

Periodontium and Orthodontic Implications: Biological Basics

Marinho Del Santo

Orthodontist in Private Practice, Neo Face Dental Clinic

Abstract As more and better orthodontic and periodontal scientific evidences are contextualized, potentially richer will be their achieved clinical results. The primary goal of this article is to study the main anatomic, histologic, physiologic and pathologic features of the periodontal tissues, and such knowledge to be applied in the Orthodontics and Periodontics fields.

Keywords Periodontics, Orthodontics, Periodontal Development, Periodontal Anatomy, Periodontal Physiology

1. Introduction

In general, the periodontal tissues of patients who seek periodontal treatment suffer constant and dynamic oscillation between the physiologic and pathologic states, and the natural ongoing process to achieve periodontal homeostasis depends upon tissue reposition, remodeling, degradation and repair. In practice, as more and better orthodontic and periodontal treatments are contextualized, more potentially richer will be the achieved clinical results.

In Orthodontics, dental movements target the correction of malocclusions. Such goal includes the leveling and alignment of the teeth in the arches, and such movements occur throughout dynamic processes of bone resorption and bone deposition. The correct dental positioning allows a better occlusal load distribution and facilitates the routinely oral hygiene of the patient. Therefore, orthodontic results increase the masticatory action, eliminate occlusal interferences and increase the probability of maintenance of periodontal health of the treated patient. Since Orthodontics allows that the protective periodontium (gingival tissues) fits better around better positioned teeth, it facilitates hygiene care and contributes for the esthetical aspects of smiling. However, when biological limits are not respected by the applied orthodontic mechanics, partial relapse of the achieved results and alveolar and root resorption may occur.

In Periodontics, the clinical or surgical therapy promotes a healthy periodontal status and targets to carry on such condition. However, the physiological periodontal homeostasis may not be maintained because diverse temporal and individual factors as oral hygiene, microorganisms virulence, integrity of defence barriers of

the tissues and host resistance. When the equilibrium is broken the periodontal disease is installed or re-installed, and diverse degenerative levels can possibly happen, and are revertible or not.

2. Goal

The main goal of this article is to study the periodontal tissues, illustrating their innate features, and highlighting pertinent clinical parameters. The target-specialists of these publications practice in the Periodontics and Orthodontics fields, and must know the mandatory periodontal requisites to recommend orthodontic treatment, and the benefits of the orthodontic treatment for the periodontal homeostasis.

3. Panoramic View: Periodontal Protection and Function

Independently of clinical and experimental advances, the most basic and undeniable biological principle in Periodontics is: the periodontium is made by protective and supportive tissues which allow the teeth to be present in the oral cavity and perform its main function mastication, indeed the discontinuity of the oral mucosa do not make it vulnerable to alien antigens.

The human body (and also of other mammals) is protected against alien microorganisms by the continuity of its tissues, fundamentally the skin and the nasopharyngeal and gastrointestinal mucosa. With dental eruption, the continuity of the oral mucosa is broken and in order to maintain the protection of the individual, a sophisticated system of biological sealing grows and develops around the teeth. Such system is the protective periodontium, mainly the junctional epithelium. However, other factors also play important role for the scenario where periodontal homeostasis is achieved and maintained, as the correct and complete oral hygiene performed by the patient.

* Corresponding author:

marinho@delsanto.com.br (Marinho Del Santo)

Published online at <http://journal.sapub.org/ijsr>

Copyright © 2012 Scientific & Academic Publishing. All Rights Reserved

Discipline and motor-skill are fundamental for the patient to achieve and maintain such goal, mainly targeting the equilibrium between the periodontal microbiota and its host, "protecting the protection". The junctional epithelium must be protected of suffering more intense aggressions that is naturally prepared to do so. Professionals deliver such equilibrium status to the patient and orient him/her how to maintain a satisfactory level of microorganism colonization.

In our opinion, to discriminate periodontal tissues as protective or supportive,¹¹⁰ although histologically correct, is unnecessary. The periodontium, under a broader view, is a unique and highly sophisticated system. The gingiva is prepared to receive the masticatory load and establish a protector collar, fibrous and well attached, around the teeth. Its junctional epithelium represents the biological sealing of the "internal periodontium environment" against external aliens. Its periodontal ligament promotes the dynamic mechanical-functional relationship between root cementum and its respective laminate alveolar bone.

Therefore, the periodontal structures shall be studied and understood as specialized epithelial and connective tissues that, up to certain limits, are phylogenetically adapted and epigenetically lapidated according to the imposed functional demand.

4. Embryological Origin and Development of the Periodontium

As a rule, the periodontium is a tissue of ectomesenchymal origin. However, its gingival epithelium (including the junctional epithelium) and the rest of Malassez cells have origin exclusively ectodermic. The gingival epithelium plays important ontogenetic modulation in the ectomesenchymal tissues,^{47,111,112,114} although such inductive mechanisms are not enough clear.¹²⁸ The enamel organ forms enamel, the dental papilla forms the dental-pulp complex and the dental follicle forms the cementum, the periodontal ligament and partially the alveolar bone.

As examples of the high complexity specialization of the periodontal tissues, we highlight the junctional epithelium and the periodontal ligament. The junctional epithelium represents a protective tool. Without it there would be no periodontal sealing and the external and internal environments were not isolated one from each other. The periodontal ligament is a complex apparatus for mechanic-functional support of the teeth, allowing their relationship with the alveolar bone.

5. Anatomy-Histology-Physiology of the Periodontium

The oral mucosa is continuous with the labial vermillion and with the soft palatal and pharyngeal mucosa. It is divided in: 1) masticatory mucosa, keratinized, which represents 25% of the internal oral surface, it is similar to the skin epithelium and it is present in the gingiva and the hard palate;

2) specialized mucosa, which covers the tongue dorsum and; 3) oral mucosa, non-keratinized, which represents 60% of the internal oral surface, it is similar to the esophageal and vaginal mucosa, it is continuous to the masticatory mucosa and it is present in the rest of the mouth.

As part of the masticatory mucosa, the gingiva is a fibrous collar tissue around the teeth, resistant, continuous and well attached, inserted in the teeth and in the crest alveolar bone. The gingiva overlaps approximately 2.0 mm the enamel-cementum junction, being that its sulcus measures approximately 0.5 mm¹²⁷ and the junctional epithelium measures approximately 1.5 mm. With the installation of the gingivitis process, the deepness of the gingival sulcus may increase, not because of some apically migration of the junctional epithelium, but because inflammation swallows the gingiva. With the possible progression of gingivitis the junctional epithelium breakage occurs and, consequently, a direct communication between the external and internal environments. An evident sign of such continuity is gingival bleeding. epithelium per se can not bleed since has no blood supply.

