

# Study of Antibacterial Effects of a Self-Standing Agar Based Film Incorporated with ZnO

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**Abstract** An agar based film was prepared and incorporated with an antibacterial agent, ZnO (Zinc Oxide). ZnO was prepared by a reflux method and the product was characterized by x-ray diffraction (XRD) and scanning electron microscopy (SEM). Synthesis of the agar based film and incorporation of the ZnO particles in the film was performed in a single pot method. The dispersion of ZnO particles in the films was revealed by SEM. A qualitative and quantitative study of the antibacterial activity of the films was carried out, after inoculation with the bacteria *Escherichia coli* and *Staphylococcus aureus*. Visual results indicate that the synthesized films inhibit the growth of the bacteria inoculated. The optimum effective concentration of ZnO /film was found by the Japanese Industrial Standard JIS Z2801:2000 method to be 10mg ZnO/film. According to the same standard method, the antibacterial efficacy (R) of the films against *E. coli* and *S.aureus* was calculated as >2.0. Bacteriological tests performed on films, containing 5, 10 and 15 mg ZnO/film, show that the films exhibit a bactericidal effect at the concentrations used. The above results indicate that agar films incorporated with ZnO, are promising candidates in the packaging sector as antibacterial food packaging materials, to increase the safety of food.

**Keywords** Antibacterial Agents, ZnO, Self-standing Film, Food Packaging Materials

## 1. Introduction

A recent subject of intense studies is the integration of antibacterial properties in films. These films could act as catalyst supports, separation membranes and implantable drug-release materials where powders and supported films cannot be used. Some examples of these studies are: DNA Lipid films, having potential dental applications[1], and DNA lipid complex films exhibiting antifungal properties[2]. Metal and metal oxide nanoparticle embedded PDMS or polymer freestanding multifunctional films exhibiting antibacterial properties were also investigated[3-6]. Food industry is also an area in which self-standing films are becoming significantly important as protective packaging. Antibacterial films act upon the food surface by contact, helping thus maintain better quality of food and increased shelf life[7-12]. Recently, natural (mostly edible) polymers have been used in the synthesis of films, especially when related to films used in food packaging[13]. Starches, because of the film forming characteristics of amylase have long been used in protecting foods from moisture and oxygen invasion, as well as, acting as carriers for

antibacterial agents[14].

Starch has also been used to form crosslinked films with polyvinyl alcohol PVA[15]. Chitosan and polyvinyl pyrrolidone were also used as polymeric networks in biomedical applications[16]. Films based on biopolymers from marine sources like alginates are also common[17]. Agar-agar has similarly been used as a polymeric matrix for antibacterial films[18]. Usage of plasticizers tends to increase the tensile strength and elongation modulus of the film, and render it more stable compared to films consisting only of agar[19].

In the present work an inorganic-organic hybrid was developed from ZnO powders. ZnO is an inorganic material of established antibacterial properties, in addition to its safety regarding human health[20, 21]. It has been used in antimicrobial fabrics[22, 23], to coat paper[24] as quantum dots[25], as well as ZnO films[26]. ZnO was dispersed in an agar-agar matrix to form a self-standing film. Plasticizers were used and the films were stable for a period of 6 months at a room temperature of 23 °C. Their antibacterial performance was evaluated qualitatively and quantitatively using *E.coli* and *S.aureus*, according to a modified JIS Z2801:2000 industrial Standard[27].

## 2. Materials and Methods

### 2.1. Synthesis of ZnO

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ZnO was synthesized using synthesis grade hexamethylenetetramine ( $C_6H_{12}N_4$ ), purchased from Merck and used as received. In a typical process, an aqueous suspension containing 50 mmol of Zn and 200 mmol  $C_6H_{12}N_4$  was refluxed for 24 h at 90°C, under  $N_2$ . The resulting white solid was filtered and washed several times with water. After drying overnight at 100°C, the crystal structure of the powder was investigated by X-ray powder diffraction (XRD, Bruker D2 Phaser) and the morphology was observed with a scanning electron microscope (SEM, Zeiss Evo). The particle size was calculated by the Debye-Scherrer Formula.

