

Isolation, Identification and Susceptibility Patterns of *Moraxella Catarrhalis* among Children with Otitis Media in Khartoum State

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Abstract Otitis media is the most frequent infection associated with *Moraxella catarrhalis* in children and comprises inflammation of the middle ear accompanied by a liquid effusion. The present study was aimed to determine the frequency and susceptibility patterns of *M. catarrhalis* in children with otitis media in Khartoum State, during the period from January to March 2011. A total of 110 specimens of middle ear discharge were collected by sterile swabs from Khartoum National Centre of Ear, Nose and Throat (ENT), Head and Neck Surgery. Specimens were cultured on chocolate agar, sheep blood agar and MacConkey agar. Ninety (82%) specimen showed significant growth while twenty (18%) did not show any growth. The identification of *M. catarrhalis* was determined based on colonial morphology, Gram stain and a number of biochemical tests. The susceptibility test was carried out using Kirby-Bauer disc diffusion method. Three (3.3%) isolates were positive for oxidase, catalase, DNase, tributyrin tests and reduced nitrate to nitrite and therefore, they were identified as *M. catarrhalis*. The clinical isolates *M. catarrhalis* were found to be sensitive to amoxiclav, azithromycin, ceftazidime, ceftriaxone, cephalexin, cefotaxime, chloramphenicol, ciprofloxacin, cotrimoxazole and erythromycin and resistance to ampicillin which might be due to the production of β -lactamase enzyme.

Keywords *Moraxella catarrhalis*, Children, Susceptibility Patterns

1. Introduction

Otitis media is inflammation of the middle ear, most commonly caused by the buildup of fluid behind the ear drum, as a result of a blockage to the Eustachian tube. It is more common in children, as their Eustachian tube is shorter and more horizontal than adults and is made up of more flaccid cartilage, which can impair its opening. Otitis media can cause a mild to moderate hearing loss, due to the fluid interfering with the transmission of sound through to the inner ear. It can often affect the tympanic membrane causing it to retract or become inflamed. The fluid can cause the tympanic membrane to bulge and become inflamed and occasionally the tympanic membrane will perforate[1].

Otitis media is the most frequent infection associated with *Moraxella catarrhalis* in children and comprises inflammation of the middle ear accompanied by a liquid effusion. Approximately 50% of children will have experienced at least one episode of acute otitis media

(AOM) by their first birthday, this proportion rising to 70% by 3 years of age, representing a tremendous disease burden for this age group and necessitating the widespread use of antibiotics[2].

Moraxella catarrhalis is a Gram-negative non-encapsulated diplococcal bacterium belonging to the family *Moraxellaceae*. The genus *Moraxella* actually comprises both coccoid and rod-shaped bacteria, and the classification of *M. catarrhalis* has been rather complex, the bacterium being alternatively named *Branhamella catarrhalis*, *Moraxella (Branhamella) catarrhalis*, and now the preferred *Moraxella catarrhalis*[3, 4]. For the most of 20th century, *M. catarrhalis* was considered a saprophyte of the upper respiratory tract associated with no significant pathogenic consequences[5]. Currently it has been proven to be a pathogen in its own right with global isolates originating from two major increased potential to bind to human epithelial cells and an older lineage. Person-to-person spread through inhalation is not considered to be the mode of transmission, while hand-to-hand nosocomial spread may be common in certain settings[6].

M. catarrhalis has been regarded as the third most important bacterial agent of acute otitis media in children, after *S. pneumoniae* and *H. influenzae*[7, 8].

Several reports showed an increased proportion of *M.*

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Catarrhalis isolation from the middle ear fluid (MEF) in acute otitis media (AOM). Kilpi *et al.*, [9], have reported an increase from 10% to 23% within 15 years, and a similar pattern has also been reported in the United States. In Costa Rica, the prevalence of *M. Catarrhalis* isolated from the MEF of children with AOM aged 3–144 months increased from 2.5% of all pathogens during 1992–1997 to 7% during 1999–2004 and was most commonly found in children aged less than 24 months during the dry season [6].

