

# Production of Ethanol by Three Yeasts in Defined Media and Hydrolyzed Cladodes of *Opuntia ficus-indica* var. Atlixco

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**Abstract** Three yeast strains of *Opuntia ficus indica* cladodes were isolated, these strains were identified as *Candida intermedia*, *Zygosaccharomyces bailii* and *Saccharomyces paradoxus*. The yeasts were grown in synthetic media using as carbon source the main sugars detected in the acid hydrolysates of the cladodes: fructose, galactose, glucose and mannose. Different sugar consumption patterns were observed for each of the yeasts under study, being the main sugars consumed; glucose and fructose. Under these conditions, *Z. bailii* showed the highest biomass production with a growth rate of 0.146 h<sup>-1</sup>. However, *C. intermedia* was the yeast with the highest ethanol yield with 0.299 g of ethanol/g of biomass. When the ethanol production was evaluated in co-fermentation with the three strains, an increase in ethanol production of 300% (0.727 g of ethanol/g of biomass) was observed with a conversion efficiency of 28.9%. Subsequently, the *Opuntia* hydrolyzate was used as the base of ethanol production medium, the yeast that showed the best behaviors was *C. intermedia* with yields of 0.321 g of ethanol/g of biomass and conversion efficiency of 63.029%.

**Keywords** Cladodes, Hydrolysis, *Opuntia ficus-indica*, Yeast

## 1. Introduction

The genus *Opuntia* in Mexico is cultivated in 29 federative entities that produce about 723,000 tons in 12,500 hectares. For 2010 the area increased 5 times and the production of forage nopal was 208,492 tons [1]. Although its chemical composition is variable, the presence of mucilage and fibers are the main components that constitute the cladodes of *Opuntia* sp. The fibers is composed of a bilateral network of lignocellulosic residues that with age harden and provide a rigid constitution, increasing the thickness and intensifying the colour [2]. The lignocellulosic residues contain cellulose (C<sub>10</sub>H<sub>10</sub>O<sub>5</sub>)<sub>x</sub> and hemicellulose (C<sub>5</sub>H<sub>8</sub>O<sub>4</sub>)<sub>m</sub> which are bound to the lignin [C<sub>9</sub>H<sub>10</sub>O<sub>3</sub>(OCH<sub>3</sub>)<sub>0.9-1.7</sub>]<sub>n</sub> [3]. Since the conversion of cellulose and hemicellulose into sugar monomers such as 5 and 6 carbon carbohydrates is complicated, one of the main aspects to consider in the use of lignocellulosic residues for the production of bioethanol is the pre-treatment to destroy the cellulose matrix and dispose of fermentable sugars [4].

Starting from the pretreatment or treatment of the lignocellulosic material, sugars such as hexoses are obtained, which can be efficiently converted to bioethanol with yields between 0.4 and 0.5 g/g with productivities greater than 1 g/L h when using *Saccharomyces cerevisiae*. Similar productivities can be obtained from mesophilic microorganisms such as *Escherichia coli*, *Klebsiella oxytoca* y *Zymomonas mobilis* *Clostridium sporogenes*, *C. indolicus*, *C. sphnoides*, *Erwinia amilovora*, *Spirocheta aurantia*, *Streptococcus lactis*, *Spirocheta litorales* and *Spirocheta stenostrepta* [5].

However, *S. cerevisiae* is unable to ferment pentoses derived from hemicellulose such as xylose and arabinose [6]. The xylose is the most predominant pentose in hemicellulose, in addition, arabinose can constitute from 2-20% of the sugars in the hemicellulose depending on the agricultural residue in question [3]. Therefore, due to the high number of pentoses present in the lignocellulosic biomass, it is important to make efficient the use of these sugars by microorganisms to reduce production costs [6].

Among the microorganisms that are able to naturally degrade pentoses such as xylose, are some yeasts such as *Candida*, *Kluiveromyces*, *Pachysolen*, and *Pichia* [7]. In addition, it has been reported that xylose and arabinose can

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Published online at <http://journal.sapub.org/ijaf>

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be fermented by genetically modified microorganisms such as *S. cerevisiae*, which in addition to expressing enzymes for the degradation of xylose has some advantages, such as tolerance to bioethanol and other inhibitory compounds present in lignocellulosic hydrolysates [3, 8]. These inhibitory compounds can be: acetic acid, formic acid, levulinic acid, 5-hydroxymethyl furfural and furfural, the effects caused by these inhibitors depend on the biomass used and the conditions tested during the pre-treatment [9].

Simi, (2009) mentions that microorganisms grow efficiently in complex media, due to the presence of components that can be used as precursors in metabolic pathways. In the fermentative stage for the production of bioethanol, different types of nutrients can be used, many of agricultural origin such as molasses, bagasse of cane among others, which contain 45-55% of fermentable sugars like sucrose, Glucose, fructose, galactose and xylose [10]. The latter can be fermented at concentrations below 100 g/L; higher concentrations may lead to a decrease in productivity and performance of the microorganism. The effect is different for each type of microorganism being studied [11]. However, these lignocellulosic substrates may be deficient in some essential elements for growth, one of these elements is the nitrogen which is at low concentrations, making necessary the addition of a nitrogen source [12]. Similarly, sources of magnesium and phosphorus can be supplied in the form of salts along with vitamins such as biotin, thiamine and pantothenic acid, which must be supplied to the medium to increase the growth speed [12]. In the present work the capacity of three yeasts isolated from *Opuntia sp.* as fermenting microorganisms are evaluated, for the production of ethanol in both, defined medium and hydrolysates of *Opuntia ficus indica* var. Atlixco.

## 2. Materials and Methods

### 2.1. Raw Material

Twelve-month-old Cladodes of *Opuntia ficus indica*, var Atlixco, were cut in 2 cm cubes of cladodes were dried at 60°C for 72 h, then milled using a commercial grain mill. The obtained flour was stored in plastic bags and kept in a dry place and mill until its use.

