

Characterization of Photoprotective Compounds in Marine Zooplankton of the Southwest Coast of India: An Ecological Perspective

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Abstract In the marine environment, organisms are under threat due to several environmental fluctuations such as ultraviolet (UV) radiation, coastal pollution etc. However, it is encouraging to note that the marine organisms try to protect themselves from UV-radiation by synthesizing photoprotective compounds. Since phytoplankton can synthesize the UV-protective compounds and are grazed by zooplankton, we investigated the presence of photoprotective compounds in zooplankton. In this context, a study has been conducted for the first time in the shelf waters of the southwest coast of Arabian Sea, India for the characterization of photoprotective compounds in the zooplankton community. The study revealed the presence of photoprotective compounds, such as porphyrins and palythine in the zooplankton community of all the stations, in addition to mycosporine glycine, in the polluted waters of the stations, Veli, Neendakari and Cochin.

Keywords Arabian Sea, Mycosporine like amino acids, zooplankton, UV-protective compounds, antioxidant metabolites

1. Introduction

Impact of UV-radiation on aquatic food webs has been stimulated by realizing the increasing levels of UV-radiation reaching earth's surface[1]. Despite the low intensity at the ground level, it causes biological damage because of the high-energy content of photon, and the DNA damage via the formation of cyclobutane and pyrimidine dimers[2-6]. One possible strategy by the organisms to protect themselves against UV-radiation is by the synthesis of UV-absorbing compounds that act as natural sunscreens. One such compound, mycosporine, identified in fungi was found to have a role in UV-induced sporulation[7-10]. Mycosporine like amino acids (MAAs) are found in marine organisms, from bacteria to fish[11, 12] and in terrestrial microorganisms like fungi[13]. High concentrations of MAAs are found in epilithic cyanobacterial or algal mats[14, 15]. Accumulation of MAAs has also been reported for the population of the copepod, *Boeckella titicacae* from the tropical high altitude Lake Titicaca[16]. Concentration of MAAs in the population of *Cyclops abyssorum* and *C. abyssorum taticus* from the lakes in the Alps has been found to increase exponentially with lake elevation and underwater UV transparency.

Copepods of alpine lakes also accumulate high amounts of carotenoids that give them the intense red appearance and provide with added protection[17-19]. Concentration of these compounds in the calanoid copepods was found to be increased with lake elevation and shallowness of the system, thus establishing a direct correlation between the concentration and the UV-intensity[20]. Such environmental factors besides UV exposure are also important in regulating the synthesis of MAAs[21].

During the past two decades, a substantial loss in the stratospheric ozone layer has been noticed that has aroused interest in studying the effects of increased ultraviolet radiation (UVR), particularly UV-B radiation (280–315 nm), on the earth's surface. Solar UV-B radiation is detrimental to most sun-exposed organisms, including humans[22]. An increase in UV-B radiation has led to search for the natural photoprotective compounds from various organisms such as microorganisms, plants and animals of marine as well as freshwater ecosystems. A number of photoprotective compounds, such as melanins, MAAs, scytonemin, parietin, usnic acid, carotenoids, phycobiliproteins, phenylpropanoids and flavonoids and several other UV-absorbing substances of unknown chemical structure have been identified from different organisms[23-25].

There have been a number of reviews about diverse classes of compounds from natural sources, including marine habitats, but the occurrence of photoprotectants from marine sources has only partially been elucidated. Ultraviolet ra-

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diation (UVR) is one of the most harmful exogenous agents and may affect a number of biological functions in all sun exposed living organisms. Solar radiation exposes the organisms to harmful doses of UV-B and UV-A (315–400 nm) radiation in their natural habitats. In response to intense solar radiation, organisms have evolved positive mechanisms such as avoidance, repair and protection by synthesizing or accumulating a series of photoprotective compounds, such as MAAs, scytonemin, carotenoids and certain other compounds to counteract the toxicity of UV (particularly UV-B) radiation[26–29]. Furthermore, MAAs is the most common compounds with a potential role as UV sunscreens in marine organisms. It has been found that MAAs provides protection from UVR not only for their producers, but also to primary and secondary consumers through the food chain[30].

