

Ecological Assessment of Brewery Effluent Impact on the Macroinvertebrates of Ikpoba River, Edo State, Nigeria

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Abstract An investigation into the impact of brewery factory effluent to the macroinvertebrate assemblage in Ikpoba River was carried out from the months of April to September 2012. Samples were collected from different three sites, designated as station 1 (upstream of effluent discharge point), station 2 (effluent discharge point) and station 3 (downstream of effluent discharge point). The collected samples were analyzed for the physicochemical characteristics while bank root substrate samples were sorted, identified and counted for macroinvertebrate composition. A total of seven macroinvertebrate species comprised of 803 individuals were identified. These include four species of Diptera and one species each for Hemiptera, Lepidoptera, and Lepidoptera. Among the macroinvertebrates, Diptera dominated contributing 61.39% of the total density occurrence, while Lumbriculida, Lepidoptera and Hemiptera, made up of 33.25%, 3.23% and 0.13, respectively. The highest total number of taxa was observed in station 1 with a total of seven taxa with 256 individuals; whereas, station 2 and station 3 each with a total of four taxa with 71 and 488 individuals, respectively. The changes observed in some of the macroinvertebrate assemblages of Ikpoba River, notably increased indicating inefficient effluent treatment in the breweries. This study has also shown that the quality of receiving water is influenced significantly by the chemical composition of effluents discharged into the River.

Keywords Brewery factory effluent, Macroinvertebrates, Ikpoba River, Water quality, Aquatic pollution

1. Introduction

In the past two decades, there has been a tremendous fear of possible global biodiversity crisis arising out of a rapidly accelerating loss of species, populations and natural habitats such as tropical rainforests and wetlands. It has been estimated that more than half of the habitable surface of the planet has already been altered by human activity [1]. McCarthy [2] also suggested that we are on the verge of mass extinctions of species. These concerns stem from the realization that our knowledge of the diversity and variability of plants, animals, microorganisms and the ecosystems in which they occur is woefully incomplete.

All water bodies, either lentic or lotic, contain vast numbers of invertebrate macrofauna (species visible to the unaided eye) which are used to assess water quality in rivers [3]. Not only are they numerically abundant, but they are also taxonomically diverse. Hynes [4] observed that the aquatic animals, which live on, in or near the substratum of running

waters, include the nematodes, annelids, insects, crustaceans and molluscs. These organisms are important in the aquatic ecosystem because they form part of the aquatic food chain. They are an essential element of the food chain, especially for larger aquatic animals. They feed on algae and bacteria which occupy the bottom of the food chain.

Biological communities have been seen as effective tools for assessing organic pollution. Macroinvertebrate animals can be sampled quantitatively and are easy to monitor because they respond to man-made disturbance. However, the assessment of biological communities present in an aquatic environment also reflects the quality of the ecosystem [5]. The monitoring of biological communities can be done at a variety of trophic levels including micro-organisms (bacteria, protists, and viruses), primary producers (algae and vascular plants), primary consumers (invertebrates) and secondary consumers (fish) [6, 7]. Macroinvertebrate assemblages are useful indicators of localized conditions because many have limited migration patterns or are sessile (non-motile). Thus, are helpful in examining site-specific impacts [5]. Changes in the environment will be reflected in variations in the species assemblage both spatially and temporally (i.e., affected and unaffected sites over time).

Some studies on macroinvertebrates and other

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related topics have been carried out in zone Nigerian rivers [8, 9, 10]. Although studies on macroinvertebrate communities in rivers in response to brewery effluent have not been given much attention in Nigeria and Africa in general, Ogbey and Egborge [8] recorded a total of 214 invertebrate taxa in the Okomu Reserve, comprising 80 zooplankton and 134 macroinvertebrates. Evagelopoulos *et al.* [11] studied the spatial variations of both phytoplankton and macroinvertebrates descriptors (composition, abundance, and biomass) at six sampling sites in the low-salinity ponds of Kalloni Saltworks. Their studies revealed 54 macroinvertebrate taxa belonging to 5 groups. Pearson and Rosenberg [12] noted that organic enrichment of sediments as a result of sewage

and other organic contaminants may lead to a series of non-linear changes in the abundance, biomass and diversity of macroinvertebrate, in both spatial and temporal patterns. Although the composition, distribution and abundance of macroinvertebrates in Nigeria have been reported in some studies [13, 14, 15]. However, no study has been carried out to assess the impact of brewery factory effluent on the macroinvertebrate community of Ikpoba River, hence, the need for this study. This study is therefore aimed at evaluating the impact of brewery effluent on the macroinvertebrate of Ikpoba River, and to establish a relationship between the current physicochemical properties and the macroinvertebrate of Ikpoba River.