### 2.2. Acid Hydrolysis of *Opuntia*

From the *Opuntia ficus indica* flour, acid pretreatment was performed in 500 mL Erlenmeyer flasks with a 400 mL working volume, adjusting the solids amount to 10% w/v and a 3% sulfuric acid concentration (v/v). The experimental units were maintained at 121°C and 1 atm pressure for 40 min. The obtained hydrolysates was neutralized with NaOH and centrifuged at 8000 rpm for 10 min. Subsequently, the filtrate was treated with a 0.45 µm membrane for ethanol production trials.

### 2.3. Microorganism, Media and Culture Conditions

Three strains of yeasts were previously isolated from cladodes of *Opuntia spp.* in the laboratory of agrobiotechnology, Polytechnic University of Pachuca, Mexico. The three strains identified as *Candida intermedia*, *Zygosaccharomyces bailii*, *Saccharomyces paradoxus* [13] and were grown in YPDA solid medium, formulated with yeast extract (10 g/L), peptone (20 g/L), glucose (20 g/L) agar (40 g/L) after sterilization at 120°C for 15 min (Lee et al., 2000) and incubated at 20°C. The inoculum of each yeast was prepared in liquid YPD medium supplemented with an antibiotic at 75 µg/mL, incubated for 24 h at 28°C. The culture of each yeast was centrifuged at 6000 x g for 5 min to separate the biomass then it was re-suspended in isotonic solution for use.

### 2.4. Fermentation Kinetics in Synthetic Medium

Yeast fermentations were performed in triplicate with a mixture of glucose, galactose, mannose, and fructose at a concentration of 20 g/L (5 g/L of each sugar) in Erlenmeyer flasks (125 mL) containing 100 mL of the fermentation medium composed of: 5 g (NH<sub>4</sub>)<sub>2</sub>HPO<sub>4</sub>, 0.75 g KH<sub>2</sub>PO<sub>4</sub>, 0.5 g K<sub>2</sub>HPO<sub>4</sub>, 0.2 g MgSO<sub>4</sub>·7H<sub>2</sub>O, 0.02 g CaCl<sub>2</sub>·2H<sub>2</sub>O, 0.1 g NaCl (Kuloyo et al., 2014), the flasks were inoculated with each microorganism by adjusting to an absorbance of 0.25 and a mixture of each of them in a 1:1 ratio. Each experimental unit was incubated on an orbital shaker at 150 rpm at 28°C for 4 days, samples of 1 mL were taken and were used to determine ethanol, reducing sugars and total sugars to each of them [14].

### 2.5. Fermentation Kinetics in *Opuntia ficus indica* Hydrolyzate

The fermentations were carried out in Erlenmeyer flasks (125 mL) containing 100 mL of the fermentation medium obtained by acid hydrolysis supplemented with (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>; K<sub>2</sub>HPO<sub>4</sub> and MgSO<sub>4</sub>. The flasks were inoculated with each microorganism by adjusting to an absorbance of 0.25, then incubated on an orbital shaker for 4 days sampling at regular intervals of time. The obtained samples were centrifuged at 13000 rpm for 5 min, the supernatant was collected and analysed for ethanol and residual sugar concentration, the residue was used for the determination of the biomass [15].

### 2.6. Determination of Reducing Sugars

The concentration of reducing sugars was determined using the dinitrosalicylic acid method described by Miller et al. (1959) and using glucose as standard.

### 2.7. Determination of Phenolic Compounds

The determination of total phenolics was carried out using the Folin-Ciocalteu method described by [17] using gallic acid as standard.

## 2.8. Quantification of Biomass<sup>†</sup>

Biomass was determined by spectrophotometry, after preparation of the calibration curve by measuring the absorbance at 600 nm [18].

## 2.9. Determination of Sugars

The concentration of glucose, galactose, mannose, and fructose and oligosaccharides in the hydrolyzate was determined by HPLC with 5  $\mu$ L automatic injection per sample, using a Rezex RCM Ca<sup>+</sup> monosaccharide column for the initial analysis and the Rezex RPM Pb<sup>+</sup> column for the Quantification and Rezex Oligosaccharides (Phenomenex), equipped with a temperature controlled oven at 80°C, the mobile phase was water HPLC grade at 0.6 mL/min and 0.3 mL/min respectively, quantification was performed prior elaboration of calibration curves for each sugar.

## 2.10. Determination of Ethanol

Ethanol determination was performed with a Trace 1310 gas chromatograph (Thermo Scientific) equipped with a Phenomenex ZB column (Phenomenex, Torrance, CA, USA) and a flame ionization detector using Helium carrier gas at 1.5 mL/min. The furnace was maintained at 40°C for 1 min, followed by a ramp at 25°C/min to 250°C with an isothermal period of 1 min, each injection was 1  $\mu$ L at a Split ratio of 66: 1. The temperature of the injector was 250°C. All the supernatants were filtered through a 0.2 mm membrane prior to chromatographic analysis.

# 3. Results and Discussion

## 3.1. Obtaining Hydrolysates

The hydrolysis process in cladode flour of *Opuntia sp.* Var Atlixco, with a solids loading of 10% w/v and sulfuric acid concentration of 3% v/v, with concentrations of both total reducing sugars (TRS) and phenolic compounds released (PHCR), At concentrations of 22.96 g/L and 1.63 g/L respectively, results equivalent to 27.7% of pretreatment yield, this low yield and concentration of TRS could be due, first to the degradation of sugars caused by the hydrolysis of oligosaccharides to compounds of low molecular weight and the subsequent degradation of the products obtained to furfural derivatives [19, 20] and second, to the solids loading in the acid pretreatment [21, 22]. In addition, the effect of a second and third hydrolysis was determined under the same conditions, with a decrease in the amount of TRS at 6 and 2 g/L for each subsequent hydrolysis stage, respectively, with an increase in the yield of the process up to 87.61%, Under the amount of solids used, [23] observed yields of up to 50% with 50 g / L of TRS for the lignocellulosic residue used. On the other hand, in a study conducted by Guevara-Figueroa et al. (2010) to determine the lyophilized phenolic compounds content of ten varieties of cactus, found that the phenolic amount present in the samples was from 2 mg/g to

20 mg/g. Additionally, [25] observed concentrations of phenolic compounds from 0.3 g/g to 0.9 g/g under drying conditions, conditions similar to those submitted to the *Opuntia ficus indica* variety studied in the present work.