Production of MAAs from marine zooplankton requires holistic understanding the transfer of MAAs from primary to higher trophic levels through the marine food chain. Findings during this investigation are considered important since much of our knowledge on MAAs is from the fresh water habitat and temperate waters. Studies on the MAAs production from the shelf waters of tropical environments are lacking. Hence, an attempt was made for the first time in this regard along the shelf waters of southwest coast, India. This study investigated the relationship among zooplankton, chlorophylla, productivity and photoprotective compounds characterization in zooplankton community with the existing environmental conditions.

2. Materials and methods

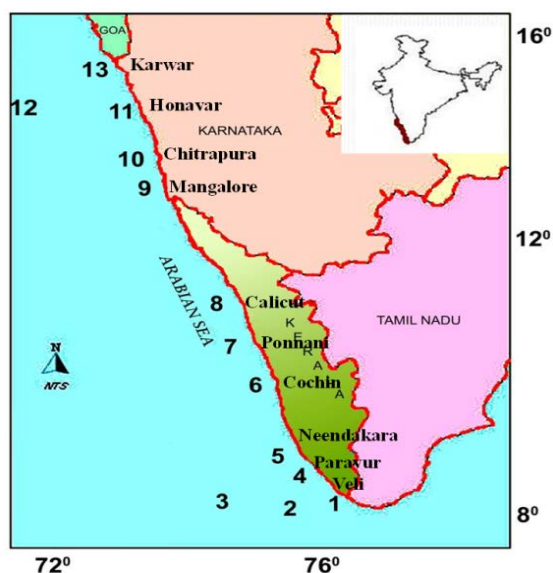
Study was carried out at thirteen stations along (~ 1400 km stretch between 8°N and 14°N lat) the shelf waters of the southwest coast of Arabian Sea, India, in which eight stations were selected from off Kerala coast and five from off

Karnataka Coast (Figure 1) Samples were collected during the *CRV Sagar Purvi* cruise from May to June 2005. The southwest coast of India is a monsoon dominated coast. Upwelling occurs along the coast during the southwest monsoon (June to September) season between 14 °N and 7 °N[31, 32].

Sub surface water samples representing the photic layer, were collected using a 5L Niskin sampler for physico-chemical parameters, such as pH, salinity, temperature, dissolved oxygen (DO), chlorophyll *a* and primary production. Temperature was recorded using a thermometer (1–51°C range within $\pm 0.1^\circ\text{C}$; Brannan, UK). pH was measured using a portable pH meter, WTW MultiLine P4, having a range of 0 to 14, resolution of 0.01 and accuracy ± 0.05 . Salinity was determined using a Digi auto salinometer (Model TSK, accuracy ± 0.1). DO was measured using Winkler's titration method of Grasshoff *et al.*[33], the analytical precision, expressed as standard deviation ($\pm 0.07\%$). 1 litre water sample was filtered through glass fiber filter (GF/F; Whatman) for chlorophyll *a* analysis. Chlorophylla retained on the filter was extracted with 90% acetone at 4°C in dark for 24h, and measured spectrophotometrically[34]. The analytical precision for chlorophylla analysis was $\pm 4\%$.

Primary production was estimated by *in-situ* method using the ^{14}C -technique[35]. Water samples were collected before the sunrise, immediately passed through 200 μ sieve to remove large sized zooplankton and transferred to 300 ml capacity Nalgene bottles (3 light bottles and 1 dark bottle). After the inoculation of 1 ml of $\text{NaH}^{14}\text{CO}_3$ (activity 5 μCi) solution, the light and dark bottles were deployed in a mooring system at respective depth for 12 h for incubation. The experiments were terminated by filtering the samples on to 47mm Whatman GF/F filters and the filters were used for subsequent analysis in a liquid scintillation counter[35] after treatment with HCl fumes to remove inorganic carbon.

