

Trends for the Application of Passion Fruit Industrial By-Products: A Review on the Chemical Composition and Extraction Techniques of Phytochemicals

Juliane Viganó, Julian Martinez *

Department of Food Engineering, College of Food Engineering, University of Campinas, Campinas, Brazil

Abstract This work presents a review on the composition and extraction techniques for recovering phytochemicals from passion fruit by-products. The review approaches on the characteristics of each phytochemical and its benefits to human health. It was observed that the by-products of passion fruit contain a range of interesting substances, and for their recovery the application of environmentally friendly extraction techniques, such as supercritical fluid extraction (SFE) and pressurized liquid extraction (PLE), are suggested as single or sequential procedures. The future perspectives of this review encourage the application of these techniques to a complete recovery of passion fruit by-products.

Keywords By-Products, Phytochemicals, Supercritical fluid extraction, Pressurized liquid extraction, Sequential extraction, Passion fruit

1. Introduction

Researches carried out in the last years have demonstrated the growing interest in recovering phytochemicals from vegetal matrices. The main goal of this recovery is to obtain compounds with biological activities for use in drugs, cosmetics and as additives for foods. Thus, the present work shows a review on a particular case. An approach on the phytochemical compounds contained in the passion fruit industrial by-products and on the emerging techniques to extract such compounds is presented. Researches performed with edible parts, by-products and aerial parts indicate that passion fruit is a rich source of phenolic compounds [1-4], fatty acids [5], tocopherols, tocotrienols [6] and carotenoids [7], compounds of high nutritional value, which are related to many health benefits.

The industrial processing of passion fruit pulp generates a considerable volume of by-products due to the separation of rind and bagasse. It is estimated that these materials correspond to 60-70% of the fruit mass [8, 9]. Therefore, the development of techniques designed to use this type of material can add value and reduce the environmental impacts of its inappropriate disposal.

Several factors lead to chemical reactions that can degrade bioactive compounds limiting their use. To

eliminate such limitations, extractions using pressurized fluids are a good alternative. The extraction techniques that employ supercritical fluids (SFE – Supercritical Fluid Extraction) or subcritical fluids (PLE - pressurized liquid extraction) show some advantages when compared to conventional extraction techniques. Among these advantages, the selectivity and low generation of solvent waste seem to be the most attractive. However, some drawbacks may occur with respect to the solubility of target compounds. In order to overcome this problem and to increase the extraction efficiency, some authors suggest sequential extractions, in which different extraction techniques are applied to the same raw material.

Few works were found addressing the recovery of bioactive compounds from passion fruit industrial by-products. In addition, there is no record of works that have fully used passion fruit by-products through sequential extraction processes. Taking the mentioned information into account, this paper has as objective to discuss the reasons that make the passion fruit by-products an interesting source of phytochemicals and presents trends for their recovery using clean and efficient technologies.

2. The Yellow Passion Fruit

The genus *Passiflora* includes about 500 species. Among them, the specie *Passiflora edulis* is the most known, being widely used by the food industry, and used in America and Europe as a sedative or tranquillizer. This specie, commonly called sour passion fruit or yellow passion fruit, is native from Brazil, but has also been grown in other continents [3].

* Corresponding author:

julian@unicamp.br (Julian Martinez)

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The fruits, as shown in Figure 1, are berry type and have the seeds involved by the pulp and the fleshy aryl. The rind is composed by epicarp, which corresponds to the external layer of green to yellow color, and by the mesocarp, which is the white internal layer [10].

Brazil is the largest producer and consumer of yellow passion fruit. This specie is produced in 95% of the passion fruit orchards [3]. According to data from the Brazilian Institute of Geography and Statistics – IBGE [11], the production of this fruit in 2012 reached 776.097 tons. Brazilian Northeast and Southeast regions are the main responsible for this production.