## 2.2. Synthesis of Self-Standing Film

Agar ( $C_{12}H_{18}O_9$ )<sub>x</sub> Mr3000-9000 (microbiology grade) used in this study was purchased from Fluka and polyvinylalcohol, [-CH<sub>2</sub>CH(OH)-]<sub>n</sub> (av  $M_w$  85,000-124,000) from Alfa Aesar. Reagent grade sucrose and glycerol were purchased from Aldrich.

About 0.20 g of agar-agar was dissolved in 15 ml deionized water with constant stirring, at 90°C until a clear solution was obtained. To this solution was added, a well blended and ground mixture of polyvinyl alcohol (PVA) and glucose. Then, glycerol was added as a plasticizer. The ratio of (Agar: PVA: Glucose: glycerol) was 1:1:0.5:0.5 by weight. Heating with continuous stirring was required until a clear solution was obtained. A suspension of ZnO powder dispersed in water (5ml) under ultrasonic treatment was added at this stage. The obtained suspension was heated (at 90°C) with stirring for another 10 min. The resulting viscous suspension was sonicated for 5 min to get rid of air bubbles and then was poured on polyethylene petri dishes (9.27±0.1 cm diameter) and let to dry in air.

Agar-agar films were also prepared in the same way without the addition of the ZnO suspension and were used as references or blanks in the tests. Different concentrations of ZnO were used, ranging from 5 to 35 mg (for films with a diameter of 9.27±0.1 cm) to find the minimum effective antibacterial concentration of ZnO.

## 2.3. Bacterial Cultures

Two species of bacteria, *S.aureus* ATCC 25923 and *E.coli* ATCC 8769, were used as model bacteria in this study. Nutrient broth, Mueller hinton agar media were used as sources for culturing *E.coli* and *S.aureus*, at 37°C in an incubator for 24 hours. The samples were then adjusted according to McFarland 0.5 standard and the necessary dilutions were made to obtain  $2.5 \times 10^5$  cfu/ml. All materials were sterilized in an autoclave before the experiments

## 2.4. Antibacterial Testing

Antibacterial properties testing for the ZnO containing self-standing films were carried out both qualitatively and quantitatively. For the qualitative test, the films on the petri-dishes were spray coated, evenly, with a bacterial solution on the surface and incubated at 35°C overnight. The qualitative test results were determined visually by

enumeration.

For the JIS Z 2801:2000 tests, a cell suspension of the bacterial strain, was prepared in 1/500 nutrient broth. The test surfaces were prepared as follows: each film (ZnO incorporated films and blanks) was placed in a sterilized petri dish making the test surface. 0.4 ml of the inoculum was instilled on each film. The instilled test inoculum was covered with sterile polyethylene. The polyethylene was pressed so that the test inoculum spread over the film. Three replicate samples per species of the ZnO containing film and 6 replicate blank samples were prepared for each ZnO concentration. A and B represent the blank films, and C, the ZnO containing film.

Group A films (blanks), were immediately diluted and underwent plate culture. The number of viable cells was determined by counting and averaging the number of colonies that grew in each Petri dish. The films corresponding to groups B (blanks) and C (ZnO containing films) were incubated and tested as described below.

The incubation times for the inoculated test surfaces were 15, 30, 60 min and 24 h, for samples containing 5, 10, 15 mg of ZnO/film. The surfaces were incubated at 35°C under aerobic conditions and 90% humidity. After incubation, the test bacteria were washed by addition of 10 ml nutrient broth. The samples were diluted and the viable cell count of bacteria was performed by the agar plate culture method. An average number of viable cells were obtained after incubation by averaging the numbers in the three replicate cells. The blanks were also processed in the same manner to provide base-line data.

The tests were performed using *S.aureus* ATCC 25923 and *E.coli* ATCC 8769. (R) The antibacterial activity was calculated with the values obtained from the above tests performed on the films.