The financial impact on global health care systems of the high incidence of *M. catarrhalis* colonization and disease is significant, and consequently, several research groups are currently involved in identifying and assessing the usefulness of putative *M. catarrhalis* vaccine candidates [10]. However, *Moraxellacatarrhalis* is estimated to be responsible for 3–4 million cases of otitis media annually, with an associated health care cost (direct and indirect) of \$2 billion each year [11].

Bacterial resistance to antimicrobial agents has become an increasing problem in the treatment of otitis media. A multicenter surveillance study, carried out in Asia and Europe, demonstrated a high prevalence of antimicrobial resistance among respiratory pathogens and important differences in antimicrobial resistance profiles between countries. Pathogens that cause acute otitis media become resistant to commonly used antibiotics. The increasing rates of antibiotic resistance are due to repeated exposure of these bacteria to antibiotics and geographic spread of resistant strains. The rapid emergence of multidrug resistant otitis media in developing countries is a new potential threat to the survival of newborn babies and children [12].

Unlikely our African countries lack the authentic data documentation system so it is very difficult to trace such information. According to the best of our knowledge this is first report of *M. catarrhalis* as pathogenic bacterium infected middle ear in the Sudan. The present study was a cross sectional study conducted to determine the frequency and susceptibility of *M. catarrhalis* in children (under 15 years) with otitis media during the period from January to March 2011 in Khartoum National Centre of ENT, Head and Neck Surgery.

2. Materials and Methods

2.1. Sample Collection and Isolation

Samples were collected from 110 children who their parent agreed to participate in this study. Swabs samples were collected from the middle ear of infected children using sterile cotton wool swabs and transported in Amies Transport Media to the laboratory for diagnosis in the same day of collection. The specimens were inoculated under aseptic condition into MacConkey agar, sheep blood agar, chocolate agar and nutrient agar, and incubated at 37°C for 48 hours at presence of 10% CO₂.

2.2. Identification

The identification of *M. catarrhalis* was based primary on colonial morphology, Gram stain and a number of biochemical tests namely; Oxidase, Catalase, Nitrate Reduction and DNase. *Moraxellacatarrhalis* was differentiated from *Neisseria* using DNase, tributyrin test and growth on nutrient agar at 35°C [13, 14, 15]. Several others biochemical tests were used to identify the causative agents of OTM other than *M. catarrhalis* (data not shown). Three control strains of *M. catarrhalis* ATCC2 5240, 25238 and 23246 from the American Type Culture Collection (ATCC) were used to confirm the results obtained by clinical isolates.

2.3. Antibiotic Susceptibility Test

The susceptibility test was done by Kirby-Bauer disc diffusion commonly used method. The following eleven antimicrobial impregnated disks were used: amoxiclav, ampicillin, azithromycin, ceftazidime, ceftriaxone, cephalexin, cefotaxime, chloramphenicol, ciprofloxacin, co-trimoxazole and erythromycin.

3. Results

Out of 110 specimens, 90 (82%) showed significant growth whereas 20 (18%) sample displayed no or insignificant growth. The colonial morphology was used as primary identification tool that differentiates *M. catarrhalis* from other microorganisms. *M. catarrhalis* was grown well on sheep blood agar and chocolate agar but not on MacConkey agar. On sheep blood agar colonies were gray to white, opaque, smooth, dry and 1–3 mm in diameter after 24 hours of incubation. With an inoculating loop, colonies could easily be slid across the agar surface, like hockey pucks, and could be stacked like disks. Colonies on chocolate agar were pinkish-brown.

The isolates which later identified as *M. catarrhalis* were showed large kidney shaped Gram negative cocci. Three (3.3%) isolates were positive for oxidase, catalase, DNase, tributary tests and reduce nitrate to nitrite, thus these strains were identified as *M. catarrhalis*, they were isolated from children aged 1 – 5 years, 1 (30.3%) from female and 2 (60.6%) from male (Table 1).

Table 1. Distribution of *M. catarrhalis* isolates according to sex and age groups

factor		Frequency	percentage
Sex	Male	2	30.3
	Female	1	60.6
Age group	1-5 year	3	100
	6-10 year	Zero	Zero
	11-15 year	Zero	Zero

The results of identification tests of clinical isolates were compared with ATCC control strains that gave the same results.