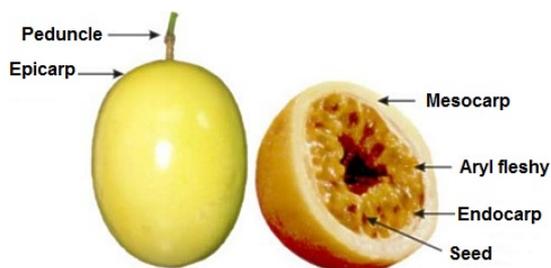


Figure 1. Passion fruit morphology

The cultivation of passion fruit is primarily focused on the juice and pulp industry, especially due to its more acidic taste and higher yield [10]. For obtaining the pulp, rind (epicarp and mesocarp) and bagasse (seed and aryl fleshy) are separated, resulting in by-products of the passion fruit processing industry. In most cases, this amount of biomass is used for animal feed or deposited on orchards to act as fertilizer. Although in small amounts, the rind has already been used for the formulation of flour, and the seeds are destined to the extraction of oil for cosmetics. The use of these wastes has environmental and economic advantages, since it prevents the inadequate disposal of biomass in the environment and can generate income through the formulation of new products with high added value.

3. Composition of Passion Fruit By-Products

The interest on the search for natural antioxidants for use in foods, aiming to decrease the lipid oxidation rate, or for pharmaceutical applications to prevent chronic diseases related to the production of free radicals, has increased in the recent years. Moreover, results of researches conducted with passion fruit by-products have been disclosed and show that the bagasse and rind are sources of several compounds of nutritional importance.

3.1. Bagasse

The passion fruit bagasse, as described previously, is composed by fleshy aryl and seeds. Bibliographic data regarding the composition of the fleshy aryl was not found. On the other hand, the seeds have been characterized and explored for different purposes. The main components of

passion fruit seeds are oil and fibers [12]. The oil is used to produce cosmetics and may be found commercially. Linoleic acid is one of the main fatty acids of passion fruit oil (around 72-73%), followed by oleic acid (13-16%) and palmitic acid (8-9%) [5, 6]. For human health, polyunsaturated fatty acids play important roles in the maintenance of cell membranes, brain function and the transmission of nerve impulses, and the growth and development of the cardiovascular system [8, 13].

Researches have also pointed that the passion fruit seed is a source of other substances, such as precursors of vitamin E (tocopherols and tocotrienols). Malacrida and Jorge [6] identified and quantified three fractions of tocopherols in passion fruit oil. They reported the following values for the fractions β -, γ - e δ - tocopherol: 54.0, 166.6 e 278.7 mg/kg oil, respectively. Vitamin E consists of eight distinct molecules α -, β -, γ - e δ -tocopherol e α -, β -, γ - e δ -tocotrienol (Table 1), collectively named as tococromanolos and tocols. These compounds have a chromanol group in one end attached to a long isoprenoid chain. Depending on the nature of the isoprenoid chain, a distinction is made between tocopherols (containing a saturated phytyl chain) or tocotrienols (unsaturated geranylgeranyl chain) [14].

Table 1. Chemical structures of tocols

<p style="text-align: center;">Tocopherol</p>			
<p style="text-align: center;">Tocotrienol</p>			
Prefix	R ₁	R ₂	R ₃
α -	CH ₃	CH ₃	CH ₃
β -	CH ₃	H	CH ₃
γ -	H	CH ₃	CH ₃
δ -	H	H	CH ₃
Source: Drotleff et al. [14]			

Numerous benefits for human health have been attributed to this class of compounds. Tocols delay the progress of a series of degenerative diseases, act on the protection against cancer and cardiovascular diseases, and have the function of reducing cholesterol in blood [15-17].

Carotenoids are an extensively studied class of compounds in the passion fruit juice. Nevertheless, works addressing the identification and quantification of carotenoids in passion fruit oil from seeds were not found. On the other hand, Ferreira et al. [18] investigated the total carotenoids of refined oil, obtained by cold pressing and Soxhlet, and the

reported carotenoid content was 2.3, 4.6 and 19.7 $\mu\text{g/g}$ oil, respectively. Carotenoids are known as responsible for many functional properties as antioxidant activity, prevention of cardiovascular disease, cancer, macular degeneration and in some cases pro-vitamin A [19].