Zooplankton samples were collected with a net (mesh size 100 μm) having a diameter of 0.6 m, towed horizontally just below the surface for a duration of 10 min (speed 1 knot). The net was fitted with a calibrated flow meter (General Oceanics, Model-2030) to quantify the volume of water filtered. Collected zooplankton samples were placed in two sets of prewashed (Milli-Q) polycarbonate bottles. One was stored at -4°C for the characterization of MAAs and the other was fixed with 4% formalin. Qualitative and quantitative analysis of zooplankton were performed following stringent methods[36–38]. Zooplankton samples were filtered through a Whatman GF/F filter and frozen at -80°C for subsequent extraction of MAAs. Extraction was done in tightly capped vials containing 0.75–1.5 ml of 25% aqueous methanol (24–48 h at 25°C , sonicated before analysis). The extracts were subsequently dried under vacuum in 2ml Eppendorf micro centrifugation tubes, using a SpeedVac concentrator at 4°C and stored at -80°C for further characterization. The concentrated extracts were resuspended in 0.5 to 2ml of 25% aqueous methanol (v/v) and they were analyzed chromatographically by isocratic high-performance liquid chromatography (HPLC)[39]. We used a Shimadzu



1. Veli-A, 2. Veli-B, 3. Veli-C, 4. TTP*, 5. Neendakara, 6. Cochin, 7. Kodungallur, 8. Calicut, 9. Mangalore, 10. Chitrapura, 11. Honavar-A, 12. Honavar-B, 13. Karwar * Travancore Titanium Products.

Figure 1. Study area map with sampling locations

LC-10AD/SCL-10A chromatograph with SPD-10AV stop-flow scanning spectrophotometric detector (recording 313, 340 nm) and a Brownlee C18 column (250 x 4.6 mm). For routine analyses, the mobile phase was 25% (vol:vol) aqueous methanol with 0.1% acetic acid, at 0.8 ml min^{-1} . The MAAs were identified by chromatography by comparing the absorption spectra and retention times with standards provided by Dr.Oliver Nixdorf, University of Bremen, Germany.

3. Results and Discussion

Variations in hydrographic characteristics along south-west waters were presented in (Figure 2 a-d).South west coast has a warm humid climate with $\sim 32^\circ\text{C}$ air temperature. Water column remained relatively cool (avg. $28.16 \pm 0.73^\circ\text{C}$) except station Veli. Salinity is one of the prime factors, which influences the abundance and distribution of the fauna and flora in the coastal waters. Salinity (avg. 28.94 ± 5.59) showed a wide fluctuation at stations proximal to estuaries. Low pH recorded at southern stations (Veli, Paravur, Neendakara) showed a great fluctuation among the stations. At Veli pH was 3.32, indicating the extremity of acidic factory effluents discharged from Travancore Titanium Product factory (TTP). This is in agreement with the observation-sofarlier studies[40]. Relatively high DO (avg. 5.04 ± 0.60) was observed at all stations except Veli. From these results, it is quite clear that the shelf waters of the Thiruvananthapuram coast have been exposed to the increased threat of industrial pollution.