Table 2. Antibiotic susceptibility patterns of *M.catarrhalis* ATCC and Clinical isolates

Antimicrobial agent	American Type Culture Collection ATCC				Clinical isolates			
	Sensitive		Resistant		Sensitive		resistant	
	Frequency	percentage	Frequency	percentage	Frequency	percentage	Frequency	percentage
Amoxiclav	3	100	zero	zero	3	100	zero	zero
Ampicillin	3	100	Zero	Zero	zero	zero	3	100
azithromycin	3	100	Zero	Zero	3	100	Zero	Zero
ceftazidime	3	100	zero	zero	3	100	zero	zero
ceftriaxone	3	100	Zero	Zero	3	100	Zero	Zero
Cephalexin	3	100	Zero	Zero	3	100	Zero	Zero
cefotaxime	3	100	Zero	Zero	3	100	Zero	Zero
chloramphenicol	3	100	Zero	Zero	3	100	Zero	Zero
ciprofloxacin	3	100	Zero	Zero	3	100	Zero	Zero
co-trimoxazole	3	100	Zero	Zero	3	100	Zero	Zero
erythromycin	3	100	zero	zero	3	100	zero	zero

On the other hand, antibiotic susceptibility profile showed that all ATCC control strains were sensitive to all antimicrobial agents. Clinical isolates were also sensitive to all except ampicillin (Table 2).

4. Discussion

In this study, 3 (3.3%) of isolates were identified as *M.catarrhalis* based on colony morphology, Gram-stain and biochemical reactions. However, Ellis[16] reported that the phenomenon of colonies characters in conjunction with oxidase and catalase positivity and typical Gram-stain is the most settings appropriate for presumptive identification of *M.catarrhalis*. Definitive identification should be differentiated it from the *Neisseria*. However, in the present study, *M.catarrhalis* was differentiating from *Neisseria* by its ability to grow into nutrient agar at 35°C, hydrolysis of tributyrin and DNase. These results were almost similar to that reported by Broides *et al.*, [17] who found that *M.catarrhalis* occurred in proportion of 4.8% in children less than 5 years and Vergison[18] who showed that *M.catarrhalis* proportion in children with otitis media was 3–20%. In this study *M.catarrhalis* was occurred most frequently in male than female and this was in alignment with previous report which suggested that the males are more liable to be infected by *M.catarrhalis* [11].

All *M.catarrhalis* isolates in the present study were sensitive to amoxiclav, azithromycin, ceftazidime, ceftriaxone, cephalexin, cefotaxime, chloramphenicol, ciprofloxacin, co-trimoxazole and erythromycin and showed resistant to ampicillin which might be due to its ability to production of β -lactamase enzyme. According to McGregor *et al.*, [19] most *M.catarrhalis* isolates produce β -lactamases and resistant to penicillins; and susceptible to most other antibiotics, including erythromycin, tetracycline, co-trimoxazole, and the combination of penicillins with a β -lactamase inhibitor (e.g., clavulanic acid). However, it has been suggested that the production of β -lactamases by *M.catarrhalis* could protect colonizing pathogens from the

effects of β -lactam antibiotic treatment [10]. More than 90% of *M. catarrhalis* strains have been shown to resist amoxicillin, and these rates vary by region. Amoxicillin-clavulanate, second- and third-generation oral cephalosporins, and trimethoprim-sulfamethoxazole (TMP-SMX) are the most recommended agents. Alternatively, azithromycin, clarithromycin, or dirithromycin can be used [11].

In any case, the financial impact on global health care systems of the high incidence of *M. catarrhalis* colonization and disease is significant, and consequently, several research groups are currently involved in identifying and assessing the usefulness of putative *M. catarrhalis* vaccine candidates [10].