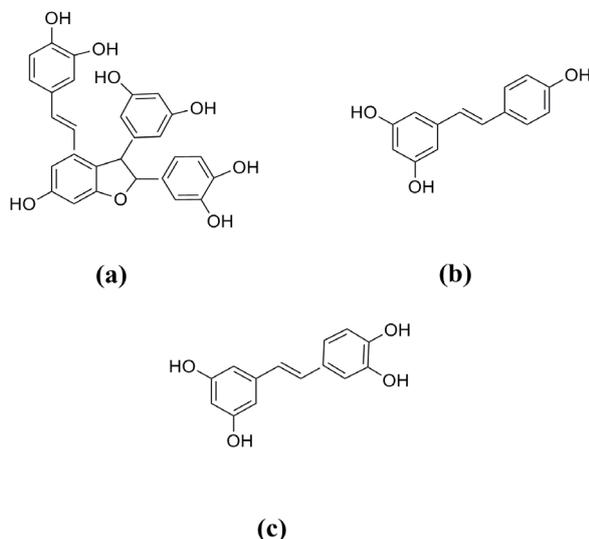


Figure 2. Chemical structure of (a) Scirpusin B, (b) resveratrol and (c) piceatannol

Recent studies have reported the presence of phenolic compounds in passion fruit seeds. Matsui *et al.* [20] found piceatannol and resveratrol (Figure 2), and reported that piceatannol has positive effect in inhibiting melanogenesis and collagen synthesis. The observed amounts of piceatannol

and resveratrol were 2.2 and 0.1 mg/g of dried seed, respectively. Sano *et al.* [21], apart from these two substances, found another phenolic compound known as Scirpusin B (Figure 2), but the authors quantified only piceatannol and Scirpusin B, which amounts were 5.7 and 3.6 mg/g of dried seed, respectively. Matsumoto *et al.* [22] used Scirpusin B for the evaluation of the coronary circulation of rats and found that this compound caused the increase in coronary blood flow via production of nitric oxide and vasodilating prostanoids. Scirpusin B is implicated to have beneficial effects on preventing cardiac events and atherosclerosis by increasing these substances responsible for vasodilatation.

Table 2 shows a comparison between substances found in passion fruit seeds and other sources. It can be observed that the fatty acid profile of passion fruit seeds is similar to that of seeds of other fruits, such as grape and guava. These three types of seeds are rich in polyunsaturated fatty acids. Regarding tocols, the presented sources do not resemble in the concentration. According the presented data, the carotenoid concentration in passion fruit seeds is lower than in pequi and buriti oils. However, this same reference (Ferreira *et al.* [18]) examined the same oil in terms of antioxidant activity, and passion fruit seed oil presented was the most active. This is possibly due to the content of tocopherols or phenolics, which proved to have antioxidant activity. As can be observed, passion fruit seeds present levels of piceatannol and resveratrol comparable to already recognized sources of these compounds.

Table 2. Comparison between the contents of substances of passion fruit seeds with other sources

Fatty acids (%)	Passion fruit seed oil [5]	Peach almond oil [23]	Grape seed oil [24]	Guava seed oil [25]
Linoleic	73	19	70-74	71
Oleic	16	72	13-17	9
Palmitic	9	6	5-9	10
Linolenic	0,3	0	0.3-0.7	0.5
Tocols (mg/100g oil)	Passion fruit seed oil [6]	Rice bran oil [26]	Grape seed oil [24]	Guava seed oil [6]
α - / β - / γ - / δ -Tocopherol	0 / 5.4 / 16.6 / 27.8	179 / 1 / 55 / 0	19.9 / 0 / 6.2 / 0	10.7 / 0.3 / 55 / 0.5
α - / β - / γ - / δ -Tocotrienol	-	96 / 0.7 / 588 / 19	2.6 / 0 / 198 / 0	-
Total carotenoids ($\mu\text{g/g}$ oil)	Passion fruit seed oil [18]	Pequi oil [18]	Babaçu oil [18]	Buriti oil [18]
	2-20	275	20	693
Phenolic (mg/g seed)	Passion fruit seed [20, 21]	Grape seed [27]	<i>Rhodomyrtus tomentosa</i> seed [28]	
Piceatannol	2-6	NQ	3-7	
Resveratrol	0.1	1.1-3.2	NI	
Scirpusin B	3.6	NI	NI	

NQ – Identified, but not quantified NI – Not identified.