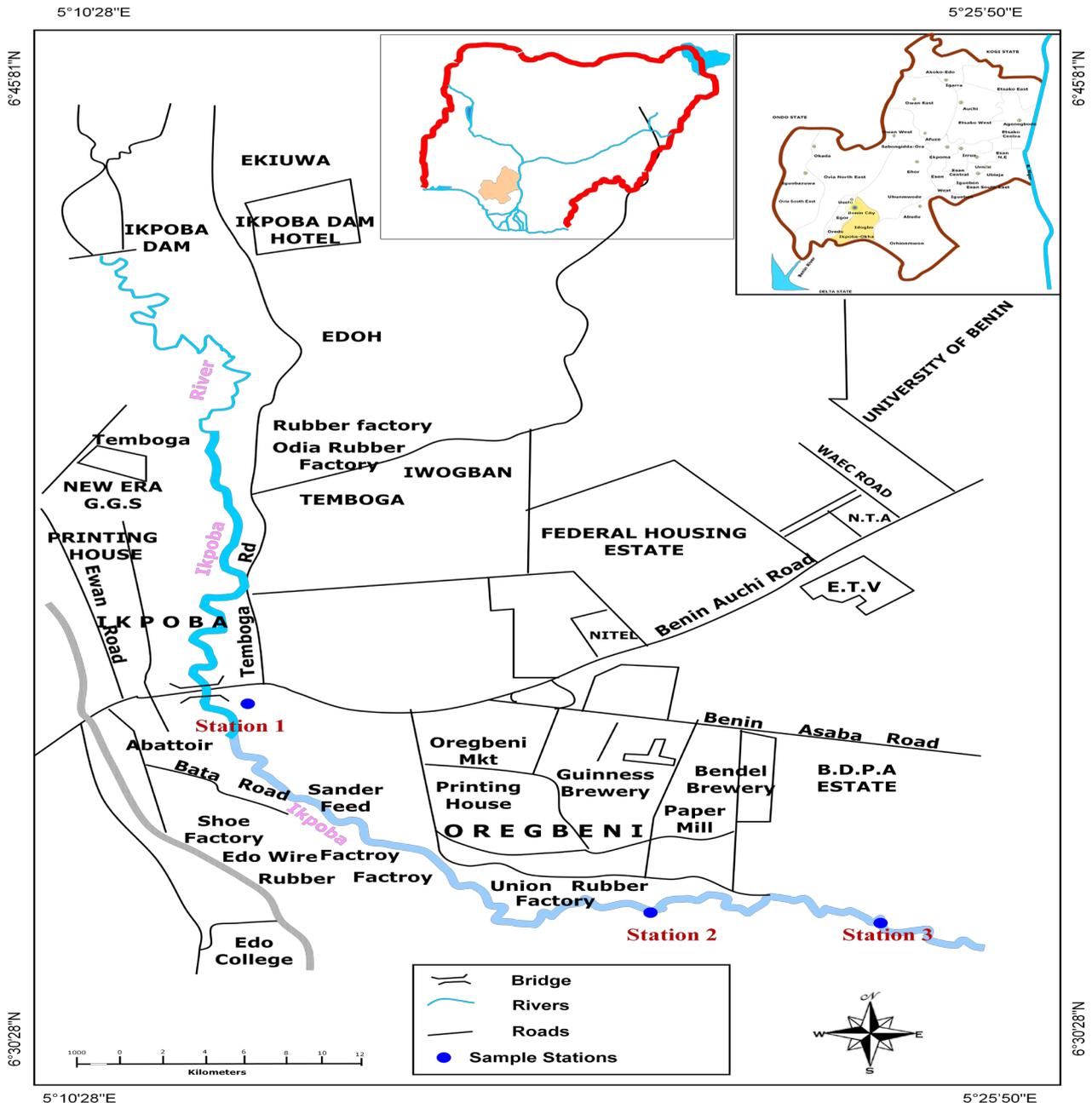


Figure 1. Map of the study area (Map of Nigeria and Edo State as inset)

2. Materials and Methods

2.1. Study Area

Ikpoba River, a fourth order stream, is located in Benin City, Edo State in South Nigeria (Lat 6.5°N, Long 5.8°E). Its headwater originates from North West of Benin City and flows north to south through the city (Benka-Coker and Ojior, 1995). The river flows through a dense rain forest where the allochthonous input of organic matter from the surrounding vegetation is derived through run-off from the surface of the soil.

Typically, the region have the characteristic features of the humid tropical wet and dry climate governed primarily by the rainfall. The vegetation of Ikpoba River consists of rainforest which is secondary in nature and has been greatly subjected to deforestation and other anthropogenic activities. The main vegetation consists of rubber palm, bamboo plant; and palm trees with their symbiont epiphytic ferns.

2.2. Sampling Stations

Three sampling stations were selected for this study. Station 1 is the upstream section of the river, the point, just after the bridge. Water velocity is high, and depth is minimal when compared to other stations. At this section the water is clear, but the substratum is silted. Human activities include fishing and domestic activities such as bathing and washing of clothes and cars, mechanic workshops and the activities of the local deity shrine.

Station 2, was located 400m downstream of station 1. It was the contact point of the brewery effluent with the river water before it flows down to station 3. The substratum is coarsely sandy with pebbles. The river bank is muddy and has a large deposit of spent grain which has accumulated over time as a result of the effluent flow. The water is highly turbid, and distinct odour of decaying organic materials. The activities in this region include fishing.