On the teeth, below the junctional epithelium, there is approximately 1.0 mm of fibrous gingival attachment: dental-gingival and dental-periodontal fibers. Therefore, the total space between the gingival free margin and the crest of the alveolar bone is approximately 3.0 mm. It is called biological space. The gingival collar is also formed by alveolar-gingival, circumferential and semi-circumferential, trans-gingival, inter-gingival and trans-septa fibers, composing a resistant soft tissue frame which contours all the teeth. Such fibrous bundles are mainly made by type I collagen (approximately 91% of incidence; it is the dense collagen) and type III collagen (approximately 8% of incidence; it is the soft collagen).^{69,70,22} Such net of collagen fibers, with longer turnover than skin fibers,⁹⁹ besides providing consistency and resistance to the gingiva, promotes its attachment to teeth and to the alveolar bone, making a supportive apparatus for the masticatory function. Moreover, such twisted fibrous system maintains the adjacent teeth inter-connected in the dental arch. The referral of pre-orthodontics dento-gingival fibrotomy targets to minimize the resistance to the orthodontic movement and also to decrease the risk of relapse, in special when dental rotation is the case.⁵⁰

Anatomically, the gingival tissue is adapted to the shape, size and positioning of the teeth. The gingival shape is of continuous segmented arches separated by inter-dental papillae. In the inter-proximal regions the gingiva is narrow between the anterior teeth and large between premolars and molars. In young patients, the inter-proximal gingiva totally fills the space between the teeth, in a region called gingival col.

Attached gingiva is the band of gingival tissue present between the free gingiva and the mucosa-gingival junction. The attached gingiva is fixed, pink (or pigmented in afro-descendent individuals), more or less thick and more or less large. Its thickness ranges approximately from 0.5 mm

to 2.5 mm and its width ranges approximately from between 3.0 to 6.0 mm, being thickest in the vestibular face of the upper incisors and in the lingual face of the lower incisors.³² The thickness of the attached gingiva also varies according to the degree of dental eruption.^{63,64,88} In the other hand, the oral mucosa is mobile, thin and relatively elastic, being highly blood supplied (with reddish appearance).

The oral epithelium is a stratified paved keratinized epithelium, presenting four cellular layers: basal, spinosum, granulosum and keratinized (differentiated or not). The cells of the granulosum and keratinized layers exfoliate on the tissue surface with a turnover of approximately 14 days. The keratinocytes are cells that produce keratin (and make the keratinized layer) and correspond to 90% of the total cellular tissue population. The keratinized layer is the principal barrier against external aggressions. Besides keratinocytes, melanocytes, Langerhans cells, Merkel cells and inflammatory cells are also found. The oral epithelium does not present inter-cellular fibrous components but proteoglycans,^{121,7} adipose tissue and water. The proteins lectin, hyaluronan, chondroitin, dermatan, decorin and syndecan,^{79,98,103,52,82} among others, are in tight contact with the adjacent connective tissue.⁷⁵ The gingival epithelium is made by the oral epithelium, sulcus epithelium and junctional epithelium.

6. Junctional Epithelium

With dental eruption, the fusion of the enamel reduced organ epithelium with the oral epithelium occurs. The dental-gingival junction is established as an epithelium collar, derived from the two epithelia, which united to make the junctional epithelium.⁹³ The junctional epithelium and the sulcular epithelium recover the gingival sulcus, where protection is the main goal. The junctional epithelium is a simple epithelium, constituted by only two layers (*strata*) of stratified squamous epithelium, with cuboid basal cells and flat supra-basal cells, parallel oriented to the dental surface. It is not keratinized and presents few desmosomes for inter-cellular union,^{90,35} with inter-cellular spaces that shelter a great quantity of neutrophils, macrophages, mastocytes and lymphocytes, present with or without inflammation.⁹¹ The sulcular fluid, secreted even without any detected pathological process,²⁰ contributes to the mechanical cleanness of the gingival sulcus and it may be accumulated in its bottom.

The junctional epithelium presents an exceptionally high turnover, being completely substituted in approximately 05 days in humans.¹⁹ Such replacement is promoted by constant mitosis in its basal layer and consequent exfoliation of supra-basal cells into the gingival sulcus. The basal layer of the junctional epithelium can regenerate from the basal cells of the oral epithelium and also, potentially, regenerates from the skin.^{55,19} Such regeneration is also possible around dental implants.^{33,65,34} As the junctional epithelium presents a significant amount of polymorphic leucocytes, it might be

compared to the epithelium of lymphoid tonsils^{92,94,80} and it is considered a very favorable environment for immunological reactions, innate or acquired.⁸⁰

If the inflammatory process is installed, polymorphic leucocytes are attracted by microbiological chemotaxis and abundantly released in the gingival sulcus.^{105,2} As important the inflammatory process is, as greater neutrophil migration occurs,^{3,48} providing the first innate immunological combat against external aggressors. Macrophages and lymphocytes are also found; however, they are less than 5% of the polymorphic leucocytes.² Lymphocytes leads acquired immunological reactions.

Clinically, adequate oral hygiene allows reduction or elimination of bacterial aggressive agents in the gingival sulcus. In an adequate ecosystem, possible repair and reinstallation of a junctional epithelium in a more coronal position, in natural teeth and in dental implants, is the best periodontal protection. The epithelium is called of long junctional epithelium. Even assuming that the losses in the alveolar bone crests are irreversible, the possibility of establishment of a long junctional epithelium significantly improves the prognosis of an adverse periodontal scenario.

7. Dentin-cementum Junction

Genetic expression for the development of the dental organ occurs in the dental papilla and in the dental follicle.¹¹² The Hertwig's epithelial root sheath is classically described as the precursor for the apical development of the dental organ, in its dentinoblastic and cementoblastic differentiation.^{116,115,60,61} The hyaline layer of Hopewell-Smith, originally described by Hopewell-Smith,³⁸ can be a variation of the dentin which recovers the dental root surfaces. It was described it as non-cellular cementum.^{49,77} It has been confirmed^{73,74,14-18} the possible existence of such non-mineralized layer between the dentin the Hertwig root sheath, continuous with the dentin and therefore called pre-dentin; however, its existence is only sporadic.¹⁰⁹ In theory, the function of the Hopewell-Smith layer would be to promote non-cellular cementum deposition over root dentin.¹¹²

8. Cementum

Cementum is a mineralized connective tissue, specialized in the recovering of the dental roots and where the collagen fibers of the periodontal ligament are anchored to promote dental sustain.⁷⁷ Cementum presents peculiar biological features.

The cementoblasts, cells that produce cementum, are originated in the dental follicle.^{106,114} The interactive relationship between dentinoprogenitor cells and cementum progenitor cells¹⁶ indicated that there is no gap between dentin and cementum in humans.¹⁷ However, the origin of cementum progenitor cells is controversial. Although in

theory they are originated from the ectomesenchyme, the hypothesis of that they are originated in the Hertwig's sheath has not been refused.^{116,61} There are different structural types of cementum, according to its location, function, chemical composition and mineralization degree.