3.2. Rind

The passion fruit rind is composed mainly by carbohydrates [29]. Its elevated pectin content has made it a source for the extraction of this substance and for the formulation of flour. However, research has revealed the rind as a source of interesting nutrients such as the phenolic compounds presented on Table 3.

The substances presented on Table 3 are phenolic compounds belonging to the class of C-glycosyl flavonoids. Their molecular structures are characterized by the presence of the flavan group, which comprises fifteen carbon atoms arranged into three rings, and five radicals, as shown in Table 4 [33].

Flavonoids are secondary metabolites that occur widely in plants. The function of those substances in the plant is wide. They act on protection against the harmful effects of UV rays and pathogens such as viruses, fungi, insects and

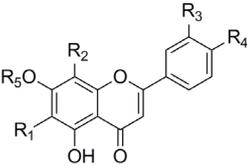
bacteria. Besides, they contribute to the pigmentation and control of plant hormones and enzymes [33, 34]. Regarding human health, flavonoids have sedative properties and play an important protective role against atherosclerosis, brain disorders and cancer [33, 35]. Part of the mentioned functions is due to the antioxidant activity of these compounds. Flavonoids are natural antioxidants that act as reducing agents, hydrogen donors, oxygen neutralizing and chelating metals that could initiate or propagate chain reactions [36].

The substances contained in the passion fruit bagasse and rind represent an opportunity for adding value to these by-products. Therefore, techniques that allow the separation of these substances from the vegetable matrix are necessary. The next section addresses extraction processes that can be applied to separate and isolate the bioactive components from passion fruit residues.

Table 3. Phenolic compounds identified in species of *Passiflora* genus

Compound	Part of plant	Specie	Reference
Vicenin-2	Leaves and pericarp Leaves	<i>P. edulis</i> var. <i>flavicarp</i> <i>P. tripartita</i> var. <i>mollissima</i>	[4]
6,8-di-C- glycosylchrysin	Leaves and pericarp	<i>P. edulis</i> var. <i>flavicarp</i>	[4]
Spinisin	Leaves and pericarp	<i>P. edulis</i> var. <i>flavicarp</i>	[4]
Swertisin	Leaves	<i>P. tripartita</i> var. <i>mollissima</i>	[4]
Vitexina-2''-O- rhamnoside	Leaves and pericarp Leaves Leaves	<i>P. alata</i> <i>P. quadrangulares</i> <i>P. manicata</i>	[4]
Isoorientin	Rind Rind Leaves and pericarp Leaves Leaves and pericarp Leaves Leaves Pulp	<i>P. edulis</i> <i>P. edulis</i> <i>P. edulis</i> var. <i>flavicarp</i> <i>P. alata</i> <i>P. tripartita</i> var. <i>molissima</i> <i>P. manicata</i> <i>P. edulis</i> <i>P. edulis</i>	[3, 4, 30-32]
Orientin	Leaves Leaves Leaves and pericarp Leaves	<i>P. edulis</i> var. <i>flavicarp</i> <i>P. alata</i> <i>P. tripartita</i> var. <i>molissima</i> <i>P. manicata</i>	[4]
Isovitexin	Leaves Leaves Leaves and pericarp Leaves Leaves	<i>P. edulis</i> var. <i>flavicarp</i> <i>P. alata</i> <i>P. tripartita</i> var. <i>molissima</i> <i>P. manicata</i> <i>P. edulis</i>	[3, 4]
Vitexin	Leaves Leaves Leaves Leaves	<i>P. edulis</i> var. <i>flavicarp</i> <i>P. tripartita</i> var. <i>molissima</i> <i>P. manicata</i> <i>P. edulis</i>	[3, 4]