In addition, the chromatographic analysis of *Opuntia ficus indica* hydrolysates showed the presence of oligosaccharides such as xylopentosa, xylotetroses, xylotriose, maltose and xylobiose (Table 1). Additionally, it was observed that this amount of both monosaccharides and oligosaccharides is variable depending on the pre-treatment conditions used. The presence of these oligosaccharides may be due mainly to the acid treatment, eliminating the insoluble hemicellulose from the surface of cellulose microfibrils giving soluble oligosaccharides [26].

## 3.2. Fermentation Kinetics and Sugars Assimilation in Defined Medium

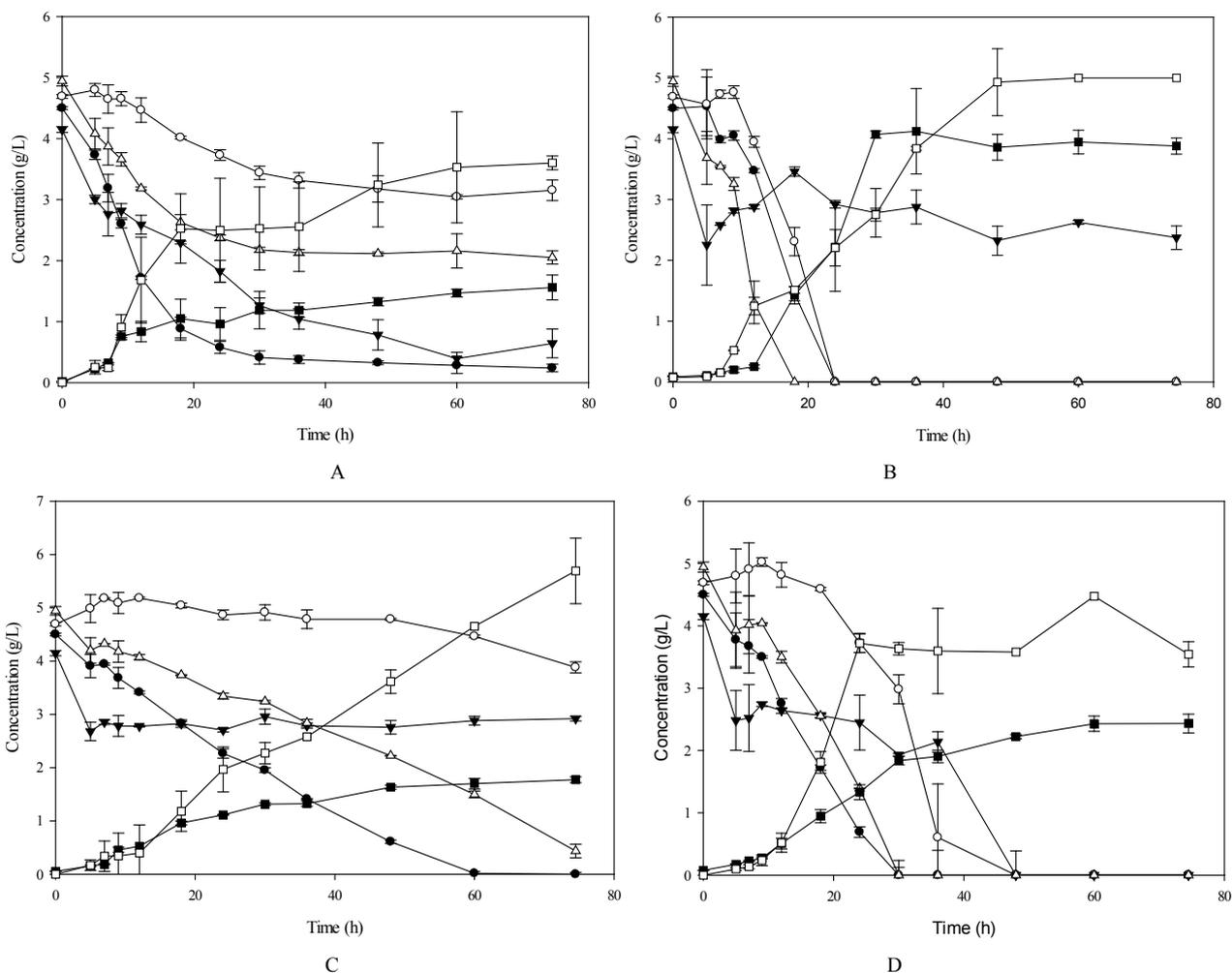
The previously isolated strains were evaluated for their ability to ferment and assimilate carbon sources such as glucose, mannose, galactose and fructose in a synthetic medium at a concentration of 20 g / L in a mixture of sugars. From the kinetics, it was observed that the *Z. bailii* strain, preferably consumed fructose, followed by glucose and mannose, during the first 25 hours of growth, in the same way, it was observed that there was a limitation in the consumption of galactose (Figure 1A). On the other hand, the strain *S. paradoxus* presented limitation to the mannose consumption, observing a preferential consumption of glucose, followed by galactose and fructose (Figure 1B), while *C. intermedia* showed a gradual consumption of sugars, being mainly glucose and fructose the most consumed sugars followed by mannose and galactose. (Figure 1C).

When evaluating the effect of isolated strains inoculated as a consortium (Fig. 1D), it was observed that the consumption of sugars as the only source of carbon was complete after 50 h of culture. The consumption of sugar was mainly of glucose, during the first 5 h, followed by fructose. These results suggest that *Z. bailii* and *C. intermedia* strains were the main microorganisms that metabolized these sugars, finally, the consumption of mannose and galactose was observed without having detected a significant increase in the production of ethanol, so it is very likely that these last two sugars are used as maintenance of the microorganism. Whereas the *S. paradoxus* strain had little assimilation of sugars, which indicated that this yeast requires special elements in the medium as growth factors, vitamins or culture conditions that allow both, the activation of the necessary enzymes as their assimilation and the obtaining of primary and secondary metabolites. [27] observed that under conditions of aerobic growth, both *Kluyveromyces marxianus* and *S. cerevisiae* preferentially consume glucose and fructose and when the conditions of the medium are changed to anaerobic, the microorganism is able to metabolize galactose, xylose, and arabinose, in addition, they observed that this consumption increases at the end of easy-to-assimilate carbon sources.