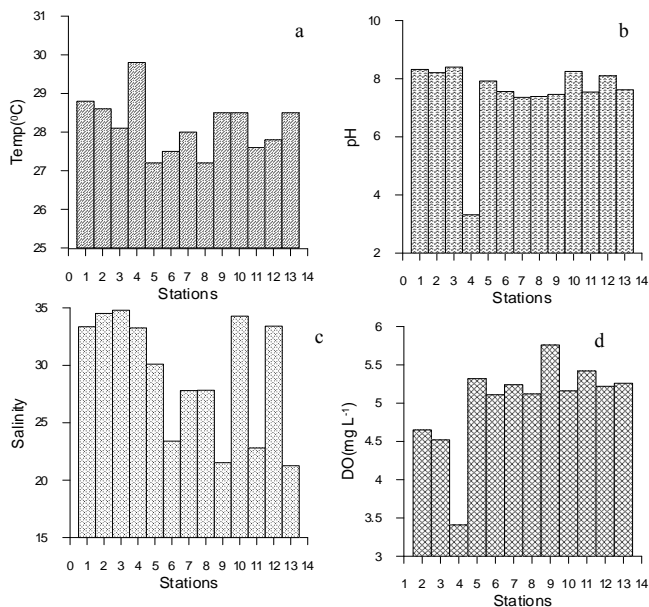


Figure 2. bSpatial variation of hydrographic characteristics along the stations (a) water temperature (b) pH (c) Salinity(d) Dissolved Oxygen

Concentration of chlorophyll *a* (Figure 3a) varied from $2.6 - 0.1 \text{ mg m}^{-3}$ with an average of $1.23 \pm 0.94 \text{ mg m}^{-3}$. High values of primary productivity of $0.34 \text{ g C m}^{-3} \text{ d}^{-1}$ was at Karwar and lowest of $0.07 \text{ g C m}^{-3} \text{ d}^{-1}$ was at Veli with an average of $0.21 \pm 0.09 \text{ g C m}^{-3} \text{ d}^{-1}$ (Figure 3b). Higher zoo-

plankton population density of 859 No m^{-3} was recorded at Calicut and lowest (64 No m^{-3}) was observed at Veli near shore (Figure 3c). Madhuprathapet al.[41] reported that death of organisms due to effluent could be due to the drastic change in pH of water, which will create stress on normal metabolism of the overall aquatic life, which was reflected in our study through the pigment concentration and primary productivity at Velitransact (Figure 3a,b).

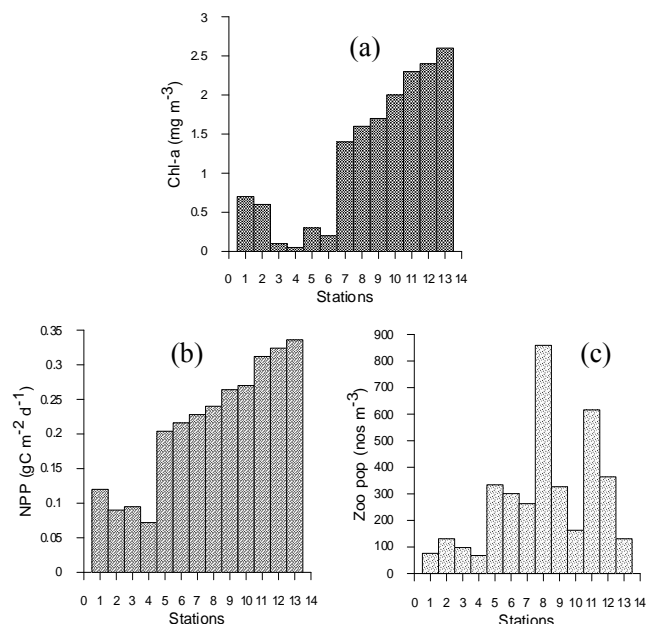


Figure 3. Spatial variation of (a) pigment concentration (b) Primary productivity (c) zooplankton abundance



Figure 4. Photomicrographs of some selected zooplankton recorded during the present study

Copepods, fish eggs, Cladocera, Appendicularians, Decapods, Ostracods, Doliolids and Amphipods were the major zooplankton groups observed along the shelf waters (Figure 4). Overall percentage compositions of zooplankton recorded at different stations are shown in Figure 5. Zooplankton community of the southwest coast shared heterogeneous assemblage of many species, covering many taxonomic groups. Population dynamics is related to the physico-chemical factors.