A standard protocol was employed to determine bactericidal effect of ZnO. A suspension of ZnO (6.5 mg/ml in 1ml isotonic water) was prepared. The bacteria under inspection were adjusted according to McFarland 0.5. Then, ZnO suspension was added to it. In this way, a series of samples containing 6.5, 4.0 and 2.0 mg/ml concentrations of ZnO were prepared. The same was repeated for blanks in test tubes containing only the broth and no ZnO. The tubes were incubated overnight at 35°C. All tests were performed as triplicates. Bacterial growth was determined visually.

To observe the bactericidal effect of ZnO, the test tubes containing the samples in which no bacterial growth was observed were used. They were inoculated once more using Mueller hinton agar plates and incubated overnight at 35°C. Then, the viable cell count of bacteria was performed as described above.

# 3. Results and Discussion

## 3.1. Characterization of ZnO

The XRD pattern of ZnO is given in Fig. 1. The XRD

pattern revealed the wurtzite structure of ZnO particles. The sharp and intense peaks show the high degree of crystallization of the oxide. The particle size was calculated using the Debye-Scherrer equation as 200 nm[28].

As seen in the SEM images of the ZnO powder (Figure 2), the synthesis method resulted in approximately equilateral powders.

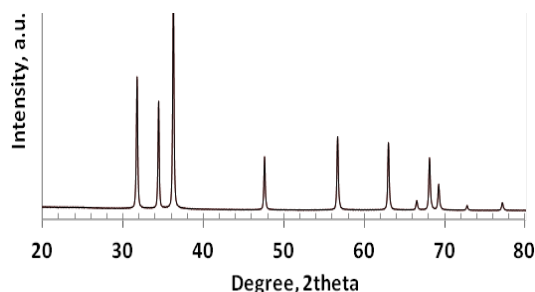


Figure 1. XRD pattern of ZnO particles

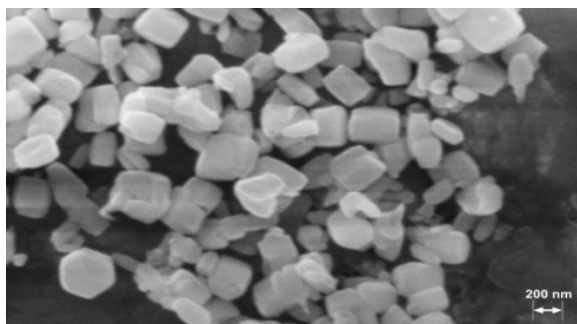


Figure 2. SEM image of the synthesized ZnO particles before film preparation

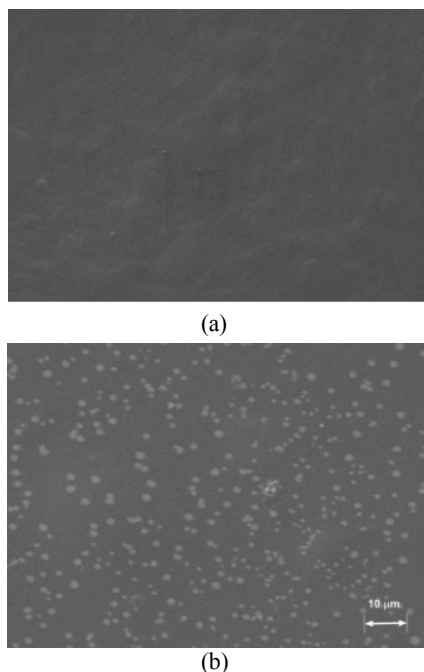


Figure 3. SEM image of the blank film (a) and of the ZnO containing film (b)

After the synthesis, ZnO particles were embedded in an agar based film as described in the experimental part. The ZnO suspension was poured into the agar-polymer solution right after ultrasonic dispersion of the ZnO in water. The thickness of the film could be controlled by the amount of solution applied on the petri dish. All films were flexible and opaque as a result of the presence of ZnO particles. After the films were dried they could be easily removed from the petri dishes. The SEM image of the blank film and ZnO containing films (10mg ZnO/film, for films with a diameter of  $9.27 \pm 0.1$  cm) are shown in Fig. 3.(a and b), at magnification 1.65 K. Direct comparison of the images reveals ZnO particles have been uniformly dispersed.