**Table 1.** Composition (g/L) of the hydrolyzate obtained from the factorial design for acid-solid treatments

Solids (% p/v)	Acid (% v/v)	Glc	Gal/Xyl	Ara/Frc	Xyl5	Xyl4	Xyl3	Mal	XylB
10	3	3.9	4.4	4.0	0.29	0.19	ND	2.40	0.90

Xyl5 = Xylopentose, Xyl4 = Xylotetrose, Xyl3 = Xylotriose, XylB = Xylobiose, Mal = Maltose



**Figure 1.** Sugar consumption kinetics for yeast strains; ● glucose; ○ manose; ▼ galactose; ▲ fructose; ■ Biomass; □ ethanol. (A) *Z. bailii*; (B) *S. paradoxus*; (C) *C. intermedia*; (D) co-culture

When evaluating the effect of the substrates for the production of ethanol, it was observed that *Z. bailii* strain presented higher biomass growth with 4.12 g/L compared to the other strains studied, which presented up to 1 g/L. It was determined that the growth rate was 0.146 h<sup>-1</sup> for the *Z. bailii* strain, 0.064 h<sup>-1</sup> for *S. paradoxus* strain, 0.053 h<sup>-1</sup> for *C. intermedia* strain and 0.070 h<sup>-1</sup> for the co-culture (*C. intermedia* and *S. paradoxus*). The results suggest that *Z. bailii* has the ability to metabolize carbon sources for biomass formation (Figure 1a). Nevertheless; in the co-culture, although the growth was slightly limited, a better production of ethanol was observed, obtaining a maximum of 5.5 g/L in a time of 60 h of culture (Figure 1d). These results suggest that consumption limitation of one type of substrate promotes the fermentation and production of ethanol in the medium. *S. cerevisiae* and *S. stipitis* have been subjected to co-culture processes for the production of

ethanol from a mixture of glucose and xylose, obtaining good yields in the production of ethanol under controlled conditions [28].

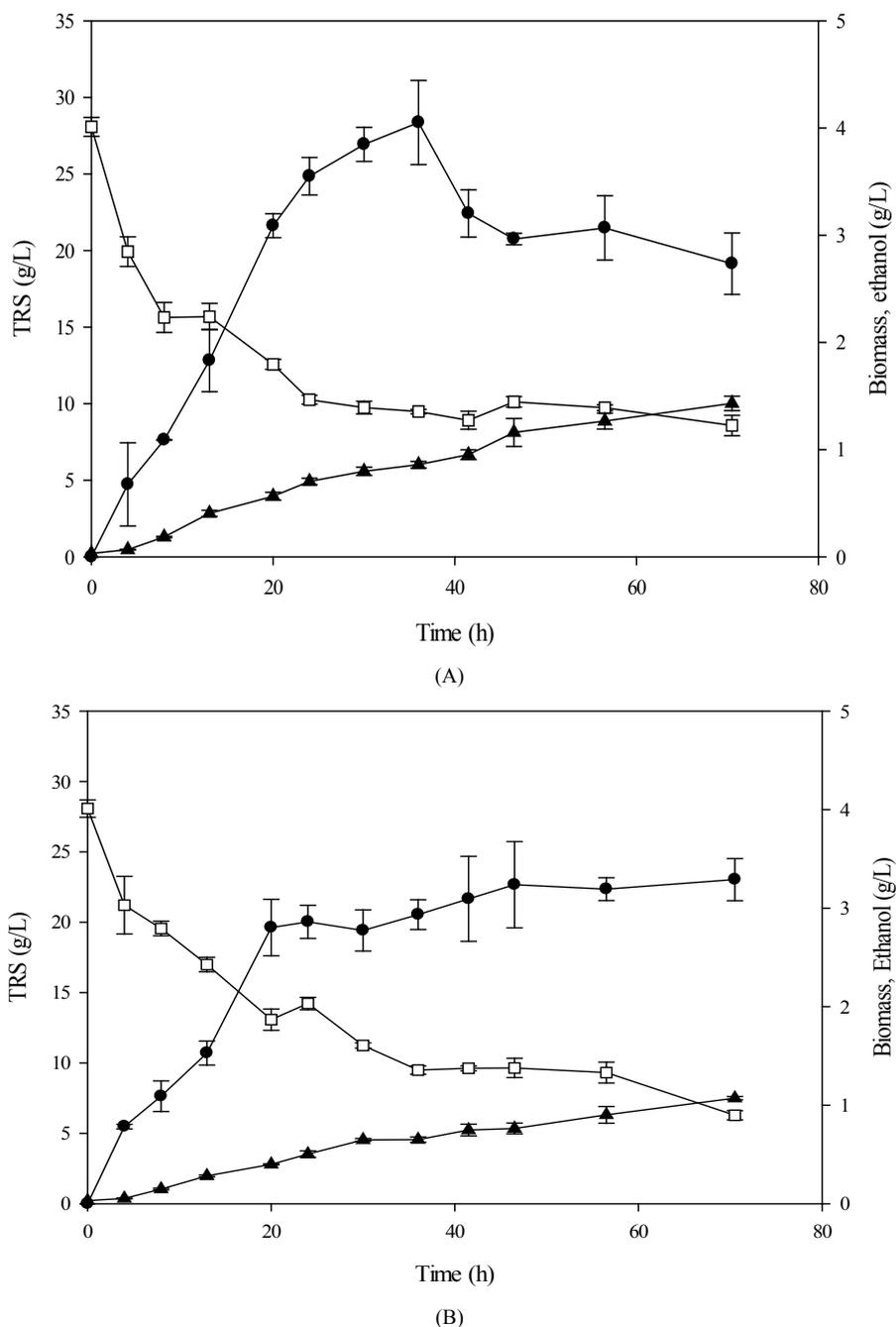
**Table 2.** Kinetic parameters obtained from cultures with the sugar mixture