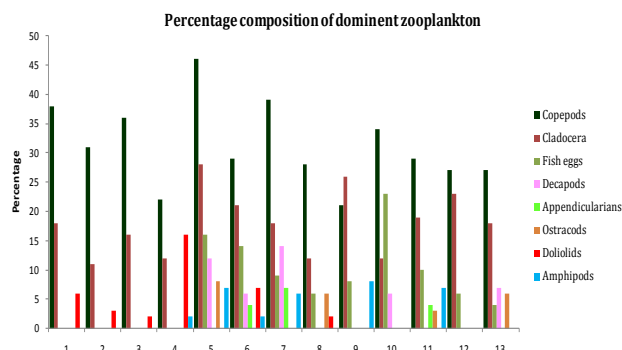


Figure 5. Percentage composition of dominant groups of zooplankton in different stations

The 3D surface plot of zooplankton population against chlorophylla and primary productivity, well demonstrated in Figure 6, indicates that the zooplankton population is not in direct tune with the high chlorophyll concentration and primary productivity. But it could be attributed to salinity variations and marginal stress from the fresh water input. Gross pollution problems at Veli transact owing to the release of untreated effluents from industries and domestic sectors which could be the reason for comparatively low levels of pigment concentration and primary productivity and zooplankton population at Cochin. From the results, it is quite clear that the surface productivity of Veli was drastically decreased especially near the shore, due to higher suspended matter and decreased transparency, apart from the acidic pH caused for factory effluents, which adversely affect the phytoplankton community, thereby corresponding

organic production. This conclusion is in agreement with Bijumon et al.[42]. However, a trend in increased primary productivity positively correlated with pigment concentration was observed along the northern transects.

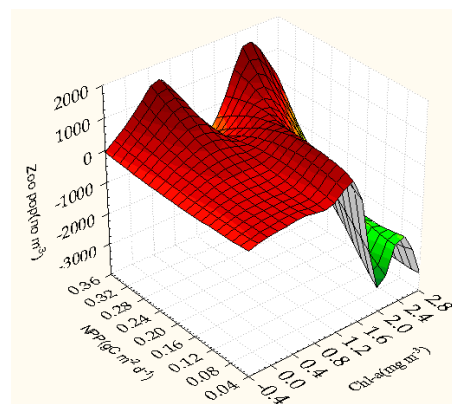


Figure 6. 3D Surface plot of zooplankton population (no m^{-3}) against chlorophylla ($mg\ m^{-3}$) and NPP ($g\ C\ m^{-3}\ d^{-1}$) zoo population (no m^{-3}) = Distance weighted least squares

Photoprotective compounds traced from the zooplankton from the shelf waters of southwest coast of India are represented in Table 1 and Figure 7. Mycosporine-glycine was found in zooplankton of all the polluted stations in Veli, Neendakara, and Cochin, probably for the protection from the adverse effect of pollution *viz.* high discharge of acidic effluents at Veli transact, sewage discharge at Neendakara and dredging activities and sewage disposal at Cochin. Among the photoprotective compounds, Mycosporine-glycine, exhibit a high antioxidant activity scavenging superoxide anions and inhibiting lipid peroxidation resulting from UV-induced production of ROS besides UV-absorption[43]. This is a self-protective mechanism intimately involved in the prevention of cell damage from the adverse impact. Thus the presence of Mycosporine-glycine in zooplankton of the polluted area is understandable as it protects the organisms from the adverse effects of pollution in addition to photoprotective action by antioxidant metabolites.

Table 1. Photoprotective compounds traced from the zooplankton from the southwest coast of India

Sl. No.	St.code	Lat.	Long.	MG	PR	PT	SH	UC
1	Veli-A	8°29.000	76°49.400	-	+	+	-	+
2	Veli-B	8°30.978	76°52.022	+	+	+	-	-
3	Veli-C	8°30.000	76°49.630	-	+	+	-	-
4	TTP	8°32.566	76°54.014	+	+	+	-	-
5	Neendakara	8°56.183	76°29.795	+	+	+	-	-
6	Cochin	9°56.793	76°11.763	+	+	+	-	-
7	Kodungallur	10°11.546	75°53.365	-	+	+	-	-
8	Calicut	11°13.569	75°44.220	-	+	+	-	-
9	Mangalore	12°57.500	74°47.001	-	+	+	-	-
10	Chitrapura	12°57.500	74°45.200	-	+	+	-	-
11	Honovar-A	14°17.400	74°22.809	-	+	+	-	+
12	Honovar-B	14°17.400	74°22.700	-	+	+	-	+
13	Karwar	14°49.800	75°50.000	-	+	+	-	-