### 3.2. Physical Properties of the Film

The thickness of the synthesized films was measured after 24hrs drying in the air. The measurements were done on 6 samples using a micrometer. The average thickness was calculated as:  $125 \pm 1.8$  mm.

The average volume of the film was calculated using the thickness value in  $V = \pi r^2 t$ , and was found to be:  $8.4 \pm 0.7$  ml. The average concentration/film was calculated as:

$2.1 \pm 0.7$ mg ZnO/ml	for a 5mg ZnO suspension
$4.2 \pm 0.7$ mg ZnO/ml	for a 10mg ZnO suspension
$6.3 \pm 0.7$ mg ZnO/ml	for a 15mg ZnO suspension

### 3.3. Qualitative Evaluation of Antibacterial Properties

As revealed in Figures 4 and 5, for the case of *Escherichia coli* and *Staphylococcus aureus* respectively, the blank films are completely covered with the bacterial colonies after 24 h while the ZnO containing films are not. The tests were done in triplicate, all cases showed a similar situation.

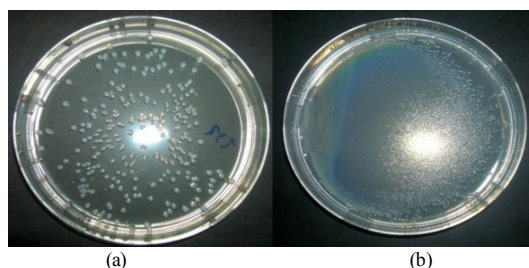


Figure 4. *E. coli* colonies completely cover the blank film (b) after 24 h as compared to the ZnO containing film (a)

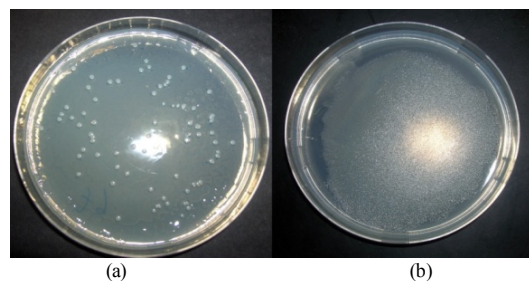


Figure 5. *S. aureus* completely covers the blank film (b) after 24 h as compared to the ZnO containing film (a)

**Table 1.** Antibacterial Activity Test of the ZnO containing Agar films against *E.coli* and *S. aureus*

<i>E.coli</i>								
sample	15min cfu/ml	% reduction in viability (mean±sd)	30 min cfu/ml	% reduction in viability (mean±sd)	60 min cfu/ml	% reduction in viability (mean±sd)	24h cfu/ml	% reduction in viability (mean±sd)
Blank	$1.0 \times 10^4$	No reduction observed	$1.0 \times 10^4$	No reduction observed	$1.0 \times 10^4$	No reduction observed	$1.0 \times 10^4$	0.0
2.1±0.7 mg ZnO/ml	$5.0 \times 10^2$	95.0±0,6					20.0	99.0±0,06
4.2±0.7 mg ZnO/ml	$2.0 \times 10^2$	98.5±1,0	80.0	99.3±0,25	10.0	99.9±0,06	0.00	100±0,05
6.3±0.7 mg ZnO/ml	$2.0 \times 10^2$	97.0±1,7	100.0	99.0±0,11	10.0	99.9±0,05	0.00	100±0,05

<i>S.aureus</i>								
sample	15min cfu/ml	% reduction in viability (mean±sd)	30 min cfu/ml	% reduction in viability (mean±sd)	60 min cfu/ml	% reduction in viability (mean±sd)	24h cfu/ml	% reduction in viability (mean±sd)
Blank	$1.0 \times 10^4$	No reduction observed	$1.0 \times 10^4$	No reduction observed	$1.0 \times 10^4$	No reduction observed	$1.0 \times 10^4$	No reduction observed
2.1±0.7 mg ZnO/ml	$1.0 \times 10^3$	90.0±1,1					10.0	99.9±0,06
4.2±0.7 mg ZnO/ml	60.0	99.5±0,06	50.0	99.5±0,06	10.0	99.9±0,06	0.00	100±0,05
6.3±0.7 mg ZnO/ml	50.0	99.5±0,06	50.0	99.5±0,06	10.0	99.8±0,06	0.00	100±0,05