Station 3 was located about 100m downstream of station 2. The water is also turbid similar to that of station 2. The activities in this station include distillation of palm wine to alcoholic beverages, fishing and farming activities.

2.3. Collection of Water and Bankroot Samples

Monthly sampling of Ikpoba River was undertaken throughout the wet season months from April to September 2012. Samples were collected between the hours of 0900 and 1200 on each sampling day and at each time, sample collection commenced in station 1 and terminated in station 3.

Standard methods and instrumentations were followed during sample collection procedures [17]. At each station, the surface water temperature was taken in-situ. Surface water samples for physicochemical analyses were collected into thoroughly cleaned 1liter polyethylene bottles and tightly closed. Each bottle was rinsed with the appropriate sample before the final sample collection. The samples were placed in a cooler box and then taken to the laboratory for

analyses. For dissolved oxygen (DO) determinations, separate samples were collected in 300 ml plain glass bottles and the samples fixed using the azide modification of Winkler's method [17]. Samples for biochemical oxygen demand (BOD) were collected into dark glass bottles for incubation and subsequent DO determination. In the laboratory, pH, total dissolved solids, conductivity, turbidity, dissolved oxygen, biochemical oxygen demand (BOD₅), chemical oxygen demand (COD), chloride, sodium, potassium, magnesium, sulphate, nitrate, phosphate, lead, iron, zinc, cadmium and copper were determined according to procedures outlined in the Standard Methods for the Examination of Water and Wastewater [17]. The pH was measured using HACH digital meter, total dissolved solid was determined using a TDS meter (Model 4076), conductivity was measured using Cybersan 510 conductivity meter, while turbidity was determined using a DR/2000 HACH spectrophotometer. The titrimetric method was applied in the determination of chemical oxygen demand (COD), chloride, sodium, potassium, and magnesium. Sulphate, nitrate, and phosphate were determined spectrophotometrically at 380nm, 470nm, and 680nm respectively. Heavy metal such as lead, iron, zinc, cadmium and copper in the water sample was determined using Atomic Absorption Spectrophotometer (Buck Scientific Model-210) at 217.0nm, 248.0nm, 213.9nm, 228.8nm and 324.8nm respectively.

The modified 'Kick sampling technique' [18, 19] was used in collecting benthic macroinvertebrates from the bank root biotope of each station. A hand net (154mm mesh size) was used in sampling 0.25m² of the substratum at four different points to the form of one composite sample per station. Collected samples were preserved with 10% formaldehyde in polyethylene containers. In the laboratory, macrobenthic invertebrates in the sample from the three stations were sorted out with the aid of a binocular dissecting microscope (25-40X American Optical Binocular microscope, model 570) and preserved in 4% formalin. The observed specimens were identified to the species level with the aid of identification keys [20, 21] while counting was done manually.

2.4. Statistical Analysis

Statistical package for Social Sciences (SPSS 16.0) to test for significant differences in the physicochemical parameters using One-way analysis of variance (ANOVA), where significant values ($p < 0.05$) were obtained, 'A posteriori' Duncan Multiple Range Test was subsequently applied to all pairs of means to detect the location of difference. Palaeontological statistical software package for education and data analysis (PAST 1.97) was used to evaluate relationships (such as diversity indices, taxa richness and species evenness) between macrobenthic invertebrates, while pearson correlation was used to establish comparison between macrobenthic assemblage and physicochemical parameters.

3. Results and Discussion

Physico-chemical parameters (water temperature, total dissolved solids, electric conductivity, turbidity, BOD, COD, pH and DO) and chemical parameters (chloride, sodium, potassium, magnesium, sulphate, nitrate, phosphate, lead, iron, zinc, and cadmium) were found highest ($p < 0.05$) at the discharge point (Station 2) than the upstream (Stations 1) and downstream (Station 3) stations. This result confirms previous reports on some aquatic studies [25]. The low dissolved oxygen concentration recorded agreed with values reported for some Nigerian waters [26]. The low dissolved oxygen values revealed anoxic condition during the study period. Such low oxygen concentration has been reported in Eruvie Stream and Ikpoba River as a result of the discharge of organic-rich effluent [25, 27]. Low dissolved oxygen has been reported to be deleterious to most aquatic fauna [33].