Basically, human teeth present three types of cementum: 1) the non-cellular and non-fibrous type, which recovers small areas of the enamel, near to the enamel-cementum junction; 2) the non-cellular fibrous type, which is present in the two coronal thirds of the dental root and; 3) the cellular fibrous type, which is present in the apical third of the dental root. However, in the anterior teeth the non-cellular fibrous type also covers part of the apical third of the root, and such coverage decreases in the posterior teeth, where more cellular fibrous type cementum is identified. Cellular cementum is present where the non-cellular cementum is not present, as furcations and apices. In general, cementum is thicker as apical as it is located, and it can invade the apical foramen.

The non-cellular cementum is more mineralized than the cellular cementum, possibly because its process of formation is slower. Then, it allows longer contact of the minerals, that come from blood vessels, with the organic matrix. The apical cellular cementum holds the capacity of quicker growth when compared to the non-cellular cementum. It also presents more intense remodeling, in response to the masticatory functional stimuli from dental occlusion. It seems that such functional stimuli reduce the mineral deposition rate in the apical area and do not increase it. The cellular cementum can overlap layers of non-cellular cementum and vice-versa.

The composition of the cementum matrix is similar to the bone matrix and to the dentin matrix, being 50% of it made of inorganic components, mainly hydroxyapatite crystals. Apparently, the cementum mineralization process is identical to the bone mineralization. Its framework is made by a collagen matrix, where mineral deposition occurs. Such organic matrix is mainly made by type I (90%) and type III collagen (5%)^{11,18,122} and by associated adhesion proteins as chondroitin, 4,6 sialoprotein, tenascin and osteonectin.⁶¹

It has been suggested that type I fibers are recovered by type III fibers,¹²⁰ avoiding its mineralization. However, other authors hold different opinion, stating that type I and type III collagen are parallel to each other, and there is no overlap.^{42,9,40} Interesting to note that although the new layers of cementum are added during the lifetime of the individual, its mineral content does not significantly change with aging,^{95,68} differently of what happens with dentin, which mineral content significantly does, provoking obliteration of the dentinary channels.

Bone participates of the systemic metabolism, as a reserve of Calcium and other minerals; however, cementum is excluded of such dynamics. There is no evidence that changes in the Calcium systemic level are associated with biochemical changes in the cementum. It is possible that Fluoride ions brought by blood vessels of the periodontal ligament plays a role. It intensively reacts with the

hydroxyapatite of the cementum surface, especially in the dental cervical region, and it armors cementum against Calcium systemic biochemical changes.

Although do not suffer continuous remodeling, cementum slightly increases its thickness during lifetime, if no periodontal disease is present. Intense Fluoride deposition on cementum surface is probably the main reason for the fact that caries and root resorption are presented in a "mined form", that means, they progress internally, before significant major changes on the cementum surface.

Usually cementum is classified of a "bone-like" tissue; however, the comparison is limited. The differences between both tissues are evident when bone and cementum biodynamics are studied during the orthodontic movement.

Quantitatively, cementum responds to tension orthodontic forces as alveolar bone; however, in a much slower speed. In the side submitted to orthodontic tension, slight cementum deposition may occur in layers.⁸¹ Under physiologic orthodontic conditions, in the side submitted to orthodontic pressure, alveolar bone suffers resorption and cementum does not, or if does, it is not evident by routine diagnostic methods. This difference between both tissues is essentially the speed of the metabolism: the bone metabolism is much faster and dynamical than the cementum metabolism.

And there are histological reasons for such difference. The main reason is blood supply, absent in non-cellular cementum and minimal in cellular cementum; however, abundant in alveolar bone. As a consequence of such minor blood support, with sparse and limited channel communications, the cementum turnover is much longer than the bone one. Besides that, although the collagen fibers of cementum and alveolar bone are both exposed to the intense metabolic activities of the periodontal ligament, the fibers that are inserted in the cementum present a turnover significantly longer than the ones of the alveolar bone. In the cementum, the attached fibers are more numerous, more stable and more mature. In the alveolar bone, the fibers (Sharpey's fibers) are unstable and suffer relatively faster remodeling.⁵⁴

In adverse orthodontic conditions, as excessive orthodontic load combined or not with extensive orthodontic treatment, cementum resorption may also occur. In such conditions, bone resorption is also altered: it is delayed and it occurs "internally" and not "superficially", due to the obliteration of the blood vessels of the periodontal ligament, with consequent necrosis and tissue hyalinization. As a direct clinical consequence, easily assessed, the orthodontic movement is delayed. Up to the timing that necrosis and tissue hyalinization are solved by the installed inflammatory process, possible cementum resorption may happen and may also be repaired. Most of the time, it is not even diagnosed. When root resorption is evident, showing shortening of the root length and rounding of the dental apice, the process is already in an advanced phase.

Under adverse conditions, dental ankylosis may also occur. Dental traumas provoke ankylosis as a consequence of the tissue damage and bleeding in some sites of the periodontal

ligament and the alveolar bone. The tissue reparative response promotes undesired fusion between cementum and alveolar bone. However, ankylosis may also have other reason: lack of masticatory function. In this scenario, lack of demand of tissue remodeling leads to atrophy of periodontal ligament fibers. The bone-cementum fusion can be an important consequence of such atrophic state.

9. Periodontal Ligament

In the human specie (and in other mammals) teeth are not rigidly united to the alveolar bone, but articulated to it by a fibrous connective mesh, called periodontal ligament. The periodontal ligament is a soft connective tissue, multifunctional and with unique proprieties, not showed by other tissues.¹⁰ The periodontal ligament is specialized in support, protection and sensorial proprioception of the masticatory system. It is a mesh of collagen fibers, of approximately only 0.2 to 0.3 mm of thickness, organized according to its functional purpose. Moreover, the periodontal ligament is the source of the progenitor cells of the anatomic-functional unit called supportive periodontium.

Although the fibroblasts of the periodontal ligament can not be originated from gingival cellular populations,⁵¹ both tissues have their development closely related. The gingival connective tissue and the periodontal ligament are fused during the dental eruption process and there is no anatomic distinction between both.⁷⁶ Besides the fibroblasts, the periodontal ligament presents endothelial cells, rest of Malassez epithelial cells, nervous sensorial system cells, blood vessels, cells associated with bone formation (formation of the alveolar bone radiographically called lamina dura) and cementoblasts.¹⁰

As other gingival tissues, most of the fibers of the periodontal ligament are made of type I and type III collagen,^{21,120,70,71,22,40} that are firmly attached to Cementum.¹⁰⁸ The type V and type XII fibers are present in smaller amount. Indeed, the blood vessels of the periodontal ligament present types I, III, IV and V collagen fibers.⁸

The extra-cellular matrix that holds periodontal ligament fibers was studied in details in sheep.⁴⁵ In such matrix there is small amount of elastin and tenascin,^{23,5,59} but the same proteoglycans of the gingival connective tissue were found. As examples, hyaluronan, heparan, dermatan and chondroitin,^{78,31} and versican and decorin,⁷⁸ are intrinsically imbricated among collagen fibers, cells and vessels. The biochemical composition of the proteoglycans of the periodontal ligament is altered in consequence of periodontal disease^{46,44} and during orthodontic movement.^{83,28,53,100}

One of the main functional features of periodontal ligament is its extremely high capacity of remodeling under local changes, presenting short cellular and protein turnover. The turnover of the collagen fibers of the periodontal ligament is double than of the attached gingiva.^{99,102,26} Such

active remodeling is necessary in a scenario where intense activity occurs, and none functional compromising is allowed. At the same time that many collagen fibers are destroyed, many others are generated.