Table 4. Chemical structure of C-glycosyl flavonoids

						
Compound	Aglycone	R ₁	R ₂	R ₃	R ₄	R ₅
Vicenin-2	Apigenin	Glu	Glu	H	OH	H
6,8-di-C- glycosylchrysin	Chrysin	Glu	Glu	H	H	H
Spinosin	Apigenin	Glu-O- Glu (1-2)	H	H	OH	CH ₃
Swertisin	Apigenin	Glu	H	H	OH	CH ₃
Vitexin-2''-O- rhamnoside	Apigenin	H	Glu-O-rham	H	OH	H
Isoorientin	Luteolin	Glu	H	OH	OH	H
Orientin	Luteolin	H	Glu	OH	OH	H
Isovitexin	Apigenin	Glu	H	H	OH	H
Vitexin	Apigenin	H	Glu	H	OH	H

Glu = glucose; rham = rhamnose
Source: Zucolotto *et al.* [4]

4. Extraction Process

The separation process of one or more components from a complex mixture is a requirement to several unit operations in food and chemical industries. Usually, the main objective of this operation is to remove a specific component to add value to the product. The specific component can be a residue, an interesting substance or both. The separation process can be classified according to the nature of the material submitted to separation, based on unit operations or classified by the type of phases in contact, such as solid-liquid, liquid-liquid, among others [37]. Basically, the separation of substances from vegetable matrices is classified as solid-liquid extraction, and can be carried out through conventional or non-conventional methods.

4.1. Conventional Extraction Methods

Conventional extraction methods are defined as classical techniques based on the extraction capacity of different solvents and/or application of heat and shaking. The classical techniques for obtaining substances from vegetable matrices are Soxhlet and maceration. Both techniques were already used to obtain extracts from passion fruit by-products. Malacrida and Jorge [6] obtained oil from passion fruit seeds using Soxhlet and Cazarin *et al.* [10] extracted bioactive compounds from the rind by maceration.

The advantage of both techniques is the simplicity of the systems and, therefore, low cost to assemble the extraction unit. On the other hand, conventional techniques have drawbacks such as high extraction times, the use of large amounts of solvent [38], low selectivity, the need for solvent evaporation and decomposition of labile compounds [13, 39,

40]. Thus, more environmentally friendly techniques that do not present health risks and provide high-quality extracts may be applied.

4.2. Non-conventional Extraction Methods

Extraction techniques that use pressurized fluids, ultrasound, enzymes, microwaves and electric pulse have shown promising results. Besides supplying the deficiencies of conventional methods, some of these techniques can be considered as "green" or environmentally friendly.

Extractions using pressurized fluids, besides reducing the extraction time and the amount of used solvent, can be more selective for obtaining target compounds. The process selectivity and velocity are consequences of the solvent physical and chemical properties, which in some cases can be controlled by changing the extraction parameters. In ideal solid-liquid extraction processes, the target compound must have high solubility in the employed solvent. Meanwhile, other components of the solid matrix should not be solubilized. In real processes this is rarely achieved. For this reason, several studies have sought to optimize conditions as the solvent/feed ratio, particle size, temperature, pressure, time and solvent flow rate to obtain the best performance in terms of the target compound [39]. Two extraction methods using pressurized fluids are commonly used: supercritical fluid extraction (SFE) and pressurized liquid extraction (PLE).

4.2.1. Supercritical Fluid Extraction (SFE)

Supercritical fluids are extensively used for extracting compounds from a large variety of matrices. Carbon dioxide (CO₂) is the most used solvent in SFE, since it is secure,

nontoxic and has high solubilization power, which provides high mass transfer rates. In addition, it is widely available, has low cost and can be obtained with high purity. Its critical point (7,34 MPa and 31,06°C) allow obtaining extracts without submitting them to thermal damage [41]. On the other hand, the disadvantage of the use of this technique is the high investment cost for the construction of equipment. This is another reason to focus on obtaining extracts with high added value from this process.

