Synthesis and Study of the Antibacterial Activity of Polyvinylpirrolidone Polymeric Complexes with Levofloxacin

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Abstract In the present work, chemical immobilization of levofloxacin into macromolecules of a synthetic polymer, polyvinylpyrrolidone, was carried out. By varying the molar ratio of PVP: levofloxacin in the reaction medium, polymer complexes were obtained in which the quantitative content of the drug was 12.3-38.5%. The structure of the synthesized polymer compounds was confirmed by IR and UV spectroscopy, and the molecular characteristics of the reaction products were also calculated. The kinetics of cleavage of levofloxacin from polyvinylpyrrolidone macromolecules at pH 2.0 and 7.4 was studied. According to data the most optimal is the use of an antibacterial complex with a molar ratio of PVP and levofloxacin respectively is 1:0.5.

Keywords Fluoroquinolones, Levofloxacin, Polyvinylpyrrolidone, Polymer complex, Antibacterial properties

1. Introduction

One of the most practical ways to obtain physiologically active polymers with a pharmacological property, is the chemical immobilization of low molecular weighted biologically active compounds to macromolecules of natural or synthetic polymers. It is known that the modification of low molecular weight drugs with various polymers is carried out in order to improve their therapeutic properties, eliminate existing side effects, increase the duration of their action in the body, ensure targeted transport of the active substance and increase bioavailability [1,2]. Due to a sharp increase in the resistance of pathogens of infectious diseases to low molecular weight antimicrobial drugs used in clinical practice, a search is being intensively aimed at creating antimicrobial macromolecular systems consisting of a polymer matrix and an active substance [3-5]. Mainly, the specific properties of the graft polymer designed according to this principle are determined by the physicochemical characteristics, molecular weight and the type of chemical bond formed between the drug and the macromolecular chain [6,7]. Therefore, by optimizing the final structure and the necessary characteristics of the created antimicrobial polymer, it is possible to obtain new drugs that are devoid of the original disadvantages and have the desired pharmacological properties.

Clinical studies have shown that fluoroquinolones are

highly effective antibacterial drugs that are very often used in the treatment of infections, including resistant forms of tuberculosis [8-10]. Despite a wide range of antimicrobial activity, the main disadvantages of fluoroquinolone preparations are low bioavailability and the presence of various side effects on the body during long-term use. For example, levofloxacin, an antibacterial drug that is part of the third-generation fluoroquinolones. Levofloxacin is often prescribed in clinical practice because it is highly effective against most strains of bacterial pathogens responsible for respiratory, urinary, gastrointestinal and abdominal infections. However, the half-life of levofloxacin is 6-8 hours. In this connection, to create the necessary therapeutic concentration in an infected organism it is necessary to increase the daily dosage of levofloxacin. And as a result, there is a clinical detected cardiotoxicity (prolongation of the QT interval), nephrotialysis, hyperkalemia, allergic reactions. etc.

Based on the above-described relevance of scientific and practical problems, in this work we carried out the chemical modification of levofloxacin with a synthetic hydrophilic polymer polyvinylpyrrolidone (PVP). The choice of PVP as a polymer matrix for levofloxacin can be substantiated by the fact that this polymer is highly soluble in water, has unique physicochemical properties, is nontoxic, and contains reactive functional groups in its structure that easily bind drugs by complex formation [10-12].

The objective of this work was to obtain and study the biological activity of polymer complexes of levofloxacin with polyvinylpyrrolidone, in which the antibacterial agent is associated with hydrophilic polymer macromolecules by a

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hydrolyzable type of chemical bond.

2. Materials and Methods

To carry out the chemical modification of levofloxacin with hydrophilic polymers, we used polyvinylpyrrolidone with a molecular weight of 12000 Da, purified by extraction with diethyl ether.

2.1. Chemical Modification of Levofloxacin with Polyvinylpyrrolidone

0.0135 mole (1.5 g) of PVP was dissolved at room temperature in 50 ml of water until a clear solution was formed. After complete dissolution of the polymer with vigorous stirring, 0.00135-0.0067 mole (0.49-2.44 g) of levofloxacin was added. The reaction solution was stirred for 2 hours and t=20°C. The resulting clear solution was then dialyzed against distilled water for 24 hours using dialysis bags with a protein cutoff of 5000 Da. The final products were isolated by sublimation of water from a pre-frozen aqueous solution.