Recent evidences suggest that such intense activity is possible because of the progenitor pluripotent stem cells supply. Such cells can also work as a reserve for cellular differentiation and potential reconstitution of the tissues affected by periodontal diseases.^{96,97,29} Such cellular differentiation is mediated by cytosines present since the dental eruption stage.^{124,125,36,56,57} Such cytosines are also identified in the inflammatory process after orthodontic movement,^{1,72,118,12} promoting a programmed bone remodeling.⁴³ This potential of response tends to decrease with aging,⁸⁶ what changes the expectation of orthodontic results in adult patients.¹⁰⁴ If orthodontic force is applied in an ankylosed tooth, which does not present periodontal ligament, bone remodeling around it is very limited. That is the same pattern of response induced on mini-implants, for temporary orthodontic anchorage.³⁷

10. Laminate Alveolar Bone

It is a specialized calcified connective tissue, dental-dependent, supported by trabecular alveolar bone, which is surrounded by the maxillary and mandibular cortical bones. Therefore, trabecular alveolar bone is recovered by laminate alveolar bone inside of the dental sockets, and surrounded by the bone cortices of the maxillary bones.⁸⁹ The alveolar bone and the cortices converge and are fused in the alveolar crests. The dental insertion system provided by the periodontal ligament is functionally adapted to the masticatory forces and to the contact between adjacent teeth.

The progenitor cells of the alveolar bone, with ectomesenchymal origin, are the osteoblasts. Osteoblasts make the bone matrix (osteoid), constituted by collagen fibers, glycoproteins and proteoglycans¹¹⁹ and mineralized by Calcium and Phosphate ions. Such deposition produces hydroxyapatite. Havers's channels allow the nutrition of osteocytes, cells that are locked in the mineralized bone and that communicate with periosteal osteoblasts. The "internal" surface of the alveolar bone is covered by endosteum, tissue that presents similar features of the periosteum.

The laminate alveolar bone is specialized as cementum and periodontal ligament. It also works for dental support. Its blood and lymphatic supply and its innervation are provided by the periodontal ligament throughout Volkman's channels, among perpendicular attachments of Sharpey's collagen fibers. The Havers' channels of the alveolar bone communicate with the Volkman's channels of the laminate alveolar bone, allowing instantaneous integration among bone, periodontal ligament and cementum.

Table 1. Top 10 most scientifically relevant articles of the paper

Year	Main Author	Title	Main Conclusions	Clinically Applied Evidence
1990	Bartold PM	A biochemical and immunohistochemical study of proteoglycans of alveolar bone.	The predominant glycosaminoglycan in human alveolar bone was chondroitin sulfate, although some hyaluronate, dermatan sulfate, and heparin sulfate were also detected. The proteoglycans were similar to those in cementum, dentin, and other bones.	No
1984	Chavrier C	Connective tissue organization of healthy human gingiva. Ultrastructural localization of collagen types I-III-IV.	Gingival connective tissue is made of an intricate pattern of type I and III collagen where type I fibers are preferentially organized in large dense bundles, whereas type III fibers present short and loose bundles pattern. Type IV collagen is the main collagen component of the basement membranes.	*
1989	Kirkham J	Site-specific variations in the biochemical composition of healthy sheep periodontium.	Collagen concentrations were highest at the alveolar bone and below the junctional epithelium adjacent to the tooth. Such patterns may influence the way in which periodontal disease is propagated through the tissue.	No
1995	Lang H	Formation of differentiated tissues in vivo by periodontal cell populations cultured in vitro	The alveolar bone contains cementogenic precursors with the potential to differentiate into active cementoblasts in the presence of a dental substrate.	No
1993	MacNeil RL	Molecular factors regulating development and regeneration of cementum.	The discussion focused on the factors/proteins important to the processes of formation and regeneration of the cementum. However, the specific cells responsible for stimulating new cementum formation have yet to be clarified.	No
1986	Pearson CH	Chemical and immunochemical characteristics of proteoglycans in bovine gingiva and dental pulp.	Chondroitin sulphate characterized the larger proteoglycans of gingiva and pulp; significant amounts of L-iduronic acid-rich dermatan sulphate or heparin sulphate were not present.	No
1997	Saffar JL	Alveolar bone and the alveolar process: the socket that is never stable.	Teeth migrate adaptively to crown attrition. The migration rate seems to peak during growth and declines afterwards without ceasing completely. Such mechanisms results in continuous plasticity of the alveolar wall around erupted and functioning teeth.	*
1997	Schroeder HE	The gingival tissues: the architecture of periodontal protection.	The tissues surrounding the teeth have been designed to provide a seal around the teeth (via the junctional epithelium and the epithelium attachment), to withstand the frictional forces of mastication and to defend the potential space between the teeth and the soft tissues against foreign invaders, such as microorganisms.	*
2004	Seo BM	Investigation of multipotent postnatal stem cells from human periodontal ligament.	PDL contains stem cells that have the potential to generate cementum/PDL-like tissue in vivo. Transplantation of these cells might hold promise as a therapeutic approach for reconstruction of tissues destroyed by periodontal diseases.	*
2008	Wise GE	Mechanisms of tooth eruption and orthodontic tooth movement.	A better appreciation of the molecular and cellular events that regulate osteoclastogenesis and osteogenesis in dental eruption and orthodontics is not only central to our understanding of these processes, but also is needed for ultimate development of the means to control them.	No

*Clinically Applied Evidence

Alveolar bone, laminate alveolar bone and periodontal ligament respond to the functional masticatory demand.⁸ In order to have such dynamic remodeling, continuously recycling of the Haversian system occurs. Bone suffers continuous resorption by osteoclasts and, at the same time, is neo-formed by osteoblasts.

The alveolar bone is trabecular (under lesser functional demand) in interproximal regions and it is compact (under greater functional demand) in the vestibular and palatine/lingual surfaces of the teeth, fusing with the laminate alveolar bone. In these later regions, there is also a

thickness difference: the bone is thicker in the vestibular and palatine/lingual surfaces in the molars region and thinner in the anterior teeth region.

The functional load imposed by antagonist teeth is not limited to the centric static position, but is also related to mandibular excursion movements. As clinical consequences, the vestibular and palatine/lingual laminae of the posterior teeth offer greater resistance to orthodontic movements.⁸⁷ Dental displacement is easier when the teeth moved in the “antero-posterior corridors” of the osseous dental arch.