	<i>Z. bailii</i>	<i>S. paradoxus</i>	<i>C. intermedia</i>	Co-culture
$\mu_{h^{-1}}$	0.146	0.064	0.053	0.070
$Y_{x/s}$ (g/g)	0.130	0.062	0.058	0.064
$Y_{p/s}$ (g/g)	0.045	0.018	0.015	0.028
$Y_{p/x}$ (g/g)	0.302	0.018	0.229	0.727
$Q_p$ (g/gh)	0.044	0.001	0.012	0.051
$\eta_{max}$	10.61 (36 h)	11.24 (48 h)	6.58 (60 h)	28.93 (60 h)

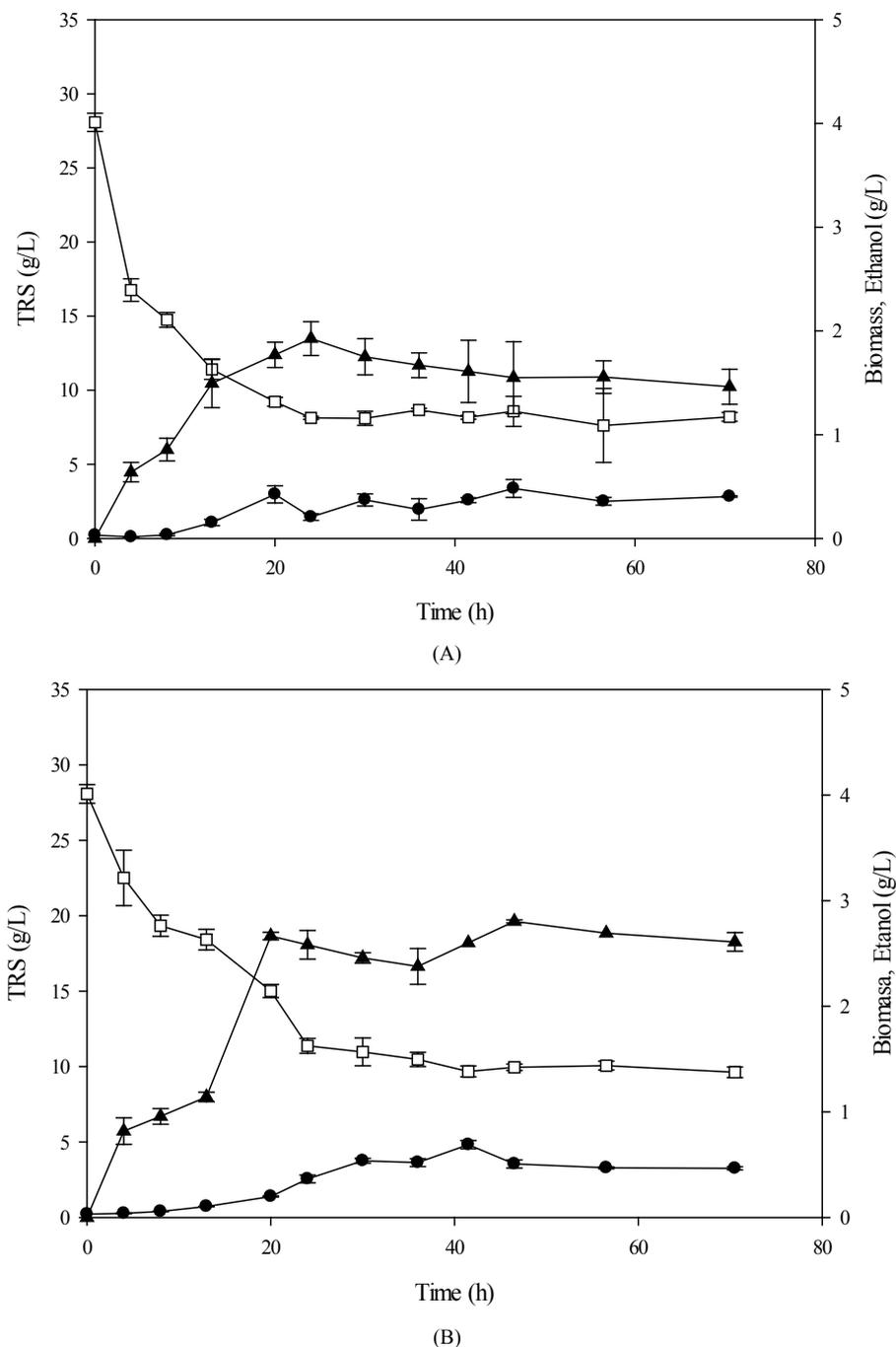
When quantifying the consumption of substrates as reducing sugars during the kinetics, it was observed that the co-culture had a  $Y_{p/x}$  yield of 0.72 g/g, which was greater than that obtained by the *Z. bailii* strain with  $Y_{p/x}$  of 0.3 g/g (Table 2). On the other hand; The  $Y_{p/s}$  yields were 0.028 and 0.045 g/g for the co-culture and *Z. bailii* strain respectively, with a maximum  $Y_{p/s}$  of 0.14 and 0.05 g/g respectively for these yeasts. In this sense, an important factor in the fermentation is the presence of a nitrogen source of easy assimilation in the culture medium. [6, 29, 30, 31, 32] use Yeast Extract as an additional nitrogen source, showing yields higher than 10 g/L.

The results obtained showed that the yeast mixture in a co-culture showed a conversion efficiency of 28.9% during the first 72 h. This result showed that the co-culture process allows a better efficiency in the conversion and transformation of the substrates for the formation of ethanol. [33] observed that the efficiency in the substrate conversion changes according to the type of carbon source being used and this efficiency ranges from 2 to 50% depending on the strain and the growing conditions.