2.2. Study of the Structure and Molecular Characteristics of Polymer Complexes of Levofloxacin with Polyvinylpyrrolidone

UV spectra of aqueous solutions of the synthesized samples were taken on a "UV 1280" spectrophotometer (Shimadzu, Japan) in the range λ =180-400 nm. IR spectra were recorded on a Vector-22 spectrometer in the wavelength range 400-4000 cm⁻¹ in KBr tablets (3 mg sample/300 mg KBr). The molecular weight characteristics of the synthesized derivatives were determined by high performance gel permeation chromatography on an Agilent 1260 Infiniti liquid chromatograph. The analysis process was monitored using a refractometric detector using a PL Aquagel OH Mixed chromatographic column 300 mm long and 7.5 mm in inner diameter (Waters, USA). A 0.1 N aqueous solution of sodium nitrate was used as the eluent. The volume flow rate of the eluent was 0.8 ml/min.

2.3. Determination of Antibacterial Properties of Polymer Complexes of Levofloxacin with Polyvinylpyrrolidone *in vitro*

We chose the following microorganisms as test cultures for determining the antimicrobial activity of levofloxacin polymer complexes: *S.aureus*, *Pr.mirabilis*, *E.faecalis*, *E.coli, and Ps.aeruginosa.* The antimicrobial effect of the synthesized compounds was studied at a concentration of 100μ g/ml by the agar diffusion method, based on the ability of the studied substances to diffuse into the nutrient medium on which the selected test cultures are sown and to suppress the growth of microorganisms (mm). The diameter of microbial growth inhibition zones less than 10 mm was evaluated as the absence of antimicrobial activity, 10-15 mm - weak activity, 15-20 mm - moderately pronounced activity, more than 20 mm - pronounced activity. Each sample was tested in three parallel experiments. The obtained data were statistically processed taking into account Student's criterion [13].

2.4. Study of the Release of Levofloxacin from PVP Macromolecules

The release rate of levofloxacin from polymeric complexes was assessed by dialysis through а semipermeable membrane with a protein transmission limit of 3000 Da. The hydrolysis of polymer complexes was studied in physiological solution with pH 2.0 and 7.4 at a temperature of 25°C using equipment for the study of equilibrium dialysis. The selected polymer complex dissolved in 5 ml of physiological solution was placed in the left cell of the setup. The optical density of the prepared solution λ_{max} =294 nm was 2.23. Then, 5 ml of physiological solution was placed in the right cell. At certain intervals, samples of 0.5 ml of the solution were taken from the right cell and diluted 10 times, and the amount of levofloxacin that passed through the cellophane membrane of the right cell was determined spectrophotometrically at λ_{max} =294 nm in the resulting solution. The degree of hydrolysis (S,%) of the polymer complex was calculated as the ratio of D_{294} in the right cell at time τ to the initial value of D₂₉₄ in the left cell and expressed in % [14].

3. Results and Their Discussion

The high reactivity of polyvinylpyrrolidone is associated primarily with its ability to form complex compounds (due to the presence of >N-C=O groups in the macromolecule) with substances containing -COOH groups in the structure. Therefore, the chemical binding of levofloxacin to PVP functional groups will proceed according to the following scheme:



Polymer complex

To determine the limiting number of lefoloxacin molecules that can bind to PVP macromolecules, we carried out the reaction at different molar ratios of the starting reagents (Table 1). The molecular characteristics of the obtained samples were also found.

 Table 1. Influence of the molar ratio of levofloxacin on the composition and molecular characteristics of the reaction products

Samples	Molar ratio of PVP: levofloxacin	Levofloxacin content,%	Molecular weight Da	[n] [*] , ml/g
1	1:0.1	12.3	13400	0.137
2	1:0.25	25.4	14200	0.145
3	1:0.5	37.2	15100	0.152
4	1:1.0	38.5	16000	0,158

Note. $[\eta]^*$ - intrinsic viscosity of the samples, calculated in a 0.1 N solution of sodium acetate in water at 25 °C.

From the data in Table 1, it can be seen that with an increase in the molar ratio of levofloxacin in the reaction medium from 0.1 to 0.5 mole, a regular increase in the quantitative content of the antimicrobial drug in the

composition of PVP from 12.3 to 37.2% is observed. Accordingly, an increase in the mass fraction of levofloxacin in the hydrophilic polymer macrochain leads to an increase in the molecular weight of the reaction products. At the molar ratio of PVP: levofloxacin=1:1 the content of the fluoroquinolone agent in the composition of the polymer complex remained practically unchanged and amounted to only 38.5%. All obtained samples, regardless of the content of levofloxacin, were highly soluble in water.