When non-controlled orthodontic forces are applied in the

vestibular direction (for example, excessive palatine expansion), the bone laminae may suffer resorption. Bone fenestration or dehiscence may occur. The same happens in undesirable vestibular projection of the anterior teeth,¹²³ during orthodontic leveling and alignment mechanics. In lower incisors bone fenestration or dehiscence mainly occurs due to disrespect to negative arch discrepancy and consequent vestibular flaring.

Since bone remodeling is a dynamic, physiologic and natural process, orthodontic movement is well predictable, if well controlled. Orthodontic movement provokes inflammation in the periodontal ligament and increases its blood supply. Therefore, with the orthodontic activation occurs: 1) pain (in the first 3 to 5 days), 2) increase of the pericementary space (swallowing in the periodontal ligament) and, 3) increase in the local temperature (theoretical supposition, hard to be assessed). Such inflammatory process is aseptical in periodontally health patients and, if well planned, releases a desired and highly sophisticated biochemical process of bone deposition and resorption.⁶² In this process occurs bone deposition in the tension side (side “behind” the related orthodontic movement) and resorption in the pressure side (side “ahead” of the related orthodontic movement). Osteoblasts make the deposition line and osteoclasts make the resorption line.

During orthodontic bone resorption, the osteoclastic action provokes the detachment of the periodontal ligament of the bone and dental anchorage in the socket is partially lost. Bone deposition in the endosteal side of such site aims to counterbalance such resorption. In the other side of the orthodontic movement, where bone deposition takes place, mineralization of the osteoid collagen matrix allows that Sharpey’s fibers are incorporated into the neo bone, promoting new periodontal insertion. In this site there is no resorption process in the endosteal side. When the periodontium of the teeth submitted to orthodontic movement is normal, good “bone balance” is achieved, and the amount of neo bone built in the deposition side equalizes the bone deficit between resorption and deposition in the resorption side.

Ontogenetically, the cells which allow tissue adaptation to orthodontic forces are present since the first stages of dental formation. The normal process of eruption also requires bone deposition (and necessarily osteoblastogenesis) and bone resorption (and necessarily osteoclastogenesis).^{25,41} Sialoproteins and osteocalcins have been identified in the process of dental mineralization and in orthodontic movements.^{24,27} Proteinases have been identified in the process of bone resorption during dental formation and also in orthodontic movements.^{39,117,102,58,41}

Therefore, theoretically, orthodontic movements follow naturally developed physiologic patterns of dental eruption, and bone remodeling is naturally mediated by periodontal tissues.^{67,66} Scientifically pertinent, the biochemical comparison between dental eruption and orthodontic movements is still an ongoing research area.^{126,41}

11. Conclusions

Integration between the specialties Orthodontics and Periodontics, especially in adult patients, becomes crucial. In the table 01 the main results of the most scientifically relevant articles of the present paper are summoned. Such synergy in the modern and serious dental practice is fundamental, since favorable orthodontic movement is only achieved if the periodontal status of the patient is satisfactory. In this article we have described in a succinct way the main anatomic, histologic, physiologic, pathologic and therapeutic aspects of such integration, allowing basic knowledge for who wants to provide the best to the patients in these both areas.

REFERENCES

- [1] Alhashimi N, Frithiof L, Brudvik P, Bakheit M. Orthodontic movement and de novo synthesis of proinflammatory cytokines. *Am J Orthod Dentofac Orthop* 2001;119:307-12.
- [2] Attström R. Presence of leucocytes in crevices of healthy and inflamed gingiva. *J Periodont Res* 1970;5:42-7.
- [3] Attström R, Egelberg J. Presence of leucocytes within the gingival crevices during developing gingivitis. *J Periodontol Res* 1971;6:110-4.
- [4] Bartold PM. A biochemical and immunohistochemical study of proteoglycans of alveolar bone. *J Dent Res* 1990;69:7-19.
- [5] Bartold PM. Connective tissues of the periodontium. Research and clinical implications. *Aust Dent J* 1991;36:255-68.
- [6] Bartold PM. Distribution of chondroitin sulfate and dermatan sulfate in normal and inflamed human gingivae. *J Dent Res* 1992;71:1587-83.
- [7] Bartold PM, Wiebkin OW, Thonard JC. Preteoglycans of human gingival epithelium and connective tissue. *Biochem J* 1993;211:119-27.
- [8] Bartold PM. Turnover in periodontal connective tissues: dynamic homeostasis of cells, collagen and ground substances. *Oral Dis* 1995;1:238-53.
- [9] Becker J, Schuppan D, Rabanus JP, Rauch R, Niechoy U, Gelderblom HR. Immunoelectron microscopic localization of collagens Type I, V, VI and of procollagen Type III in human periodontal ligament and Cementum. *J Histochem Cytochem* 1991;39:103-10.
- [10] Beertsen W, McCulloch CAG, Sodek J. The periodontal ligament: a unique, multifunctional connective tissue. *Periodontology* 2000 1997; 13:20-40.
- [11] Birkadel-Hansen H, Butler WT, Taylor RE. Proteins of the periodontium. Characterization of the insoluble collagens of bovine Cementum. *Calcif Tiss Re* 1977;23:39-44.
- [12] Bletsa A, Berggreen E, Brudvik P. Interleukin-1 alpha and tumor necrosis factor-alpha expression during the early phases of orthodontic tooth movement in rats. *Eur J Oral Sci* 2006;114:423-9.