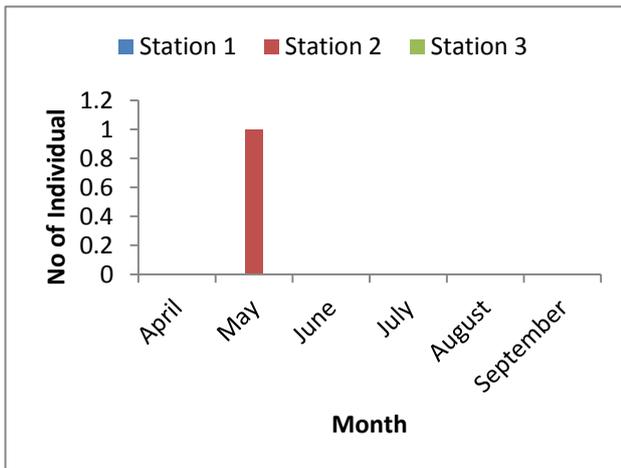


Figure 2. Monthly variation of the family Hemiptera

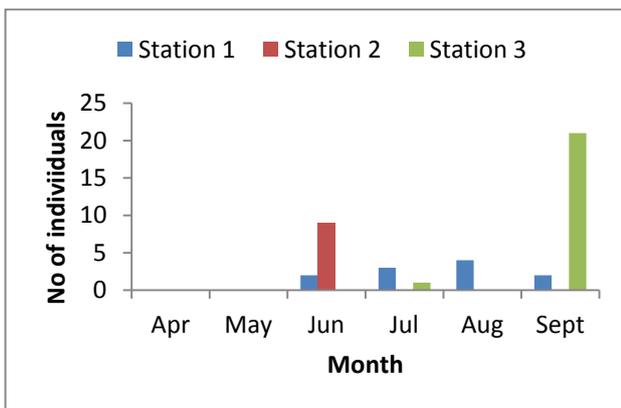


Figure 3. Monthly variation of the family Lepidoptera

Hemiptera represented by *Pelocoris femoratus* was present only at station 2 in the month of May (Fig 2). Lepidoptera represented only by *Elophila turbata* was present in all stations. It appeared in station 1 throughout the months of June, July, August, and September with the highest value reported in August (Fig 3). It appeared only once in station 2 at the month of June; in station 3, *Elophila turbata* appeared briefly in low quantity in July while a higher number was reported in September. Diptera was

represented by *Ablabesymia* species, *Callicera spinolae*, *Chironomus riparius* and *Eristalis tenax*. These macrobenthos species in station 1 increased steadily throughout the period of study. In station 2, dipterans appeared in the months of July until August. Whereas, in station 3, it was present in higher number than the other stations from the month of June to September. Lumbriculida was represented only by *Lumbriculus variegatus*. It was present in station 3 throughout the period of study. It decreased in number from April to September (Fig 5).

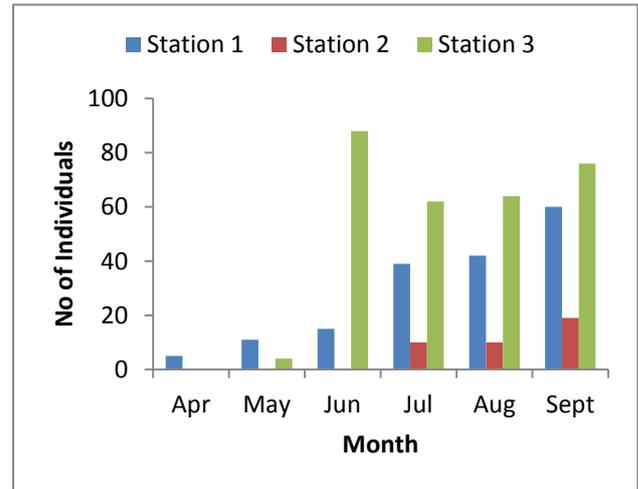


Figure 4. Monthly variation of the family Diptera

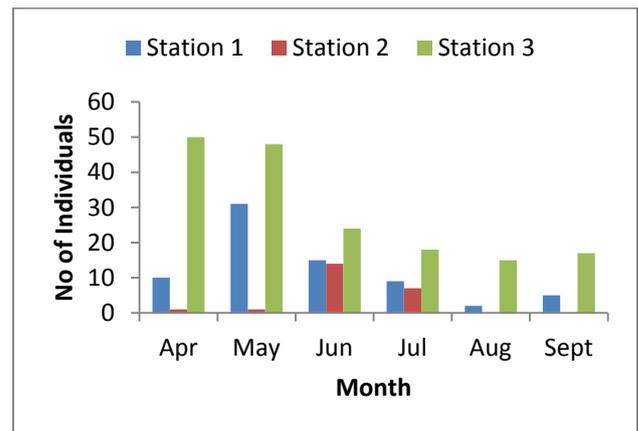


Figure 5. Monthly variation of the family Lumbriculida

A total of 7 macrobenthic invertebrate taxa comprising 803 individuals include 1 species each of Hemiptera, Lumbriculida and Lepidoptera; and 4 species of Diptera. The overall taxa composition, distribution, and abundance of macrobenthic invertebrates collected during the study period are presented in Table 1. This value was very low when compared with over 55 taxa reported for tropical streams [6, 13, 28, 29]. However, amongst the study station, the lowest abundance was reported in station 2. The low species diversity could be due to some physicochemical conditions of the river like high temperature, TDS, conductivity, turbidity, BOD, COD, chloride, sodium, potassium, magnesium, sulphate, low dissolved oxygen and high heavy metal of the water. Again, the abundance of the benthic

macroinvertebrate improved at station 3. According to Emere and Nasiru [30] diversity tends to be low in physically controlled systems. These factors probably caused disruption of life cycle, reproductive cycle, food chain, and migrations or imposed physiological stress on even the tolerant macroinvertebrates [29]. In this study, the responses of the macrobenthic organism to brewery factory effluent pollution varied greatly among orders and individual species.