Supercritical CO₂ extraction processes (SC-CO₂) present an important tool based on the tunable solvent properties, i.e., the selectivity of SC-CO₂ can be adjusted by varying temperature and pressure in order to obtain fractions enriched in desirable compounds [42-44]. The economic value of extracts can be increased through the recovery of fractions rich in desired components, and this can be achieved after comprehending the SC-CO₂ mechanisms.

There is a region above the critical point in which every substance is a single phase fluid, non-condensing and showing some typical physicochemical properties typical of gases and liquids (Table 5), i.e., the density approaches that of a liquid while its viscosity is close to those of gases. The solvation capacity of CO₂ in the supercritical state depends on its density. The higher the density, smaller spaces between the molecules and bigger interactions between them are reached [45, 46]. The highest density values are attained by combining low temperatures (but no lower than the critical temperature) and high pressures.

Table 5. Density and viscosity values of CO₂

State	Density (kg/m ³)	Viscosity (μPa.s)
Liquid (27 °C; 50 MPa)	1029	133
Gas (40 °C; 0,1 MPa)	2	16
Supercritical (40 °C; 10 MPa)	632	17

Source: Williams and Clifford [47].

The SFE process is carried out in solid matrixes by means of the continuous contact between the solvent and the solid phase. In most cases, the solid is placed in a fixed bed and the solvent flows through it. After the extraction procedure, the solvent loaded with the extracted solute leaves the extractor and migrates to the precipitator or separator. The precipitation of the solute is made by simple reduction of the pressure below the critical point.

Extraction phenomena can be separated in three parts (Figure 3). The first corresponds to the constant extraction rate (CER) period, which is characterized by the extraction of the solute that covers the outer surface of the particles of the vegetable matrix. This is considered an easily accessible solute. In this step the mass convection mechanism predominates in the process. The second period, falling extraction rate (FER), starts when there is insufficient amount of solute to maintain the constant extraction rate. The last period is diffusion controlled (DC), in which the process is controlled only by the diffusion of the solvent inside the

particles and the diffusion of the solvent/solute to their surface [48].

The application of supercritical CO₂ results in good performance to extract nonpolar substances such as fatty acids, carotenoids, tocopherols and tocotrienols. Liu et al. [5] suggested SFE at temperature of 56 °C, pressure of 26 MPa and extraction time of 4 h to recover oil from passion fruit with global yield of 25.83%. However, the authors did not identify or quantify minor lipids such as carotenoids and tocopherols. Although few studies have reported data on the extraction conditions of passion fruit by-products, there is a vast amount of literature work whose data show the best conditions to recover target compounds from other vegetable matrices using SFE.

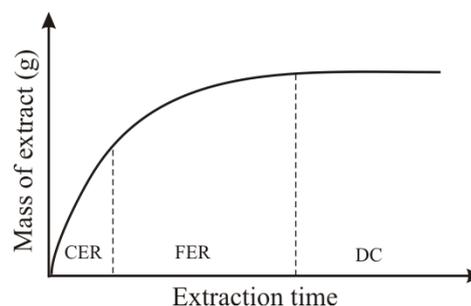


Figure 3. Typical extraction curve of a SFE process

The commonly studied variables in SFE are pressure, temperature, solvent flow rate, extraction time, particle size and extraction bed properties as height/diameter ratio. However, most studies focus efforts on the study of temperature and pressure. The employed temperatures are generally between 40 and 70°C. The lower limit is due to the proximity of the critical temperature of CO₂ and the upper limit affects the density of this solvent. Lower densities are achieved with increasing temperature, resulting in lower extraction yield and high energy consumption. Regarding pressure, most works explore the range 10-50 MPa. Pressure has the opposite effect of temperature on CO₂ density, i.e., increasing the pressure the density of CO₂ increases, and consequently its solvation capacity becomes greater. Pressures above the presented range lead to high energy costs and lower pressures approach the limit of the critical pressure of CO₂.