- [13] Bondevik O. Tissue changes in the rat molar periodontium following application of intrusive forces. *Eur J Orthod* 1980;2:41-9.
- [14] Bosshardt DD, Schroeder HE. Initiation of acellular extrinsic fiber Cementum on human teeth. A light- and electron-microscopic study. *Cell Tiss Res* 1991a;263:311-24.
- [15] Bosshardt DD, Schroeder HE. Establishment of acellular extrinsic fiber Cementum on human teeth. A light- and electron-microscopic study. *Cell Tiss Res* 1991b;263:325-36.
- [16] Bosshardt DD, Schroeder HE. Initial formation of cellular intrinsic fiber Cementum in developing human teeth. A light- and electron-microscopic study. *Cell Tissue Res* 1992;267:321-35.
- [17] Bosshardt DD, Schroeder HE. Cementogenesis reviewed: a comparison between human premolars and rodent molars. *Anat Rec* 1996;245:267-92.
- [18] Bosshardt DD, Selvig KA. Dental Cementum: the dynamic tissue covering of the root. *Periodontology* 2000 1997;13:41-75.
- [19] Braga AM, Squier CA. Ultrastructure of regenerating junctional epithelium in the monkey. *J Periodontol* 1980;51:386-92.
- [20] Brill N, Krasse B. The passage of tissue fluid into the clinically healthy gingival pocket. *Acta Odontol Scand* 1958;16:233-45.
- [21] Butler WT, Hansen HB, Beegle WF, Taylor RE, Chung E. Proteins of the periodontium: identification of collagens with the $[\alpha 1(1)]_2\alpha 2$ and $[\alpha 1(III)]_3$ structures in bovine periodontal ligament. *J Biol Chem* 1975;250:8907-12.
- [22] Chavrier C, Couble ML, Magloire H, Grimaud JA. Connective tissue organization of healthy human gingiva. Ultrastructural localization of collagen types I-III-IV. *J Periodont Res* 1984;19:221-9.
- [23] Chavrier C, Hartmann DJ, Couble ML. Distribution and organization of the elastic system fibers in healthy human gingiva. Ultrastructural and immunohistochemical study. *Histochemistry* 1988;89:47-52.
- [24] Chen J, Shapiro HS, Sodek J. Developmental expression of bone sialoprotein mRNA in rat mineralized connective tissue. *J Bone Miner Res* 1992;8:987-97.
- [25] Craddock HL, Youngson CC. Eruptive tooth movement – the current state of knowledge. *Brit Dent J* 2004;197:385-91.
- [26] Deporter DA, Svoboda EL, Howley TP, Shiga A. A quantitative comparison of collagen phagocytosis in periodontal ligament and transseptal ligament of the rat periodontium. *Am J Orthod* 1984;85:59-62.
- [27] Domon S, Shimokawa H, Yamaguchi S. Temporal and spatial mRNA expression of bone sialoprotein and type I collagen during rodent tooth movement. *Eur J Orthod* 2001;23:339-48.
- [28] Edwards JG. A study of the periodontium during orthodontics rotation of the teeth. *Am J Orthod* 1968;54:441-61.
- [29] Feng F, Akiyama K, Liu Y, Yamaza T, Wang T-M, Chen J-H et al. Utility of PDL progenitors for *in vivo* tissue regeneration: a report of three cases. *Oral Dis* 2010;16:20-8.
- [30] Gazit E, Lieberman M. Occlusal and orthodontic considerations in periodontally involved dentition. *Angle Orthod* 1980;50:346-9.
- [31] Gibson GJ, Pearson CH. Sulfated galactosaminoglycans of bovine periodontal ligament. Evidence for the presence of two major types of hybrid but no chondroitin sulfate. *Connect Tiss Res* 1982;10:161-71.
- [32] Goasland GD, Robertson PB, Mahan CJ, Morrison WW, Olson JV. Thickness of facial gingiva. *J Periodontol* 1977;48:768-71.
- [33] Gould TRL, Westbury L, Brunette DM. Ultrastructural study of the attachment of human gingiva to titanium *in vivo*. *J Prosthet Dent* 1984;52:418-20.
- [34] Hashimoto Y, Akagawa H, Nikai H, Tsuru H. Ultrastructure of the peri-implant junctional epithelium on single-crystal sapphire endosseous dental implant loaded with functional stress. *J Oral Rehabil* 1989;16:261-70.
- [35] Hashimoto S, Yamamura T, Shimono M. Morphometric analysis of intercellular space and desmosomes of rat junctional epithelium. *J Periodont Res* 1986;21:510-20.
- [36] Heinrich J, Bsoul S, Barnes J, Woodruff K, Abboud S. CSF-1, RANKL and OPG regulate osteoclastogenesis during murine tooth eruption. *Arch Oral Biol* 2005;50:897-908.
- [37] Hohlt WF, Roberts WE. Rigid implants for orthodontic anchorage. In: Davidovitch Z, ed. *Biological Mechanisms of Tooth Eruption, Resorption and Replacement by Implants*. Birmingham: EBSCO Media, pp.661-6, 1994.
- [38] Hopewell-Smith A. *The Histology and Pathology of the Teeth Associated Parts*. London: The Dental Manufacturing Company, 1903, apud Ten Cate AR. The role of epithelium in the development, structure and function of the tissues of tooth support. *Oral Dis* 1996;2:55-62.
- [39] Howard PS, Kucich U, Taliwal R, Korostoff JM. Mechanical forces alter extracellular matrix synthesis by human periodontal ligament fibroblasts. *J Periodontol Res* 1998;33:500-8.
- [40] Huang YH, Ohsaki Y, Kurisu K. Distribution of type I and type III collagen in the developing periodontal ligament of mice. *Matrix* 1991;11:25-35.
- [41] Huang XF, Zhao YB, Zhang FM, Han PY. Comparative study of gene expression during tooth eruption and orthodontic tooth movement in mice. *Oral Dis* 2009;15:573-9.
- [42] Keene DR, Sakai LY, Bachinger HP, Burgeson RE. Type III collagen can be present on banded collagen fibrils regardless of fibril diameter. *J Cell Biol* 1987;105:2393-402.
- [43] King GJ, Keeling SD, Wronski TJ. Histomorphometric study of alveolar bone turnover in orthodontic tooth movement. *Bone* 1991;12:401-9.
- [44] Kirkham J, Robinson C, Smith AJ. The effect of periodontal disease on sulphated glycosaminoglycan distribution in the sheep periodontium. *Arch Oral Biol* 1992;37:1031-7.
- [45] Kirkham J, Robinson C, Spence JA. Site-specific variations in the biochemical composition of healthy sheep periodontium. *Arch Oral Biol* 1989;34:405-11.
- [46] Kirkham J, Robinson C, Spence JA. Effect of periodontal disease ("Broken Mouth") on the distribution of matrix