REFERENCES

- [1] Alsaimary, E. I., Alabbasi, M. A., and Najim, M. J. 2010. Antibiotics susceptibility of bacterial pathogens associated with otitis media. *J. Bacteriol. Res.* 2(4): 41-50.
- [2] Falkow, S., Rosenberg, E., Schleifer, K.-H. 2006. Stackebrandt, E. *Proteobacteria: Gamma Subclass*, Volume 6: *The Prokaryotes*, 3rd ed., p: 965, Springer, USA,
- [3] Karalus, R., Campagnari, A. 2000. *Moraxella catarrhalis*: a review of an important human mucosal pathogen. *Microbes. Infect.* 2(5):547-559
- [4] Murphy F, T and Parameswaran, I.G. 2009. *Moraxella catarrhalis*, a Human Respiratory Tract Pathogen. *Clin. Pract.* 49: 124-131.
- [5] *Moraxella catarrhalis* Infection. Online Available: <http://emedicine.medscape.com/article/222320-overview>. Accessed on 23.11.2012
- [6] Guevara, S., Soley, C., Arguedas, A., Porat, N., Dagan, R. 2008. Seasonal distribution of otitis media pathogens among Costa Rican children. *Pediatr. Infect. Dis. J.* 27(1):12-16
- [7] Joseph O A and Odeh N E. 2011. Prevalence, Haemolytic Activities and Fluoroquinolones Susceptibility Profiles of

- Moraxella catarrhalis*, *Streptococcus pneumoniae* and *Haemophilus influenzae* Associated with Acute Otitis Media. *Nat. Sci.* (6):85-92.
- [8] Hays, J. P. 2009. *Moraxella catarrhalis*: A mini review. *J. Pediatr. Infect. Dis.* 4 (3): 211-220.
- [9] Kilpi, T., Herva, E., Kajjalainen, T., Syrjänen, R., Takala, A.K. 2001. Bacteriology of acute otitis media in a cohort of Finnish children followed for the first two years of life. *Pediatr. Infect. Dis. J.* 20(7):654-62.
- [10] de Vries, W, P, S., Bootsma, J, H., Hays, P, J and Hermans, M, W, P. 2009. Molecular aspects of *Moraxella catarrhalis* Pathogenesis. *Microbol. Mol. Biol. Rev.* 73(3) 389–406
- [11] The health science-*Moraxella Catarrhalis* Infections. Online Available <http://thehealthscience.com/showthread.php?168061-Moraxella-Catarrhalis-Infections>. Accessed on 23.11.2012
- [12] Al-Shara M. 2012. A Five-year Review on the Etiology and Antimicrobial Susceptibility Pattern of Otitis Media Pathogens in Jordanian Children. *O. M. J.* 27(5): 358-363
- [13] Verduin, C.M., Hol, C., Fleer, A., van Dijk, H., van Belkum, A. 2002. *Moraxella catarrhalis*: from Emerging to established pathogen. *Clin. Microbiol. Rev.* 15(1): 125–144.
- [14] Ahmed, F., Young, H., Mcleod, T, D., Crouhghan JM and Calder, A, M. 1987. Characterisation of *Branhamella catarrhalis* and differentiation from *Neisseria* species in a diagnostic laboratory. *J. Clin. Pathol.* 40:1369-1373
- [15] Doern, V. G and Morse A, S. 1980. *Branhamella* (*Neisseria*) *catarrhalis*: Criteria for Laboratory Identification *J. Clin. Microbiol.* 11(2):193-195.
- [16] Ellis, E, M. 1998. *Infectious Diseases of the Respiratory Tract*, 1st ed. 120-123. Cambridge University press. England.
- [17] Broides, A., Dagan, R., Greenberg, D., Givon-Lavi, N., Leibovitz, E. 2009. Acute otitis media caused by *Moraxella catarrhalis*: epidemiologic and clinical characteristics. *Clin. Infect. Dis.* 49(11):1641-7
- [18] Vergison, A. 2008. Microbiology of otitis media: A moving target. *Vaccine*; 26:5-10
- [19] McGregor, K., Chang, B. J., Mee, B. J., Riley TV. 1998. *Moraxella catarrhalis*: clinical significance, antimicrobial susceptibility and BRO beta-lactamases. *Eur. J. Clin. Microbiol. Infect. Dis.* 17(4):219-34.