In the IR spectra (Fig. 1), the formation of the complex is characterized by a decrease in the intensity of the absorption band of the initial polyvinylpyrrolidone at 2930 cm⁻¹, related to the intramolecular hydrogen bond, and a shift in the band of the intermolecular H-bond of the polymer from the region of 3430 cm^{-1} to the region of 3390 cm^{-1} . Also, the presence of a hydrogen bond in the polymer complex is proved by the strengthening and merging of the absorption bands of the functional groups of the substance of levofloxacin -COO-(1725 cm⁻¹) and C=O (1620 cm⁻¹) of the groups belonging to PVP, which characterizes the interaction of levofloxacin with polyvinylpyrrolidone through acid - COOH group.



Figure 2. UV spectra of povinylpyrrolidone, levofloxacin, and synthesized polymer complexes 1 and 3

As can be seen from Fig. 2, the UV spectrum of levofloxacin is characterized by an absorption band in the region of λ_{max} =294 nm. In contrast to the UV spectrum of levofloxacin, the synthesized polymer complexes have several absorption maxima (λ_{max} =215 and 294 nm). Absorption in the region of λ_{max} =294 nm proves the presence of a fluoroquinolone agent in the composition of PVP. The resulting absorption bands of the synthesized complexes in the region of λ_{max} =215 nm belong to the PVP lactam cycle. It should also be noted that with an increase in the quantitative content of levofloxacin in PVP, the intensity of the band at λ_{max} =294 nm increases.

Since levofloxacin is used orally and intravenously in the body, the kinetics of the release of the fluoroquinolone agent from PVP macromolecules was studied by dialysis through a semipermeable membrane in a buffer solution with pH 2.0 and 7.4.

As can be seen from the results presented in Fig. 3, the rate of release of levofloxacin from PVP proceeds at different rates. Data comparison shows that at pH 2.0 the release of levofloxacin from PVP is more intense than at pH 7.4. For example, the degree of hydrolysis of the polymer complex at pH 2.0 after 24 and 48 hours is -74 and 82.5%, and at pH 7.4 - 53 and 62%. This indicates the different stability of the formed hydrogen bonds between levofloxacin and PVP functional groups to the pH conditions of the medium. Also, from these data it follows that the chemical attachment of levofloxacin to PVP gives a prolonged release of antibacterial drug molecules into the physiological environment.

Figure 4 shows the chromatograms of PVP and the polymer complex corresponding to the maximum of the chromatographic peak.

In the presented chromatograms of PVP and the polymer complex, there are no other low-intensity peaks responsible for the presence of free levofloxacin. The available peaks are of narrow intensity, which indicates that the synthesized polymer complex is narrowly dispersed and does not contain impurities of unreacted levofloxacin.



Figure 3. Kinetics of Levofloxacin Release from the Polymer Complex at pH 2.0 and 7.4 (Levofloxacin Content in the Polymer Complex is 37.2%)



Figure 4. Gel chromatograms of PVP and polymer complex-3 with a quantitative content of levofloxacin 37.2%

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Microorganisms	Levofloxacin	Sample №1	Sample №2	Sample №3	Sample №4
S.aureus	35±1.3 mm	27±0.8 mm	30±1.5 mm	35±1.2 mm	35±1.0 mm
Pr.mirabilis	35±1.0 mm	30±1.5 mm	32±1.3 mm	36±1.0 mm	40±1.1 mm
E.faecalis	38±0.8 mm	35±1.2 mm	35±1.5 mm	35±0.6 mm	33±0.8 mm
E.coli	42±1.2 mm	34±1.3 mm	40±1.0 mm	42±1.3 mm	42±1.4 mm
Ps.aeruginosa	34±0.5 mm	25±1.0 mm	30±0.5 mm	33±1.5 mm	35±1.0 mm

Table 2. Antibacterial activity of levofloxacin and its polymer complexes in vitro

Note: the concentration of the studied drugs was 100 $\mu\text{g/ml}$

The antibacterial properties of PVP polymer complexes with levofloxacin against *S.aureus*, *Pr.mirabilis*, *E.faecalis*, *E.coli and Ps.aeruginosa* were studied. The results of microbiological tests under in vitro conditions are shown in table 2.

The results presented in Table 2 indicate that all synthesized samples are characterized by pronounced antibacterial properties, since they effectively inhibit the growth of pathogenic bacteria. At the same time, the data obtained demonstrate a different level of antibacterial activity of the obtained polymer complexes against microorganisms. From *in vitro* tests, it follows that the size of the growth inhibition zone of bacterial cultures increases with an increase in the quantitative content of levofloxacin in the composition of PVP. The highest antibacterial activity, comparable with low molecular weight levofloxacin at the same concentration, is shown by complex 3 and 4, in which the content of the active substance is 37.2 and 38.5%, which is almost 2.7 and 2.6 times less compared to levofloxacin.