All tests performed in the dark

2.1±0.7 mg ZnO/ml = 5mg ZnO suspension/film  
 4.2±0.7 mg ZnO/ml = 10mg ZnO suspension/film  
 6.3±0.7 mg ZnO/ml = 15mg ZnO suspension/film

### 3.4. Quantitative Evaluation of Antibacterial Properties

The antibacterial activity of the ZnO containing films, were evaluated for different concentrations of ZnO, for different time periods. Films containing 35 and 35 mg of ZnO/film were very opaque and brittle. Tests were not performed on them because of the brittleness, which makes the film difficult to handle. Besides, excellent results were obtained with lower concentrations of ZnO/film. From the results listed in Table 1, it can be seen that 10 mg of ZnO/film (4.2±0.7 mg/ml) is the optimum effective concentration to be used with both strains.

### 3.5. Calculation of the Value of Antibacterial Efficacy

$$R = [\log(B/A) - \log(C/A)] = [\log(B/C)] \quad [27].$$

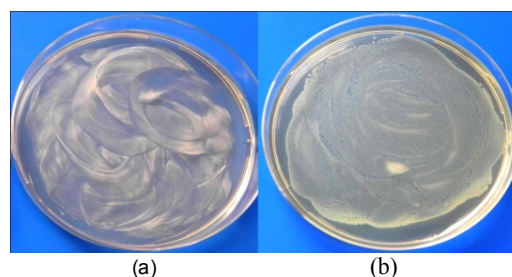
Here R is the value of antibacterial efficacy. This value shows the difference in the logarithmic value of viable cell counts between antibacterial products and untreated products after inoculation and incubation of bacteria. A is the average of the number of viable bacteria on blank films (group A). B represents the average number of viable bacteria on incubated blank films (group B) and C the average number of viable bacteria on incubated ZnO containing films (group C).

The R value was calculated for the inoculation time and minimum ZnO concentration / film for which the test was effective (4.2±0.7 mg ZnO/ml or 10mg ZnO suspension/film, incubated for 15 min).

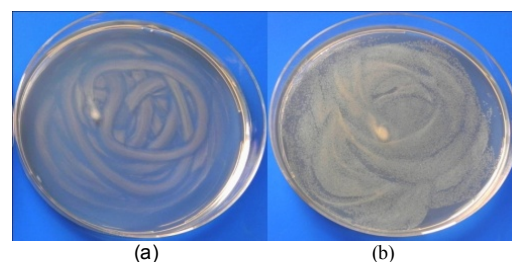
R obtained by the testing methods of this standart should not be less than 2.0 for the antibacterial efficacy of antibacterial products. In this study, R for *E. coli* and *S. aureus* was found to be > 2. Disk diffusion tests were not performed as the ZnO dispersion diffuses poorly.

### 3.6. Bactericidal Effect

Samples containing 6.3, 4.2 and 2.1 mg/ml of ZnO were used in this test. After inoculation and incubation (section 2.4), the samples on which bacterial growth was inhibited where inoculated and incubated again. No bacterial activity was observed on incubation of the colonies inoculated on the ZnO samples at any of the ZnO concentrations used. It can be concluded that ZnO effectively inhibits bacterial growth and the effect is bactericidal. The same results were obtained in the case of *E. Coli* and *S. aureus*. (Figures 6 and 7)



**Figure 6.** State of *E.coli* culture on solid agar after overnight incubation at 35°C. (a) ZnO containing medium (b) Blank



**Figure 7.** State of *S.aureus* culture on solid agar after overnight incubation at 35°C. (a) ZnO containing medium (b) Blank