The total number of taxa reported for station 1 was 7 with a total number of 256 individuals (Table 2). Stations 2 and 3 both have a total of four taxa with 71 and 488 individuals, respectively. The result showed a low diversity and density of benthic macroinvertebrates in the Station 2 which may be attributed to the increased influx of effluent from brewery factory and human interference that perhaps have led to the alteration of the biotic and abiotic characteristics of the river. Similar findings have been reported by Chowdhary and Sharma [31], who reported decrement in the quality and quantity of the diversity and density of benthic macroinvertebrate in Tawi River, India as a result of an increase in pollution load along the river channel.

Individual organisms were dominated by Diptera, which constituted 61.39% density occurrence, while Hemiptera, Lepidoptera and Lumbriculida made up of 0.13, 3.23 and

33.25% respectively. The order Diptera chiefly comprised of four species, out of which *Chironomus* sp. dominated and recorded the highest number of individuals in the highly polluted sections of Ikpoba River. It is well documented that the family Chironomidae is considered to be a pollution tolerant families of order Diptera is due to the presence of haemoglobin pigment that helps them to collect oxygen directly from the atmosphere. In addition to this, other plausible reasons of increment in Dipteran population in the downstream sections of Ikpoba River may be due to reduced flow of water, high nutrient load, increased human interference and increased decomposition. Numerical abundance of dipterans in the downstream sections of the water bodies have also been reported by many workers from their respective works [32, 33, 34, 35].

The presence of *Pelocoris femoratus*, *Callicera spinolae* and *Eristalis tenax* only in the upstream (Station 1) may be attributed to higher oxygen content, lesser organic load, and lesser human activities compared to other stations. Whereas, their complete absence in station 2 perhaps revealed the sensitivity of these species to polluted waters. It has been observed that different species in the same genus may react differently to pollution [36].

Table 1. Some physical and chemical conditions of Ikpoba River water at three sampling stations from April to September 2012

Environmental Parameter	Station 1			Station 2			Station 3			WHO Limit
	Mean (\pm SE)	Min	Max	Mean (\pm SE)	Min	Max	Mean (\pm SE)	Min	Max	
pH	6.43 \pm 0.26	5.57	6.90	6.04 \pm 0.45	4.58	6.86	6.63 \pm 0.36	5.49	7.34	6.5-9
Temperature ($^{\circ}$ C)	27.83 \pm 0.48 ^b	26.00	29.00	37.00 \pm 0.52 ^a	36.00	39.00	29.67 \pm 0.33 ^b	29.00	31.00	27
TDS (mg/l)	14.59 \pm 1.65 ^b	11.22	21.10	552.97 \pm 74.12 ^a	432.00	918.00	33.88 \pm 5.15 ^b	17.10	47.00	1000
Conductivity (μ s/cm)	22.62 \pm 2.18 ^b	18.00	33.08	829.65 \pm 31.82 ^a	755.00	931.00	41.98 \pm 6.73 ^b	28.00	73.70	-
Turbidity (NTU)	4.75 \pm 1.01 ^b	2.02	9.05	88.61 \pm 75.92 ^a	9.47	468.18	8.88 \pm 4.31 ^b	1.96	23.97	5
Dissolved Oxygen (mg/l)	3.29 \pm 0.52	0.99	4.30	1.81 \pm 0.21	1.10	2.37	2.94 \pm 0.76	1.12	6.55	5
BOD ₅ (mg/l)	2.68 \pm 0.94	1.20	6.86	2.85 \pm 1.06	1.11	8.06	2.38 \pm 0.92	0.72	5.36	28-30
COD (mg/l)	4.12 \pm 0.56	2.83	5.96	5.69 \pm 0.56	3.71	7.85	2.97 \pm 0.43	1.83	4.38	-
Chloride (mg/l)	3.41 \pm 0.73 ^b	1.18	5.29	90.96 \pm 26.24 ^a	3.88	138.76	7.74 \pm 2.36 ^b	1.73	14.52	400
Sodium (mg/l)	1.48 \pm 0.17 ^b	0.91	1.98	39.71 \pm 27.64 ^a	7.27	176.89	3.34 \pm 0.68 ^b	1.04	6.11	-
Potassium (mg/l)	1.81 \pm 0.74 ^b	0.53	5.29	44.57 \pm 26.12 ^a	1.11	145.32	4.30 \pm 2.16 ^a	0.59	14.52	
Magnesium (mg/l)	0.51 \pm 0.13	0.13	1.08	5.36 \pm 3.02	0.78	18.36	1.19 \pm 0.46	0.29	2.82	150
Sulphate (mg/l)	1.64 \pm 0.31 ^b	0.36	2.42	21.25 \pm 10.83 ^a	2.52	74.48	2.31 \pm 0.60 ^b	0.82	4.42	400
Nitrate (mg/l)	2.60 \pm 1.90 ^b	0.39	12.08	14.60 \pm 8.29 ^a	1.14	43.29	3.88 \pm 1.93 ^b	0.52	10.95	50
Phosphate (mg/l)	0.58 \pm 0.17 ^b	0.21	1.10	13.11 \pm 7.76 ^a	0.63	37.64	1.32 \pm 0.39 ^a	0.44	2.50	-
Lead (mg/l)	0.01 \pm 0.00 ^b	0.00	0.01	0.14 \pm 0.05 ^a	0.04	0.37	0.01 \pm 0.01 ^b	0.00	0.03	0.05
Iron (mg/l)	0.64 \pm 0.14 ^b	0.02	0.90	8.10 \pm 5.85 ^a	1.24	37.24	0.84 \pm 0.43 ^b	0.00	2.35	1.0
Zinc (mg/l)	0.14 \pm 0.10 ^b	0.01	0.61	0.58 \pm 0.12 ^a	0.11	0.97	0.08 \pm 0.02 ^b	0.01	0.14	5
Cadmium (mg/l)	0.01 \pm 0.01 ^b	0.00	0.04	0.47 \pm 0.24 ^a	0.01	1.41	0.02 \pm 0.01 ^b	0.00	0.05	0.01
Copper (mg/l)	0.04 \pm 0.03	0.00	0.20	0.31 \pm 0.17	0.01	0.93	0.17 \pm 0.10	0.00	0.52	1.5