### 3.3. Fermentation Kinetics in *Opuntia ficus indica* Hydrolyzate



**Figure 2.** Ethanol production kinetics for *C. intermedia* with physicochemical (A) and nutritional (B) factors. ▲ Biomass, ● Ethanol, ■ TRS (g/L)



**Figure 3.** Kinetics of ethanol production for *S. paradoxus*, in physicochemical (A) and nutrients (B) factors. ▲ Biomasa, ● Ethanol, ■ TRS (g/L)

**Table 3.** Kinetic parameters obtained from kinetics with yeasts *C. intermedia* and *S. paradoxus*

Parameter	<i>Candida intermedia</i>		<i>Saccharomyces paradoxus</i>	
	Physicochemical	Nutrients	Physicochemical	Nutrients
$\mu$ ( $h^{-1}$ )	0.199	0.120	0.264	0.103
$Y_{x/s}$ (g/g)	0.066	0.042	0.052	0.022
$Y_{p/s}$ (g/g)	0.321	0.248	0.157	0.248
$Y_{p/x}$ (g/g)	4.646	5.573	2.556	11.646
$Q_p$ (g/Lh)	0.923	0.666	0.676	1.197
$\eta$	63.029	48.565	30.740	48.694

Previous studies (data not shown) allowed to adjust several environmental conditions as well as the addition of nutrients for each of the yeasts studied. Conditions such as pH of 6.5, 150 rpm of stirring and 30°C were adjusted as conditions, allowing the maximum ethanol production for *C. intermedia* yeast, while for the addition of nutrients the  $\text{KH}_2\text{PO}_4$  salt was evaluated as the only additional nutrient at a concentration of 4 g/L. Figure 2B shows the kinetics performed during 72 h, where it was observed that the maximum amount of ethanol that could be obtained was 3.5 g/L and 4 g/L for the addition kinetics of P (phosphorus) as an additional nutrient and environmental conditions respectively (Figure 2A). These amounts were slightly compared to that obtained using a defined medium, this effect could be due to the process of adaptation of yeast cells to the medium used and the presence of various compounds in the medium such as phenolic compounds, which were 1.63 g/L in the hydrolysates, since the microorganism was grown initially in nutrient-rich medium (YPD).

In a study conducted by [34] with sugar cane leaves, under a treatment with sulfuric acid was able to obtain up to 800 mg/L of ethanol using *Klebsiella oxytoca* ATCC 68831 and *Pichia stipitis* ATCC 58376 as fermenting microorganisms.

Phenolic compounds have been reported as compounds capable of lowering ethanol yields and altering glycerol and acetate yields during fermentation of *S. cerevisiae* [21]; In addition, compounds such as hydroxymethyl furfural and furfural, occur during the process of chemical hydrolysis as an inevitable consequence of the degradation of sugars [19, 35] generating a negative effect on growth. In a study conducted by [36] evaluated the effect of inhibitory compounds, noting that furfural and HMF affect yeasts during the lag phase of *Saccharomyces cerevisiae* growth, however; The yeasts have adaptability during the stationary phase from 12 h and 24 h for HMF and furfural. This same effect was observed for the yeasts evaluated in the present study, observing a limitation of the growth during the first 24 h of culture, however; It was observed that at the end of the growing period, the amount of biomass increased slightly until obtaining about 1 g/L. The kinetics showed the ability of *Candida* and *S. paradoxus* to tolerate inhibitory compounds present in the culture medium obtained from *Opuntia* under an acid pretreatment; This possibly due to their ability to convert them into less harmful compounds under aerobic and anaerobic conditions [35].

On the other hand, 5 g/L of the nitrogen source added as  $(\text{NH}_4)_2\text{SO}_4$  and 0.5 g/L of Mg added as  $\text{MgSO}_4$  (Figure 3B) and anaerobic conditions, pH of 6.8 were evaluated for *S. paradoxus*, at a stirring speed of 200 rpm and 32°C (Figure 3A).

From the kinetics, it was observed that the yeast did not show a high capacity to produce ethanol, obtaining a maximum at 20 h of culture, with a production of 2.5 g/L of ethanol in the medium, although the consumption of reducing sugars was gradual. These results suggest that *S. paradoxus* has the ability to tolerate the compounds of the medium. Coupled with the presence of inhibitory

compounds in the culture medium as previously described, the amount of available sugars was an important factor in the fermentation process [37]; In addition to some divalent cations in the medium could directly affect the production of ethanol. These negative effects were visible on the *Z. bailii* yeast, which did not show growth on the hydrolysates, indicating that this microorganism has a minimum affinity on the substrate, making it unviable for the production of ethanol on the hydrolysates obtained from *Opuntia ficus indica*.

Table 3 shows the kinetic parameters obtained for the yeasts *C. intermedia* and *S. paradoxus*, for the conditions of physicochemical and nutrient parameters; of the obtained results could be observed that adding nutrients to the medium, the degree of conversion of the substrate to product was similar for the two evaluated yeasts with a 48% efficiency, whereas for the physicochemical parameters it had a better conversion efficiency with the yeast *C. intermedia*, whereas for *S. paradoxus* it was smaller with 63 and 45% respectively. On the other hand, yields were similar for the conditions evaluated except for the evaluation of physicochemical parameters, observing a  $Y_{p/s}$  of 0.32 g/g. In a study conducted by [38], with hydrolysates obtained by *Opuntia* with acid and enzymatic treatment, showed growth rates 0.4 and 0.7  $\text{h}^{-1}$  for *C. utilis* and *K. marxianus*, which makes it evident that the growth of the microorganisms used in this work was lower, with rates of Growth of approximately 0.2  $\text{h}^{-1}$  for the evaluation of physicochemical parameters and 0.1  $\text{h}^{-1}$  for the evaluation of nutritional parameters (Table 3), possibly coupled with inhibitory effects, low growth rates are directly related to the amount of available TRS as mentioned earlier. On the other hand, [39] reported similar results with growth rates  $< 0.3 \text{ h}^{-1}$  for microorganisms such as *S. cerevisiae*, *K. marxianus*, and *Pichia stipitis*, in addition; These same microorganisms showed yields ( $Y_{x/s}$ )  $< 0.1 \text{ g/g}$  resulting in yields of product as a function of the substrate ( $Y_{p/s}$ ) greater than 0.32 g/g for these microorganisms. These results suggest that the microorganisms used in the present work have the capacity to produce ethanol as a result of their des-assimilative metabolism but an increase in a number of sugars available for fermentation is necessary.

