 2b, show that as the amount of mucoadhesive agent increases, adhesiveness of gels increases.

The cohesiveness is important to determine the reconstruction ability of the gel after application[45,46]. The high cohesiveness value increases the performance of the product at the application site by providing full structural recovery following gel application[42,44]. But Figure 2c shows that with increase in amount of mucoadhesive agent, cohesiveness values of gels decreases and this is observed both in case of conventional and *in situ* gels.

Mucoadhesive strength determines force required to detach gel applied on nasal mucosa. It is also measured with the help of Texture Profile Analyser. Mucoadhesive strength mainly depends on two factors - concentration of mucoadhesive strength and contact time of formulation with nasal mucosa[47]. It was observed that on increasing contact time beyond 5 min, there was hardly any significant change in mucoadhesive strength. Hence, evaluation of this parameter was done keeping the contact time at 5 min only. From the results shown in Figure 3, it can be deciphered that *in situ* formulations are more mucoadhesive than the conventional ones. And with increase in amount of mucoadhesive agent used, there was a corresponding increase in amount of mucoadhesive strength of the gels.

Ex vivo drug permeation study was conducted in phosphate buffer (pH 6.0) and results are shown in Figure 4. It is observed that formulations containing enhancer exhibited faster release than the ones without enhancer. This may be attributed to the fact that the enhancer which is used here is sodium taurocholate, which being a bile salt can enhance paracellular transport of drugs by opening the tight junctions of the nasal mucosa[46]. Moreover, drug release from *in situ* gels was faster than conventional ones. This happens because *in situ* gels contain Pluronic F127, a block copolymer, which enhances transport of drugs by opening tight junctions of nasal mucosa[49].

In vivo drug absorption profiles depicted in Figure 5, show that drug absorption from *in situ* gels was far better than from conventional gels. For *in situ* gels, C_{\max} was higher, AUC was higher, $t_{1/2}$ was more, MRT of drug was better and clearance was lesser than conventional gels. However, T_{\max} was same for both types of gels. From the figure, it is evident that a better plasma absorption profile of midazolam was shown by *in situ* formulations than conventional gels.

5. Conclusions

Midazolam is an important drug used widely for treatment of status epilepticus. Hence, nasal gel of midazolam will provide a firm platform from which drug will be released directly into systemic circulation and therapeutic effect can be obtained for a prolonged period. In comparison to conventional formulations, thermoreversible or *in situ* formulations provided reproducible results in all aspects as is evident

from the above study. Moreover, it is easier to administer such formulations into nasal cavity as they remain liquid at room temperature and convert to gels at nasal temperature. Thus, thermoreversible nasal gel of midazolam prepared from a natural mucoadhesive agent will provide a cost effective dosage form for the treatment of status epilepticus.

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