- macromolecules in the sheep periodontium. *Arch Oral Biol* 1991;36:257-63.
- [47] Kollar EJ, Lumsden AGS. Tooth morphogenesis: the role of innervations during induction and pattern formation. *J Biol Buccale* 1979;7:49-60.
- [48] Kowashi Y, Jaccard F, Cimasoni G. Sulcular polymorphonuclear leucocytes and gingival exudate during experimental gingivitis in man. *J Periodont Res* 1980;15:151-8.
- [49] Kronfeld R. Coronal Cementum and coronal resorption. *J Dent Res* 1938;17:151-9., apud Ten Cate AR. The role of epithelium in the development, structure and function of the tissues of tooth support. *Oral Dis* 1996;2:55-62.
- [50] Kusters ST, Kuijpers-Jagtman AM, Maltha JC. An experimental study in dogs of transeptal fiber arrangement between teeth which have emerged in rotated or non-rotated positions. *J Dent Res* 1991;70:192-7.
- [51] Lang H, Schüler N, Arnhold S, Nolden R, Mertens T. Formation of differentiated tissues *in vivo* by periodontal cell populations cultured *in vitro*. *J Dent Res* 1995;74:1219-25.
- [52] Larjava H, Häkkinen L, Rahemtulla F. A biochemical analysis of human periodontal tissue proteoglycans. *Biochem J* 1992;284:267-74.
- [53] Last KS, Donkin C, Embery G. Glycosaminoglycans in human gingival crevicular fluid during orthodontic movement. *Arch Oral Biol* 1988;33:907-12.
- [54] Lindhe J. *Tratado de Periodontia Clínica e Implantologia Oral*. 5ª Edição, Rio de Janeiro: Guanabara-Koogan, pp. 32, 2008.
- [55] Listgarten MA. Electron microscopic features of the newly formed epithelial attachment after gingival surgery. *J Periodont Res* 1967;2:46-52.
- [56] Liu D, Yao S, Pan F, Wise GE. Chronology and regulation of gene expression of RANKL in the rat dental follicle. *Eur J Oral Sci* 2005;113:404-9.
- [57] Liu D, Yao S, Wise GE. Effect of interleukin-10 on gene expression of osteoclastogenic regulatory molecules in the rat dental follicle. *Eur J Oral Sci* 2006;114:42-9.
- [58] Luan X, Ito Y, Holliday S, Walker C, Daniel J, Galang TM, et al. Extracellular matrix-mediated tissue remodeling following axial movement of teeth. *J Histochem Cytochem* 2007;55:127-40.
- [59] Lukinmaa PL, Mackie EJ, Thesleff I. Immunohistochemical localization of the matrix glycoproteins-tenascin and the ED-sequence containing form of cellular fibronectin in human permanent teeth and periodontal ligament. *J Dent Res* 1991;70:19-26.
- [60] MacNeil RL, Somerman MJ. Molecular factors regulating development and regeneration of Cementum. *J Periodont Res* 1993a;28:550-9.
- [61] MacNeil RL, Thomas HF. Development of the murine *periodontium*. Role of basement membrane in formation of a mineralized tissue on the developing root dentin surface. *J Periodontol* 1993b;64:95-102.
- [62] Masella RS, Meister M. Current concepts in the biology of orthodontic tooth movement. *Am J Orthod Dentofac Orthop* 2006;129:458-68.
- [63] Mazeland GRJ. The mucogingival complex in relation to alveolar process height and lower anterior face height. *J Periodont Res* 1980a;15:345-52.
- [64] Mazeland GRJ. Longitudinal aspects of gingival width. *J Periodont Res* 1980b;15: 429-33.
- [65] McKinney RV Jr., Steflik DE, Koth DL. Evidence for a junctional epithelial attachment to ceramic dental implants. A transmission electron microscopic study. *J Periodontol* 1985;56:579-91.
- [66] Merzel J, Salmon C. Growth and the modeling/remodeling of the alveolar bone of the rat incisor. *Anat Rec* 2008;291:827-34.
- [67] Moxham BJ, Berkovitz BKB. The periodontal ligament and physiological tooth movements. In: Berkovitz BKB, Moxham BJ, Newman HN, eds. *The Periodontal Ligament in Health and Disease*. London: Mosby-Wolfe, pp. 183-210, 1995.
- [68] Nakata TM, Stepnick RJ, Zipkin I. Chemistry of human dental Cementum: the effect of age and fluoride exposure on the concentration of ash, fluoride, calcium, phosphorus and magnesium. *J Periodontol* 1972;43:115-24.
- [69] Narayanan AS, Page RC. Biochemical characterization of collagens synthesized by fibroblasts derived from normal and diseased human gingiva. *J Biol Chem* 1976;251:5464-71.
- [70] Narayanan AS, Page RC. Connective tissues of the periodontium. A summary of current work. *Coll Rel Res* 1983;3:33-64.
- [71] Narayanan AS, Clagett JA, Page RC. Effect of inflammation on the distribution of collagen types I, III, IV and V and type I trimmer and fibronectin in human gingivae. *J Dent Res* 1985;64;1111-6.
- [72] Oshiro T, Shiotani A, Shibasaki Y, Sasaki T. Osteoclast induction in periodontal tissue during experimental movement of incisors in osteoprotegerin-deficient mice. *Anat Rec* 2002;266:218-25.
- [73] Owens PDA. Patterns of mineralization in the roots of premolar teeth in dogs. *Arch Oral Biol* 1975;20:709-12.
- [74] Owens PDA. Ultrastructure of Hertwig's epithelial root sheath during early root development in premolar teeth in dogs. *Arch Oral Biol* 1978;23:91-104.
- [75] Oyarzun-Droguett A. Ultracytochemical localization of basal lamina anionic sites in the rat epithelial attachment apparatus. *J Periodont Res* 1992;27:256-63.
- [76] Palmer RM, Lubbock MJ. The soft connective tissues of the gingiva and periodontal ligament: are they unique? *Oral Dis* 1995;1:230-7.
- [77] Paynter KJ, Pudy G. A study of the structure, chemical nature, and development of Cementum in the rat. *Anat Rec* 1958;131:233-51, apud Ten Cate AR. The role of epithelium in the development, structure and function of the tissues of tooth support. *Oral Dis* 1996;2:55-62.
- [78] Pearson CH, Gibson GJ. Proteoglycans of bovine periodontal ligament and skin. Occurrence of different hybrid-sulphated galactosaminoglycans in distinct proteoglycans. *Biochem* 1982;201:27-37.