Note: Similar letters indicate means that there are not significant differences

Table 2. Percentage abundance of species and individuals of the major groups

	No of taxa	% taxa	No of individuals	% individual
Hemiptera	1	14.29	1	0.13
Lepidoptera	1	14.29	42	3.23
Diptera	4	57.14	493	61.39
Lumbriculida	1	14.29	267	33.25

Table 3. General ecological indices for macroinvertebrate from Ikpoba river

	Station 1	Station 2	Station 3
Diversity indices	1.196	1.295	1.041
Richness indices	1.082	0.7038	0.4846
Evenness indices	0.4723	0.9128	0.7077

Diversity indices provide important information about rarity and commonness of species in a community [37]. From table 5 it was noted that the diversity indices of the macroinvertebrate from Ikpoba River slightly increased in station 2 when compared to station 1, and decreased significantly in station 3; again the species evenness or equitability increased from station 1 to 2 then decreased in station 3; while species richness decreased across that station (from 1 to 3) during the period of study (Table 3). The major factor that affects the occurrence and distribution of macroinvertebrate in lotic and lentic systems include, physicochemistry, the nature of the substrate, (bed material), water current, food availability, flood, drought, vegetation and shade [38].

Table 4. Composition, Distribution, and Abundance of Macroinvertebrates in Ikpoba River, Benin City from April – September 2012

	Station 1	Station 2	Station 3
Hemiptera			
<i>Pelocoris femoratus</i>	1	0	0
Lepidoptera			
<i>Elophila turbata</i>	11	9	22
Diptera			
<i>Ablabesymia</i> species	41	12	37
<i>Callicera spinolae</i>	1	0	0
<i>Chironomus riparius</i>	129	27	257
<i>Eristalis tenax</i>	1	0	0
Lumbriculida			
<i>Lumbriculus variegatus</i>	72	23	172
Total	256	71	488

The relationship between the physicochemical parameters and the occurrence of the macroinvertebrate of Ikpoba River was examined using the Pearson's correlation analysis, and the results were shown in the correlation matrix (Table 4). *Ablabesymia* species correlated significantly with pH ($r = 0.499$), *Callicera spinolae* correlated significantly only with the DO ($r = 0.586$), *Chironomus riparius* correlated inversely with total dissolved solid ($r = -0.491$), EC ($r = -0.513$) and COD ($r = -0.569$); while it varied directly with dissolved oxygen ($r = 0.548$). *Lumbriculus variegatus* correlated inversely with total dissolved solid ($r = -0.492$), EC ($r = -0.485$) and turbidity ($r = -0.527$).

Table 5. Correlation Matrix for physical and chemical parameters of Ikpoba River from April to September 2012