## 4. Conclusions

Based on the results obtained, it was possible to determine that the yeast *C. intermedia* is a good candidate to be used in the production of ethanol under different culture conditions, since ethanol production was observed in both synthetic medium and in hydrolysates obtained from *Opuntia ficus indica* as a culture medium, however, it is necessary to increase the release of fermentable sugars to improve yields, so as to allow the use of *Opuntia ficus indica* as a possible candidate suitable for the production of biofuels. During the hydrolysis of *Opuntia* flour, the loading of solids is essential to obtain a greater quantity of free sugars. The more solids,

the hydrolysis time should be longer but with the possibility of releasing more toxic compounds (such as furfurals). Therefore, it is necessary to complement the chemical hydrolysis with a biological hydrolysis or complement with other physicochemical methods to increase the release of sugars. It was observed in the hydrolysates, the presence of a great variety of oligosaccharides product of the hydrolysis of the hemicellulose and mucilage that can be an indicative of a partial hydrolysis. A limitation of the process is the chemical complexity of the hydrolysates which avoids high ethanol yields, hence the need to use microorganisms that are capable of fermenting most of the carbohydrates present in the hydrolyzate that would be possible with the use of microorganisms genetically modified.

## ACKNOWLEDGMENTS

This work was conducted thanks to the support received from Mexico's National Science and Technology Council (CONACyT) and Sectorial Fund SAGARPA-CONACyT, project No. 195157.

## REFERENCES

- [1] SIAP (Servicio de Información Agroalimentaria y Pesquera), 2011, SAGARPA. <http://www.siap.gob.mx>.
- [2] Aquino, G.L.V., Rodríguez, R.J., Méndez, R.A.M. and Hernández, A. E.S., 2012, Extracción y caracterización de fibra de nopal (*Opuntia ficus indica*). *Nat y Des*, 12(1), 48–62.
- [3] Balat, M., Balat, H. and Öz, C., 2008, Progress in bioethanol processing, *Progress in Energy and Combustion Science*, 34(5), 551–573.
- [4] Lipnizki, F., 2010, Membrane process opportunities and challenges in the bioethanol industry, *Desalination*. 250(3), 1067–1069.
- [5] Hernández N.M.T., 2007, Tendencias actuales en la producción de bioetanol, *Boletín Electrónico N° 08*. Facultad de Ingeniería-Universidad Rafael Landívar, (8), 1–17.
- [6] Kuloyo, O.O., 2012, Ethanol production by yeast fermentation of an *Opuntia ficus-indica* biomass hydrolysate. *Magister Scientiae Thesis*. University of Free State, Bloemfontein South Africa.
- [7] Silva, J. P. A., Mussatto, S. I., Roberto, I. C. and Teixeira, J. A., 2011. Ethanol production from xylose by *Pichia stipitis* NRRL Y-7124 in a stirred tank bioreactor. *Brazilian Journal of Chemical Engineering*, 28(1), 151–156.
- [8] Kaparaju, P., Serrano, M., Thomsen, A.B., Kongjan, P., and Angelidaki, L., 2009, Bioethanol, biohydrogen and biogas production from wheat straw in a biorefinery concept, *Bioresource Technology*. 100(9), 2562–2568.
- [9] Jönsson, L.J., Alriksson, B. and Nilvebrant, N., 2013, Bioconversion of lignocellulose: inhibitors and detoxification, *Biotechnology for biofuels*, 6(1), 16.
- [10] Simi, J.P., 2009, Process optimization for mass production of marine yeast *Candida* sp S27 and its nutritive characterization. Doctor in Philosophy in Microbiology Thesis. Cochin University of Science and Technology. India.
- [11] McMillan, J. D., 1993, Xylose Fermentation to Ethanol: A Review, National Renewable Energy Laboratory, (January), p. 45.
- [12] Gómez-Pastor, R., Pérez-Torrado, R., Garre, E., and Matallana, E., 2011, Recent Advances in Yeast Biomass Production, *Biomass: Detection, production and usage*, 201–222.
- [13] Pérez-Cadena R., Medina-Moreno, S.A., Martínez, A., Lizardi-Jiménez, M.A., Espinosa-Solares, T., Téllez-Jurado, A., 2018, Effect of concentration of salts in ethanol production from acid hydrolysis cladodes of *Opuntia ficus indica* Var. Atlitxco. *Revista Mexicana de Ingeniería Química*, 17(1), 349-364.
- [14] Sreenath, H. and Jeffreis, T., 2000, Production of ethanol from wood hydrolysate by yeasts. *Bioresource Technology*, 72, 253–260.
- [15] Maruthai, K., Thangavelu, V. and Kanagasabai, M., 2012, Statistical screening of medium components on ethanol production from cashew apple juice using *Saccharomyces diastolicus*. *Int J Chem Biol Engg*, 6(6), 108–111.
- [16] Miller, G.L., 1959, Use of Dinitrosalicylic Acid Reagent for Determination of Reducing Sugar, *Analytical Chemistry*, 31(3), 426–428.
- [17] De Ascensao, A.R.F.D.C. and Dubery, I.A., 2003, Soluble and wall-bound phenolics and phenolic polymers in *Musa acuminata* roots exposed to elicitors from *Fusarium oxysporum* f.sp. *cubense*. *Phytochemistry*, 63(6), 679–686.
- [18] Xavier, A.M.R.B., Correia, M.F., Pereira, S.R. and Evtuguin, D.V., 2010, Second-generation bioethanol from eucalypt sulphite spent liquor. *Bioresource Technology*, 101(8), 2755–2761.
- [19] Behera, S., Arora, R., Nandhagopal, N. and Kumar, S., 2014, Importance of chemical pretreatment for bioconversion of lignocellulosic biomass. *Renewable and Sustainable Energy Reviews*, 36, 91–106.
- [20] Parajó, J. C., Garrote, G., Cruz, J. and Dominguez, H., 2004, Production of xylooligosaccharides by autohydrolysis of lignocellulosic materials. *Trends in Food Science & Technology*, 15(3–4), 115–120.
- [21] Adeboye, P. T., Bettiga, M. and Olsson, L., 2014, The chemical nature of phenolic compounds determines their toxicity and induces distinct physiological responses in *Saccharomyces cerevisiae* in lignocellulose hydrolysates. *AMB Express*, 4, 46.
- [22] Qin, L., Li, W.-C., Liu, L., Zhu, J.-Q., Li, X., Li, B.-Z. and Yuan, Y.J., 2016, Inhibition of lignin-derived phenolic compounds to cellulase. *Biotechnology for Biofuels*, 9(1), 70.
- [23] Idrees, M., Adnan, A., Bokhari, S. A. and Qureshi, F.A., 2014, Production of fermentable sugars by combined chemo-enzymatic hydrolysis of cellulosic material for bioethanol production. *Brazilian Journal of Chemical Engineering*, 31(2), 355–363.
- [24] Guevara-Figueroa, T., Jiménez-Islas, H., Reyes-Escogido, M.