MG-Mycosporine glycine, PR – Porphyra, PT – Palythine, SH – Shinorine, UC – Unknown compounds.

4. Conclusions

In the south west coast of India, presence and characterization of the sun screening compounds (MAAs) in the zooplankton community has been reported for the first time. Palythine and porphyra have been detected at all the stations studied. However, at four stations, mycosporine-glycine was present, where high pollution loads were there. Presence of this particular UV-absorbing compound, suggests that it is in response to pollution related environmental stress existing in these areas. High pollution loads in the stations have been reflected through the decreased levels of chlorophylla and of primary productivity. In addition to palythine, porphyra and mycosporine-glycine and some other unknown compounds were also present at Veli and Honavar. Quantification of MAAs of different zooplankton species can be performed to figure out their commercial application.

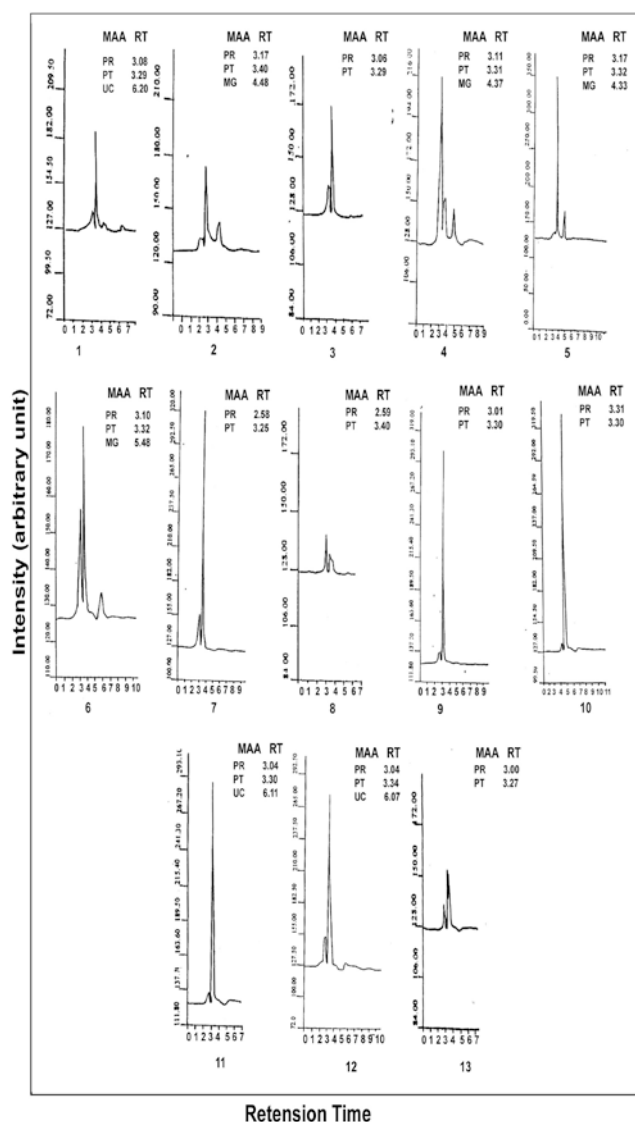


Figure 7. HPLC of mycosporine - like amino acids from zooplankton (stations 1 to 13) Reverse-Phase C₁₈ Column; mobile phase 0.2 % acetic acid; flow rate 1.0 ml min⁻¹ chart speed 0.50 cm/min. Measurements of absorbance at 330 nm

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