	<i>Pelocoris femoratus</i>	<i>Elophila turbata</i>	<i>Ablabesymia</i> species	<i>Callicera spinolae</i>	<i>Chironomus riparius</i>	<i>Eristalis tenax</i>	<i>Lumbriculus variegatus</i>
pH	-0.227	0.381	0.499	-0.215	0.289	-0.227	0.06
Temp	-0.326	-0.022	-0.244	-0.208	-0.362	-0.326	-0.492
TDS	-0.166	-0.125	-0.382	-0.172	-0.491	-0.166	-0.485
EC	-0.18	-0.115	-0.373	-0.177	-0.513	-0.18	-0.527
Turb	-0.058	-0.129	-0.229	-0.074	-0.229	-0.058	-0.216
DO	-0.273	0.054	0.215	0.586	0.548	-0.273	-0.191
BOD	-0.169	-0.282	-0.34	0.600	-0.173	-0.169	-0.005
COD	0.223	-0.378	-0.458	0.255	-0.569	0.223	-0.350
Cl ⁻	-0.129	-0.055	-0.254	-0.113	-0.368	-0.129	-0.398
Na	-0.095	-0.144	-0.28	-0.081	-0.299	-0.095	-0.312
K	-0.109	-0.213	-0.313	-0.115	-0.333	-0.109	-0.342
Mg	-0.069	-0.182	-0.333	-0.098	-0.302	-0.069	-0.238
SO ₄ ²⁻	-0.117	-0.108	-0.278	-0.087	-0.311	-0.117	-0.335
NO ₃ ⁻	0.098	-0.228	-0.38	-0.111	-0.308	0.098	-0.165
PO ₄ ³⁻	-0.08	-0.173	-0.312	-0.091	-0.307	-0.08	-0.295
Pb	-0.117	-0.215	-0.317	-0.12	-0.383	-0.117	-0.406
Fe	-0.071	-0.137	-0.252	-0.071	-0.27	-0.071	-0.272
Zn	-0.212	0.074	-0.299	-0.217	-0.403	-0.212	-0.373
Cd	-0.062	-0.112	-0.22	-0.061	-0.237	-0.062	-0.238
Cu	-0.032	-0.197	-0.32	-0.098	-0.248	-0.032	-0.165

NB: Bold font: Significant correlation

4. Conclusions

The changes observed in some of the macrobenthic assemblages of Ikpoba River, notably increased indicating inefficient effluent treatment in the breweries. This study has also shown that the quality of receiving water is influenced significantly by the chemical composition of effluents discharged into the River. High values of diversity indexes in the upstream sections of Ikpoba River reflected the stability of the physical and chemical characteristics of the river in the upstream sections. On the other hand, low values of diversity found in the downstream sections may be due to the increased influence of human activities. Because Ikpoba River serves as a source of domestic water supply for the local community for drinking, washing, fishing and swimming, all specified quality criteria must be met by the breweries especially before the discharge.

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REFERENCES

- [1] Hengeveld, R. *Dynamic Biogeography*. Cambridge University Press, Cambridge. UK, 1990.
- [2] McCarthy M.P. Freshwater ecosystem management from theory to application. *International Journal of Water* 2002, 2(1), 1–16.
- [3] Elliot, J.M.; Drake, C.M. A comparative study of four dredges used for sampling macro-invertebrates in rivers. *Freshwater Biology* 1981, 11, 245–261.
- [4] Hynes, H.B.N. The ecology of stream insects. *Annual Review of Entomology* 1972, 15, 25–42.
- [5] Olumukoro, J.O.; Ezemonye, L.I.N. Assessment of the macroinvertebrate fauna of rivers in southern Nigeria. *African Zoology* 2007, 42(1), 1-11.
- [6] Victor, R.; Ogbeibu, A.E. Macrobenthic invertebrates of a stream flowing through a farmland in southern Nigeria. *Environmental Pollution (Series A)* 1985, 39, 337-349.
- [7] Edokpayi, C.A.; Osimen, E.C. Hydrobiological Studies on Ibiekuma River at Ekpoma, Southern Nigeria, after Impoundment: The Faunal Characteristics. *African J. of Sci. and Tech.* 2001, 2(1), 72-81.
- [8] Ogbeibu, A.E.; Egborge, A.B.M. Hydrobiological studies of water bodies in the Okomu Forest Reserve (Sanctuary) in Southern Nigeria. 1. Distribution and diversity of the Invertebrate fauna. *Tropical Freshwater Biology* 1995, 4, 1–27.
- [9] Imoobe, T.O.T.; Ohoizebau, E. Pollution status of a tropical forest river, using aquatic insects as indicator. *African Journal of Ecology* 2009, 48, 232 – 238.
- [10] Olumukoro J.O.; Dirisu, A. Macroinvertebrate community of a post Lindane treated stream flowing through derived savannah in Southern Nigeria. *Tropical Freshwater Biology*, 2012, 21(1) 67- 82.
- [11] Evagelopoulous, A.; Spyrakos, E.; Karydis, M.; Koutsoubas, D. The biological System of Kalloni Saltwork and Variations of Macrobenthic Invertebrate Community Structure along the Salinity gradient in the low Salinity ponds. *Proceedings of the annual Ecologist and the Greek Zoological Society, Mytilene*, 2005, 18, 58 – 64.
- [12] Pearson, T. H.; Rosenberg, R. Macrobenthic succession in relation to organic enrichment and pollution of the marine environment. *Oceanogr. Mar. Biol. a. Rev.* 1978, 16, 229–311.
- [13] Edokpayi, C.A.; Okenyi, J.C.; Ogbeibu A.E.; Osimen, E.C. The effect of human activities on the macrobenthic invertebrates of Ibiekuma Stream, Ekpoma, Nigeria. *Biosci. Res. Commun.* 2000, 12, 79-87.
- [14] Edema, M.O.; Omemu, A.M.; Fapetu, O.M. Microbiological and physicochemical analysis of different sources of drinking water. *Nigerian Journal of Microbiology* 2001, 15, 57-61.
- [15] Ogbeibu, A.E.; Oribhator, B.J. Ecological impact of river impoundment using benthic macroinvertebrates as indicators. *Water Research* 2002, 36, 2427–2436.
- [16] Benka-Coker, M.O.; Ojior, O.O. Effect of slaughterhouse wastes on the water quality of Ikpoba River, Nigeria. *Biosci. Tech.* 1995; 52, 5-12.
- [17] American Public Health Association. Standard methods for the Examination of Water and Waste water. 20th edn. American Public Health Association, Washington. 1998; 1134 pp.
- [18] Armitage, P.D. Downstream changes in the composition, numbers and biomass of bottom fauna in the Tees below Cow Green Reservoir and an unregulated tributary, Maize Beck, in the first five years after impoundment. *Hydrobiologia* 1978, 58, 145–56.
- [19] Lenat, D.R.; Penrose, D.L.; Eagleson, K.W. Variable effects of sediment addition on stream benthos. *Hydrobiologia* 1981, 79, 187–94.
- [20] Macan, T.T. *A guide to fresh water invertebrates animal*. Longman, London. 1959; 307pp.
- [21] Mellanby, H. *Animal life in freshwater – A guide to freshwater invertebrate*. Chapman and hall Ltd, London. 1963; 308pp.
- [22] Green, R.H. *Sampling Design and Statistical method for Environmental biologist*. John Willey and Sons, Toronto. Ont. 1971; 257pp.
- [23] Robinson, A. H. and Robinson, P. K. Seasonal distribution of Zooplankton in the Northern Basin of Lake Chad. *J. 2001 land*, 1971, 163, 25 – 61.
- [24] Zar, J.H. *Biostatistical Analysis*, 2nd edn. Prentice Hall Inc., New Jersey. 1984; 348pp.