- L., Mortensen, A. G., Laursen, B. B., Lin, L.W., Barba de la Rosa, A.P., 2010, Proximate composition, phenolic acids, and flavonoids characterization of commercial and wild nopal (*Opuntia spp.*). *Journal of Food Composition and Analysis*, 23(6), 525–532.
- [25] Santos-zea, L., Guti, J. A., Serna-saldivar, S. O., Monterrey, D., Eugenio, A. and Sada, G., 2011, Comparative Analyses of Total Phenols, Antioxidant Activity, and Flavonol Glycoside Profile of Cladode Flours from Different Varieties of *Opuntia* spp. *Journal of Agricultural and Food Chemistry*, 59, 7054–7061.
- [26] Qing, Q., Li, H., Kumar, R. and Wyman, C. E., 2013, Xylooligosaccharides Production, Quantification, and Characterization in Context of Lignocellulosic Biomass Pretreatment. In *Aqueous Pretreatment of Plant Biomass for Biological and Chemical Conversion to Fuels and Chemicals* (pp. 391–415).
- [27] Kuloyo, O. O., du Preez, J. C., García-Aparicio, M. del P., Kilian, S. G., Steyn, L. and Görgens, J., 2014, *Opuntia ficus-indica* cladodes as feedstock for ethanol production by *Kluyveromyces marxianus* and *Saccharomyces cerevisiae*. *World Journal of Microbiology and Biotechnology*, 30(12), 3173–3183.
- [28] Hanly, T. J. and Henson, M. A., 2013, Dynamic metabolic modeling of a microaerobic yeast co-culture: predicting and optimizing ethanol production from glucose/xylose mixtures. *Biotechnology for Biofuels*, 6(1), 44.
- [29] Blomqvist, J., South, E., Tiukova, L., Momeni, M. H., Hansson, H., Ståhlberg, J., Passoth, V., 2011, Fermentation of lignocellulosic hydrolysate by the alternative industrial ethanol yeast *Dekkera bruxellensis*. *Letters in Applied Microbiology*, 53(1), 73–78.
- [30] Bravo, V., Camacho, F., Sánchez, S. and Castro, E., 1995, Influence of the concentrations of D-xylose and yeast extract on ethanol production by *Pachysolen tannophilus*. *Journal of Fermentation and Bioengineering*, 79(6), 566–571.
- [31] Kahr, H., Helmberger, S. and Jäger, A.G., 2011, Yeast Adaptation on the Substrate Straw, *World Renewable Energy Congress*, 492–499.
- [32] Lee, T., Kim, M.-D., Kim, K., Park, K., Ryu, Y. and Seo, J., 2000, *Pichia stipitis*. *Biotechnology and Bioprocess Engineering*, 5(1), 27–31.
- [33] Mussatto, S.I., Machado, E.M.S., Carneiro, L. M. and Teixeira, J.A., 2012, Sugars metabolism and ethanol production by different yeast strains from coffee industry wastes hydrolysates. *Applied Energy*, 92, 763–768.
- [34] Shields, S. and Boopathy, R., 2011, Ethanol production from lignocellulosic biomass of energy cane. *International Biodeterioration & Biodegradation*, 65(1), 142–146.
- [35] Parawira, W. and Tekere, M., 2011, Biotechnological strategies to overcome inhibitors in lignocellulose hydrolysates for ethanol production: review. *Critical Reviews in Biotechnology*, 31(1), 20–31.
- [36] Tian, S., Zhu, J. and Yang, X., 2011, Evaluation of an adapted inhibitor-tolerant yeast strain for ethanol production from combined hydrolysate of softwood. *Applied Energy*, 88(5), 1792–1796.
- [37] Kurian, J.K., Minu, A.K., Banerji, A. and Kishore, V.V.N., 2010, Bioconversion of hemicellulose hydrolysate of sweet sorghum bagasse to ethanol by using *Pichia stipitis* ncim 3497 and *Debaryomyces hansenii* sp. *BioResources*, 5(4), 2404–2416.
- [38] Akanni, G.B., du Preez, J.C., Steyn, L. and Kilian, S.G., 2015, Protein enrichment of an *Opuntia ficus-indica* cladode hydrolysate by cultivation of *Candida utilis* and *Kluyveromyces marxianus*. *Journal of the Science of Food and Agriculture*, 95(5), 1094–1102.
- [39] Rouhollah, H., Iraj, N., Giti, E. and Sorah, A., 2007, Mixed sugar fermentation by *Pichia stipitis*, *Sacharomyces cerevisiae*, and an isolated xylose fermenting *Kluyveromyces marxianus* and their cocultures. *African Journal of Biotechnology*, 6(9), 1110–1114.