- [79] Pearson CH, Pringle GA. Chemical and immunochemical characteristics of proteoglycans in bovine gingiva and dental pulp. *Arch Oral Biol* 1986;31:541-548.
- [80] Perry ME. The specialized structure of crypt epithelium in the human palatine tonsil and its functional significance. *J Anat* 1994;185:111-27.
- [81] Polson AM, Subtelny JD, Meitner SW, Poison AP, Sommers EW, Iker HP, Reed BE. Long-term periodontal status after orthodontic treatment. *Am J Orthod Dentofac Orthop* 1988;93:51-8.
- [82] Potter-Perigo S, Prather P, Baker C, Altman LC, Wight TN. Partial characterization of proteoglycans synthesized by human gingival epithelial cells in culture. *J Periodont Res* 1994;28:81-91.
- [83] Reitan K. Tissue rearrangement during retention of orthodontically rotated teeth. *Am J Orthod* 1959;29:105-13.
- [84] Reitan K. Clinical and histologic observations on tooth movement during and after orthodontic movement. *Am J Orthod* 1967;53:721-45.
- [85] Reitan K. Biomechanical principles and reactions. In: *Current Orthodontic Concepts and Techniques*. Graber TM & Swain BF eds., St. Louis: CV Mosby, pp. 101-92, 1985.
- [86] Ren Y, Maltha JC, Stokroos L, Liem RSB, Kuijpers-Jagtman AM. Age-related changes of periodontal ligament surface areas during force application. *Angle Orthod* 78:1000-5, 2008.
- [87] Ricketts RM. Perspectives in the Clinical Application of Cephalometrics: The First Fifty Years. *Angle Orthod* 1981;5:115-50.
- [88] Saario M, Ainamo A, Mattila K, Ainamo J. The width of radiologically-defined attached gingiva over permanent teeth in children. *J Clin Periodontol* 1994;21:666-9.
- [89] Saffar JL, Lasfargues JJ, Cherruau M. Alveolar bone and the alveolar process: the socket that is never stable. *Periodontology* 2000 1997;13:76-90.
- [90] Saito I, Watanabe O, Kawahara M, Igarashi Y, Yamamura T, Shimono M. Intercellular junctions and the permeability barrier in the junctional epithelium. A study with freeze-fracture and thin sectioning. *J Periodont Res* 1981;16:467-80.
- [91] Schroeder HE. Quantitative parameters of early human gingival inflammation. *Arch Oral Biol* 1970;15:383-400.
- [92] Schroeder HE. Transmigration and infiltration of leucocytes in human junctional epithelium. *Helv Odontol Acta* 1973;17:6-18.
- [93] Schroeder HE, Listgarten MA. Fine structure of the developing epithelial attachment of human teeth. *Monographs in Developmental Biology*. Karger: Basel, Vol. 2, 1977.
- [94] Schroeder HE, Listgarten MA. The gingival tissues: the architecture of periodontal protection. *Periodontology* 2000 1997;13:91-120.
- [95] Selvig KA, Selvig SK. Mineral content of human and seal Cementum. *J Dent Res* 1962;41:624-32.
- [96] Seo BM, Miura M, Gronthos S, Bartold PM, Batouli S, Brahimi J, et al. Investigation of multipotent postnatal stem cells from human periodontal ligament. *Lancet* 2004;364:149-55.
- [97] Seo BM, Miura M, Sonoyama W, Coppe C, Stanyon R, Shi S, et al. Recovery of functional postnatal stem cells from cryopreserved human periodontal ligament. *J Dent Res* 2005;84:907-12.
- [98] Shibutani T, Murahashi Y, Iwayama Y. Immunohistochemical localization of chondroitin sulfate and dermatan sulfate proteoglycan in human gingival connective tissue. *J Periodont Res* 1989;24:310-3.
- [99] Sodek J. A comparison of the rates of synthesis and turnover of collagen and non-collagen proteins in adult rat periodontal tissues and skin using a microassay. *Arch Oral Biol* 1977;22:655-65.
- [100] Sodek J. Collagen turnover in periodontal ligament. In: *Biology of Tooth Movement*. Norton AL & Burstone CJ ed., Boca Raton: CRC Press, pp. 157-81, 1989.
- [101] Svoboda ELA, Shiga A, Deporter DA. A serologic analysis of collagen phagocytosis by fibroblasts in three soft connective tissues with differing rates of collagen turnover. *Anat Rec* 1981;199:473-80.
- [102] Takahashi I, Onodera K, Nishimura M, Mitnai H, Sasano Y, Mitani H. Expression of genes for gelatinases and tissue inhibitors of metalloproteinases in periodontal tissues during orthodontic tooth movement. *J Mol Histol* 2006;37:333-42.
- [103] Takata T, Nikai H, Miyauchi M, Ito H, Kobayashi J, Ijuhin N. Lectin binding of rat gingival epithelia. *J Periodont Res* 1990;25:151-5.
- [104] Tanne K, Yoshida S, Kawata T, Sasaki A, Knox J, Jones ML. An evaluation of the biomechanical response of the tooth and periodontium to orthodontic forces in adolescent and adult subjects. *Br J Orthod* 1998;25:109-15.
- [105] Temple TR, Snyderman R, Jordan HV, Mergenhagen SE. Factors from saliva and oral bacteria, chemotactic for polymorphonuclear leucocytes: their possible role in gingival inflammation. *J Periodontol* 1970;41:71-80.
- [106] Ten Cate AR, Mills C, Solomon G. The development of the periodontium. A transplantation and autoradiographic study. *Anat Rec* 1971;170:365-79.
- [107] Ten Cate AR. Morphological studies of fibrocytes in connective tissue undergoing rapid remodeling. *J Anat* 1972;112:401-14.
- [108] Ten Cate AR. Formation of supporting bone in association with periodontal organization in the mouse. *Archs Oral Biol* 1975;20:137-8.
- [109] Ten Cate AR. A fine structural study of coronal and root dentinogenesis in the mouse. Observations on the so-called "von Korff fibers". *J Anat* 1978;125:183-97.
- [110] Ten Cate AR. *Oral Histology: Development, Structure and Function*. 4th Edition. St. Louis: Mosby, pp.53, 1994.
- [111] Ten Cate, AR. The experimental investigation of odontogenesis. *Int J Dev Biol* 1995;39:5-11.
- [112] Ten Cate AR. The role of epithelium in the development,

- structure and function of the tissues of tooth support. *Oral Dis* 1996;2:55-62.
- [113] Ten Cate AR, Deporter DA, Freeman E. The role of fibroblast in the remodeling of periodontal ligament during physiologic tooth movement. *Am J Orthod* 1976;69:155-68.
- [114] Ten Cate AR. The development of the periodontium - a largely ectomesenchymally derived unit. *Periodontol* 2000 1997; 13:9-19.
- [115] Thomas HF. Root formation. *Int J Dev Biol* 1995;39:231-7.
- [116] Thomas HF, Kollar EJ. Differentiation of odontoblasts in grafted recombinations of murine epithelial root sheath and dental mesenchyme. *Arch Oral Biol* 1989;34:27-35.
- [117] Tsubota M, Sasano Y, Takahashi I, Kagayama M, Shimauchi H. Expression of MMP-8 and MMP-13 mRNAs in rat periodontium during tooth eruption. *J Dent Res* 2002;81:673-8.
- [118] Tuncer BB, Ozmeric N, Tuncer C, Teoman I, Cakilci B, Yucel A. Levels of interleukin-8 during tooth movement. *Angle Orthod* 2005;75:539-44.
- [119] Waddington RJ, Embery G. Structural characterization of human alveolar bone proteoglycans. *Conn Tiss Res* 1991;36:859-66.
- [120] Wang HM, Nanda V, Rao LG, Melcher AH, Heersche JNM, Sodek J. Specific immunohistochemical localization of type III collagen in porcine periodontal tissues using the peroxidase-anti-peroxidase method. *J Histochem Cytochem* 1980;28:1215-23.
- [121] Wiebkin OW, Thonard JC (1981). Mucopolysaccharide localization in gingival epithelium. *J Periodont Res* 1981;16:600-10.
- [122] Wiesmann HP, Mever U, Plate U, Höhling HJ. Aspects of collagen mineralization in hard tissue formation. *Int Rev Cytol* 2005;242:121-56.
- [123] Wingard CE, Bowers GM. The effect on facial bone from facial tipping of incisors in monkeys. *J Periodontol* 1976;47:450-4.
- [124] Wise GE, Lumpkin SJ, Huang H, Zhang Q. Osteoprotegerin and osteoclast, differentiation factor in tooth eruption. *J Dent Res* 2000;79:1937-42.
- [125] Wise GE, Yao S, Odgren PR, Pan F. CSF-1 regulation of osteoclastogenesis for tooth eruption. *J Dent Res* 2005;84:837-41.
- [126] Wise GE, King GJ. Mechanisms of tooth eruption and orthodontic tooth movement. *J Dent Res* 2008;87:414-34.
- [127] Wolfram K, Egelberg J, Hornbuckle C, Oliver R, Rathbun E. Effect of tooth cleaning procedures on gingival sulcus depth. *J Periodont Res* 1974;9:44-9.
- [128] Yamashiro T, Tummers M, Thesleff I. Expression of bone morphogenetic proteins and Msx genes during root formation. *J Dent Res* 2003;82:172-6.