- [25] Imoobe, T.O.T.; Okoye, P.I. Assessment of the impact of effluent from a soft drink processing factory on the physico-chemical parameters of Eruvbi stream Benin City, Nigeria. *Bayero Journal of Pure and Applied Science* 2011, 4(1), 126 – 134.
- [26] Ofojekwu, P.C.; Umar, D.N.; Onyeka, J.O.A. Pollution status of Industrial wastes and their effects on macroinvertebrate distribution along Anglo–Jos water channel, Jos, Nigeria. *Journal of Aquatic Science* 1996, 11, 1-6.
- [27] Ekhaise, F.O.; Anyasi, C.C. Influence of Breweries Effluent Discharge on the Microbiological and Physico-chemical Quality of Ikpoba River, Nigeria. *Biotechnology* 2005, 4(10), 1062 – 1065.
- [28] Ogbeibu A.E. Distribution Density and Diversity of Dipterans in a Temporary pond in Okomu Forest Reservoir, Southern Nigeria. *Journal of Aquatic Science* 2001, 16, 43-52.
- [29] Adakole, J.A.; Anunne, P.A. Benthic macroinvertebrates as indicators of environmental quality of an urban stream, Zaria, Northern, Nigeria. *Journal of Aquatic Science* 2003, 18(2), 85-92.
- [30] Emere, M.C.; Nasiru, C.E. Macroinvertebrates as indicators of the water quality of an urbanized stream in Kaduna Nigeria. *Journal of Fishery International* 2007, 2(2), 152-157.
- [31] Chowdhary, S.; Sharma, K.K. Evaluation of macrobenthic invertebrates in the longitudinal profile of a River (Tawi), originating from Shivalik Hills. *Journal of Global Biosciences* 2013, 2(1), 31-39.
- [32] Arimoro, F.O.; Ikomi, R.B.; Efemuna, E. Macroinvertebrate community patterns and diversity in relation to water quality status of river Ase, Niger Delta. *Nigeria. Journal of fisheries and Aquatic Science* 2007, 2(5), 337-344.
- [33] Edokpayi, C.A.; Olowoporoku, A.O.; Uwadiae, R.E. The hydrochemistry and macrobenthic fauna characteristics of an urban draining creek. *International Journal of Biodiversity and Conservation* 2010, 2(8), 196-203.
- [34] Gopal, B.; Sah, M. Conservation and Management of rivers in India: Case study of the river Yamuna. *Environmental Conservation*. 1993, 20(2), 243-253.
- [35] Sharma, A. Limnological studies of Ban-Ganga and distributional pattern of stream bottom fauna. Ph.D. Thesis. University of Jammu, Jammu, 1999.
- [36] Hynes, H. B. N. The Invertebrate fauna at Welsh Mountain Stream. *Arch. Hydrobiol.*, 1961, 57, 344 – 388.
- [37] Akaahan T.J.A. Studies on Benthic Fauna as Bio Indicators of Pollution in River Benue at Makurdi, Benue State, Nigeria. *International Research Journal of Environment Sciences* 2014 3(7), 33-38.
- [38] Hussan Q.A. and Pandit A.K. Macroinvertebrates In Streams: A Review Of Some Ecological Factors, *Int. J. of Fish. and Aquaculture*, 2012 4(7), 114-123.