Many works have concentrated efforts to identify the variables of SFE that produce better results for the extraction of interesting compounds. The best conditions for extracting tocopherols seem to be dependent of raw material. Ciftci et al. [49] identified 70 °C and 49.6 MPa as the best condition to obtain total tocopherols from corn dried grains. Liu et al. [50] found 50°C and 15 MPa as the optimal condition to obtain total tocopherols from pomegranate. Finally, Sarmiento et al. [51] identified 40°C and 20 MPa to obtain total tocopherols from parboiled rice bran. With regard to carotenoids, Wijngaard et al. [39] made an extensive review on the SFE conditions and found that the extraction occurs at temperatures between 50-100°C and pressures between 30-40 MPa. In terms of fatty acids, the non-polar nature of these compounds make

them easily extractable with supercritical CO₂ under moderate pressures and temperatures, and in the case of thermolabile compounds, values should be set in the range of 35-50°C. Low pressures are required to extract fatty acids, generally from 10 MPa, but the general rule is: the higher is the pressure, the larger is the solvent power and the smaller is the extraction selectivity [52]. SFE also can be used to recover phenolic compounds. However, in most cases the use of a cosolvent is needed. A wide variety of extraction temperatures have been reported as optimal for polyphenol extraction (30-100°C). However, most authors usually affirm that the best temperatures are between 40 and 60°C, and the optimal pressures are between 20 and 30 MPa. Water and ethanol are the recommended cosolvents, but amounts higher than 10% (m/m) are not recommended because the solvent can leave the supercritical state [39]. In addition, SFE can show low efficiency to obtain polar compounds like polyphenols. An alternative to overcome this problem is the use of PLE processes [39, 53].

4.2.2. Pressurized Liquid Extraction (PLE)

PLE is based on the use of solvents at high pressure and temperature, but not above the critical point. The main objective of this technique is to promote the extraction of compounds from solid or semisolid matrices in short time and using small amount of solvent. The main advantage of PLE over conventional extraction methods is that pressurized solvents remain in the liquid state when taken to temperatures higher than their boiling points. These conditions improve the solvation power of the liquids and the desorption kinetics from the solid matrix. In addition, as well as SFE, this extraction method is carried out in an environment free from oxygen and light, which reinforced its potential use in extractions of nutraceutical compounds [54, 55].

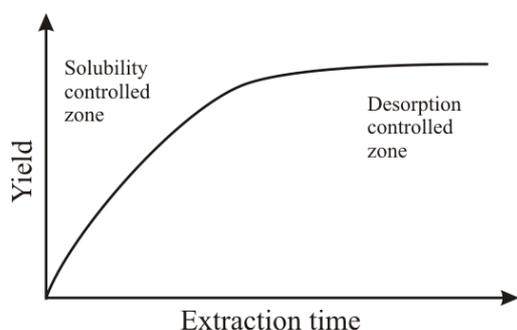


Figure 4. Typical curve of a PLE process. Adapted from Mustafa and Turner [56]

The PLE process can be divided in two stages. The first is a period when the extraction is controlled by the solubility, and the second is controlled by the diffusion of solutes in the solvent, as shown in Figure 4. The main solvents used in PLE are methanol, isopropanol, acetone, hexane and ether. Water and ethanol have been increasingly employed in the extraction of polyphenols, such as flavonoids and phenolic acids, because they are considered "green" solvents.

According to Mustafa and Turner [56], the use of solvent mixtures of two substances can help improving the solubility and increase the interaction with the target compound, i.e., one substance improves the solubility and the other improves the solute desorption.

No works were performed with passion fruit by-products employing PLE as extraction technique. However, several researches have indicated PLE as an alternative to recover phenolic compounds from a series of vegetable matrices. Wijngaard and Brunton [57] reported that the antioxidant activity of PLE extracts from apple pomace was increased 2.4 times in comparison to traditional methods.