### 3.7. Discussion of the Antibacterial Mechanisms

A number of mechanisms have been proposed to interpret the antibacterial behavior of ZnO. Swinkels, *et al.*, believe that the antibacterial effect is due to the zinc aqua complex  $\text{Zn}(\text{H}_2\text{O})$  attaining maximum acidity at pH 7, making ionization of water  $\text{H}_2\text{O}$  to hydroxyl ions  $\text{OH}^-$  possible[29], even in the absence of light. Hydroxyl ions would then yield, hydroxyl radicals (dissolved) which would destroy bacterial cell[30]. But there is no comprehensive report concerning this mechanism, to our knowledge. Xhang *et al.*, showed by electrochemical analysis that the antibacterial effect of ZnO, or the bacteria membrane damage may be partly due to the direct interaction of ZnO nanoparticles and bacteria membrane wall, in addition to active oxygen species generation[31].

Sawai *et al.*, attributed the antibacterial effect of ZnO to the production of  $\text{H}_2\text{O}_2$  from a ZnO slurry. They investigated this property using four kinds of antibiotics. Changes in sensitivity of the bacteria used, to the antibiotics, suggested that  $\text{H}_2\text{O}_2$  was a factor in the antibacterial activity of the ZnO powder slurry[32]. In another study by Sawai *et al.*, hydrogen peroxide generated from ZnO, MgO and CaO slurries was detected using oxygen electrode and chemiluminescence analysis[33]. The amount of  $\text{H}_2\text{O}_2$  generated was found to be proportional to the antibacterial activities of the above oxides. Furthermore in a study performed on activated carbon spheres containing ZnO, the generation of  $\text{H}_2\text{O}_2$  was observed in physiological saline dispersed with activated carbons. The antibacterial activity was found to be caused by the generation of  $\text{H}_2\text{O}_2$  from ZnO[34].

## 4. Conclusions

A simple method was developed to obtain stable self-standing films as organic-inorganic hybrids of ZnO incorporated in an agar based matrix. The film components are all used as food additives and represent no health hazard [35]. The ZnO containing film exhibited antibacterial effect against *E.coli* and *S. aureus* at 5,10 and 15 mg ZnO/film, with a bactericidal effect, in comparison with films containing no ZnO. The antibacterial activity of the film were attributed to ZnO. This antibacterial effect is believed to be due to the generation of hydrogen peroxide from the surfaces of ZnO particles (section 3.7). The inhibitory effect of  $\text{H}_2\text{O}_2$  is due to the fact that most living cells contain the enzyme catalase which attacks  $\text{H}_2\text{O}_2$  and converts it to water ( $\text{H}_2\text{O}$ ) and oxygen ( $\text{O}_2$ ). Oxygen atoms attract electrons from the cell walls of bacteria, thus destroying the basic molecular structure of the cell proteins and the layer of lipids on the cell surface. The oxidized cells walls become damaged or even completely break apart[36].

For practical applications, further studies would be necessary for measuring and optimizing mechanical properties such as tensile strength and elongation, as well as moisture absorbance and resistance to water disintegration .

The antibacterial effects of ZnO against other food pathogens , which may cause health problems or food spoilage that shorten the shelf life of foods should also be investigated.

10mg ZnO/film was found to be the optimum concentration to be used in food packaging , as it resulted in 100% reduction in viability after 15min contact (section 3.4).

No irradiation is needed to activate the antibacterial agent: for example, it would inhibit bacterial growth on food, in dark storage areas as well as when food is on display under a light source.

From the above, it could be concluded that the synthesized agar based films incorporated with ZnO, could possibly serve as antibacterial food packaging material for minimizing the risk of contamination and growth of bacteria, such as *E. coli* and *S. aureus*.