4. Conclusions

Thus, by complexation of the >N-C=O groups of PVP and -COOH groups of levofloxacin, water-soluble polymer complexes with different drug content of 12.3-38.5% were obtained for the first time. By carrying out the release kinetics of the polymer complex, it was proved that the degree of hydrolysis of levofloxacin from PVP macromolecules at pH 2.0 after 24 and 48 hours is -74 and 82.5%, and at pH 7.4 - 53 and 62%. Based on the comparative studies conducted, it can be concluded that the preparation of an antibacterial complex with a molar ratio of PVP: levofloxacin = 1:0.5 according to the conditions of synthesis, therapeutic activity, technological and practical reproducibility is the most optimal.

REFERENCES

- I. Ekladious, Y.L. Colson, M.W. Grinstaff, Polymer-drug conjugate therapeutics: advances, insights and prospects, Nature Reviews Drug Discovery. 2018. doi:10.1038/s41573-018-0005-0.
- [2] T.G. Barclay, C.M. Day, N. Petrovsky, S. Garg, Review of polysaccharide particle-based functional drug delivery,

Carbohydrate Polymers. 221, 2019, 94-112. doi:10.1016/j.carbpol.2019.05.067.

- [3] Sh.A., Shomurotov, O.R. Akhmedov, A.S. Turaev, G.Kh. Mamadullaev, Antituberculosis activity and pharmacokinetics of polymer conjugates of isoniazid and ethambutol, Pharmaceutical Chemistry Journal. 6, 2021, 67-71. doi:10.30906/0023-1134-2021-55-6-23-27.
- [4] N.D. Stebbins, M.A. Ouimet, K.E. Uhrich, Antibioticcontaining polymers for localized, sustained drug delivery, Advanced Drug Delivery Reviews. 78, 2014, 77-87. doi:10.1016/j.addr.2014.04.006.
- [5] O.R. Akhmedov, S.A. Shomurotov, G.G. Rakhmanova, A.S. Turaev Synthesis and Study of Biological Activity of Sulfamic Polysaccharide Derivatives, Russian Journal of Bioorganic Chemistry. 43(7), 2017, 716-721. doi:10.1134/S1068162017070020.
- [6] A. Chen, H. Peng, I. Blakey, A.K. Whittaker, Biocidal Polymers: A Mechanistic Overview, Polymer Reviews. 57(2), 2016, 276-310. doi:10.1080/15583724.2016.1223131.
- [7] G.G. Zhanel, K. Ennis, L. Vercaigne, A. Walkty, A.S. Gin, J. Embil, D.J. Hoban, A Critical Review of the Fluoroquinolones, Drugs. 62(1), 2002, 13-59. doi:10.2165/00003495-200262010-00002.
- [8] I.M. Le-Deygen, A.A. Skuredina, E.V. Kudryashova, Drug delivery systems for fluoroquinolones: New prospects in tuberculosis treatment, Russian Journal of Bioorganic Chemistry. 43(5), 2017, 487-501. doi:10.1134/s1068162017050077.
- [9] N. G. Bush, I. Diez-Santos, L.R. Abbott, A. Maxwell, Quinolones: Mechanism, Lethality and Their Contributions to Antibiotic Resistance, Molecules. 25(23), 2020, 5662. doi:10.3390/molecules25235662.
- [10] Y. Luo, Y. Hong, L. Shen, F. Wu, Multifunctional Role of Polyvinylpyrrolidone in Pharmaceutical Formulations, AAPS PharmSciTech, 22(34), 2021, 1-16. doi:10.1208/s12249-020-01909-4.
- [11] P. Franco, I. De Marco, The Use of Poly(N-vinyl pyrrolidone) in the Delivery of Drugs: A Review, Polymers. 12(5), 2020, 1114. doi:10.3390/polym12051114.
- [12] P. Franco, I. De Marco, The Use of Poly(N-vinyl pyrrolidone) in the Delivery of Drugs: A Review, Polymers (Basel). 12(5), 2020, 1114. doi:10.3390/polym12051114.
- [13] S. Glants Medical and Biological Statistics. Moscow: Praktika; 1999. 459 p.
- [14] M.V. Solovskii, N.V. Nikol'skaya, N.A. Zaikina, Synthesis and properties of polymeric schiff bases of the antibiotic

spiramycin, Pharmaceutical Chemistry Journal. 36(2), 2002, 7-11. doi:10.1023/a:1016051612229.

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