According to Mustafa and Turner [56], temperature during the extraction is one of the critical factors that affect the efficiency and selectivity in PLE. The use of high temperatures improves the efficiency of the extraction as it helps the disruption of analyte-sample matrix interactions, decreases the surface tension of the solvent, solutes and matrix, decreases the viscosity of a liquid solvent and improves diffusion rate. On the other hand, the amount of co-extracted analytes might increase at higher temperatures, resulting in a decreased the selectivity of extraction. In addition, high temperatures might affect thermo-labile compounds that are subjected to disintegration and hydrolytic degradation. Regarding pressure, the use of high pressure helps forcing the solvent within the matrix pore to contact and extract the analytes. The use of high pressure during the extraction could result in the disruption of cells of the matrix, thus enhancing the mass transfer rate of the analyte from the sample to the solvent. However, the effect of pressure on the recovery of most substances is usually negligible in PLE, since liquids are known as incompressible fluids.

It has been observed that among the process variables, temperature and solvent type are the most studied, since pressures above 10 MPa have shown no significant effect on the characteristics of the extracts. PLE has been suitably used for the recovery of phenolic compounds from plant sources. The used temperature range is very wide (40-200°C), while the most used solvents are ethanol and water, individually or in mixture [39]. In this type of process, both temperature and solvent are strongly dependent of the characteristics of the target compounds, due to the thermal degradation of the compound and its polarity.

4.2.3. Sequential Extraction

The researches mentioned in Section 3 indicate the passion fruit by-products as potential sources to extract fatty acids, tocopherols, carotenoids and phenolic compounds. However, drawbacks occur because fatty acids, tocopherols and carotenoids are nonpolar compounds, while phenolics are polar. Therefore, it becomes unviable to obtain separate fractions of these components through extractions with the same solvent. To deal with this problem and to increase the efficiency and selectivity of extractions some authors suggest sequential processes [39, 58, 59], i.e., from the same

solid matrix, extraction techniques as SFE and PLE could be applied to obtain fractions of extract with distinct chemical characteristics. The selectivity of SC-CO₂ enables obtaining different fractions of extracts, each one rich in tocots (at low SC-CO₂ density), fatty acids (at intermediate SC-CO₂ density) and carotenoids (at high SC-CO₂ density). Next, PLE can lead to the recovery of phenolic compounds in which the power of each solvent might be explored.

Some works involving combinations of high pressure techniques to extract bioactive compounds from vegetable sources have been reported. Paula et al. [60] obtained phenolic compounds from *Arrabidaea chica* employing sequential techniques and using CO₂, water and ethanol as solvents. The authors achieved better results in terms of global yield and total phenolics when compared with conventional extraction. Serra et al. [61] extracted, in sequential steps, bioactive substances from cherries using CO₂ and ethanol and concluded that the sequential process produces extracts more concentrated and with higher biological activity. Similar conclusions were obtained by Garmus et al. [62] and [63] for sequential extraction of pitanga and pepper-rosmarin leaves.

5. Conclusions and Future Perspectives

The chemical composition of passion fruit by-products encourages their recovery from industrial processes for novel applications and products. The researches reported in this work show that the passion fruit by-products are source of several phytochemical compounds of interest to food and pharmaceutical industries. The use of this type of material allows reducing the environmental damages, as well as adds value to a material that would be usually discarded.

The application of extraction techniques that employ pressurized fluids reinforces the appeal of using by-products, since the proposed techniques are considered environmentally friendly and enable the use of GRAS solvents, which are favourable for applications in food. In the same way, sequential extraction processes are seen with expectation for the recovery of phytochemicals from food by-products. Combined techniques promote a better exploration of the vegetal matrix, leading to the extraction of different classes of substances from the same source, and accordingly, leading to fractions of extract concentrated in target compounds. The passion fruit bagasse can be used as example to this approach because it is rich in polar compounds (oil, tocots and carotenoids) that can be recovered in steps by SFE, and nonpolar compounds such as polyphenols that can be properly obtained by PLE with ethanol and water as solvents.

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