## REFERENCES

- [1] Fukushima, T.e.a., Intercalation behavior and tensile strength of DNA-lipid films for the dental application. *Biomaterials*, 2004. 24: p. 5991-7.
- [2] Inoue, I.e.a., Antifungal activity of DNA-lipid complexes and DNA-lipid films against *Candida* species. *Journal of Biomedical Materials Research Part A*, 2005. 76A(1): p. 1265-132.
- [3] Goyal, A.K., A.; *et al.*, In Situ Synthesis of Metal nanoparticle Embedded Free Standing Multifunctional PDMS Films. *Macromolecular Rapid Commun*, 2009. 30: p. 1116-1122.
- [4] Porel, S.R., D.; *etr. al.*, Polymer Thin Film with in situ synthesized silver nanoparticles as a potent reusable bactericide. *Current Science*, 2011. 101(7): p. 927-934.
- [5] Chawengkijwanich, C.H., Y., Development of TiO<sub>2</sub> powder-coated food packaging film and its ability to inactivate *Escherichia coli* in vitro and in actual tests. *International Journal of Food Microbiology*, 2008. 123: p. 288-292.
- [6] Xihong, L.X., Yage; Jiang, Yunhong; Ding, Yulong; Weili, Li, Antimicrobial activities of ZnO powder coated PVC film to inactivate food pathogens. *International Journal of Food Science and Technology*, 2009. 44(11): p. 2161-2168.
- [7] Quintavalla, S.V., L., Antimicrobial food packaging in meat industry. *Meat Science*, 2002. 62: p. 373-380.
- [8] Kandemir, N.e.a., Production of antimicrobial films by incorporation of partially purified lysozyme into biodegradable films of crude exopolysaccharides obtained from *aureobasidium oullulans* fermentation. *Food Technology and Biotechnology*, 2005. 43(4): p. 343-350.
- [9] Perez-Perez, C.e.a., Incorporation of antimicrobial agents in food packaging films and coatings. *Advances ,n Agricultural and Food Biotechnology*, 2006. 2: p. 194-216.
- [10] Bastarrachea, L.D., S., Engineering properties of polymeric-based antimicrobial films for food packaging. *Food Engineering Reviews*, 2011. 3: p. 79-93.

- [11] Appendini, P.H., J. H., Review of antimicrobial food packaging. *Innovative Food Science and Emerging Technologies*, 2002. 3: p. 113-126.
- [12] Kavas, G.K., N., Gıda ambalajlamasında biyonanokompozit yanilebilir filmler. *Gıda Dergisi*, 2010. 1: p. 95-97.
- [13] Hoffman, K.L.H., I. Y.; Dawson, P.L., Antimicrobial Effects on Corn Zein Films Impregnated with Nisin, Lauric Acid and EDTA. *Journal of Food Protection*, 2001. 64: p. 885-889.
- [14] Liu, Z.H., J. H. , Film-forming Characteristics of Starches. *Journal of Food Science*, 2005. 70(1): p. E31-E36.
- [15] Chen, L.I., Syed, H.; Gordon, Sherald, H.; Greene, Richard, V., Starch-Polyvinyl Alcohol Crosslinked Film-Performance and Biodegradation. *Journal of Environmental Polymer Degradation*, 1997. 5(2111-117).
- [16] Risbud, M.V.B., S. V. , Properties of polyvinyl pyrrolidone/ $\beta$ -chitosan hydrogel membranes and their biocompatibility evaluation by haemorheological method. *Journal of Materials Science: Materials in Medicine*, 2001. 12: p. 75-79.
- [17] Cha, D.S.C., Jin Hyuk; Chinnan, Manjeet S.; Park, Hyun Jin, Antimicrobial Films Based on Na-alginate and  $\kappa$ -carrageenan. *Food science & technology*, 2002. 36(8): p. 715-719.
- [18] Muthuswamy, E.R., S. Sree; Vasan H.N; Garcia Cecile; Noe, Laure; Verelst Marc Highly stable Ag nanoparticles in agar-agar matrix as inorganic-organic hybrid. *Journal of Nanoparticle Research*, 2006. 9: p. 561-567.
- [19] Sarasawathy, N.e.a., A Preliminary investigation of tumeric-agar composite film as bioactive wound dressing material on excision in rat model. *Indian Journal of Natural Products and Resources*, 2012. 3(2): p. 237-241.
- [20] Nohynek, G.A., E; Re, T; Toutain, H, Safety assessment of personal care products/cosmetics and their ingredients. *Toxicology and Applied Pharmacology*, 2010. 243(2): p. 239-259.
- [21] Padmavathy, N.V., R., Enhanced Bioactivity of ZnO nanoparticles- an antimicrobial study. *Science and Technology of Advanced Materials*, 2008. 9.
- [22] Rajendran, R., C. Balakumar, Hasabo A. Mohammed Ahammed, S. Jayakumar, K. Vaideki and E.M. Rajesh, Use of zinc oxide nano particles for production of antimicrobial textiles. *International Journal of Engineering, Science and Technology*, 2010. 2(1): p. 202-208.
- [23] Perelshtein, I., G. Applerot, E. Wehrchetz, A. Hasmann, M. Guebitz and A. Gedanken, Antibacterial Properties of an In Situ Generated and Simultaneously Deposited Nanocrystalline ZnO on Fabrics. *American Chemical Society*, 2008. 1(2): p. 361-366.
- [24] Ghule, K., Anil Vithal Ghule, Bo-Jung Chen, and Yong-Chien Ling, Preparation and characterization of ZnO nanoparticles coated paper and its antibacterial activity study. *Green Chemistry*, 2006. 8: p. 1034-1041.
- [25] Jin, T.S., D. ; Su, J. Y. ; Zhang, H. ; SWue, H. J., Antimicrobial Efficacy of Zinc Oxide Quantum Dots against *Lysteria monocytogenes*, *Salmonella enteridis* and *Escherichia coli* O157:H7. *Journal of Food Science*, 2009. 74(1): p. M46-M52.
- [26] Sharma, S.T., Alex; Nalamasu, Omkaram and Dutta, P. S, Spin-Coated ZnO Thin Films Using ZnO Nano-Colloid. *Journal of Electronic Materials*, 2010. 35(6): p. 1237-1240.
- [27] IMSL, JIS Z2801:2000. Antimicrobial product-test for antimicrobial activity and efficacy. 2001, Japanese Industrial Standard.
- [28] Patterson, A., The Scherrer Formula for X-Ray Particle Size Determination. *Physical Review*, 1939. 56(10): p. 978-982.
- [29] S Swinkels, J.W.G.M.K., Ervin T.; Verstegen, Martin W.A., Biology of Zinc and Biological Value of Dietary organic Zinc Complexes and Chelates. *Nutrition Research Reviews*, 1994. 7(1): p. 129-149.
- [30] Applerot, G.; et al.; Enhanced Activity of nanocrystalline ZnO Due to Increase ROS-Mediated Cell Injury. *Advanced Functional materials*, 2009. 19: 842-852.
- [31] Zhang, L.; et al. Investigation into the antibacterial behaviour of suspensions of ZnO nanoparticles (ZnO nanofluids). *Journal of Nanoparticle Research*, 2007. 9: p. 479-489.
- [32] Sawai, J.S., S; Igarashi, H; Hashimoto, A; Kokugan, T; Shimizu, M; Kojima, H, Hydrogen peroxide as an antibacterial factor in zinc oxide powder slurry. *Journal of Fermentation and Bioengineering*, 1998. 86(5): p. 521-522.
- [33] Sawai, J. ; Kawada, E. ; Kanou, F.; Igarashi, H.; Hashimoto, A.; Kokugan, T.; Shimizu, M., Detection of Active Oxygen Generated from Ceramic Powders Having Antibacterial activity. *Journal of Chemical engineering of Japan*, 1996. 29(4): p. 627-633.
- [34] Yamamoto, O.; Sawai, J.; Sasamoto, T., Activated Carbon Sphere with Antibacterial Characteristics. *Materials Transactions*, 2002. 43(5): p. 1069-1073.
- [35] URL <http://www.codexalimentarius.org/>
- [36] Zalan, Z.; Nemeth, E.; Barath, A.; Halasz, A., Influence of Growth Medium on Hydrogen Peroxide and Bacteriocin Production of *Lactobacillus* Strains. *Food Technology and Biotechnology*, 2005. 43(3): p. 219-225