# Synthesis and Characterization of $\gamma$-irradiated PVA/PEG/CaCl $\mathbf{2}_{\mathbf{2}}$ Hydrogel for Wound Dressing 

Joydeep Dutta<br>Department of Chemistry, Disha Institute of Management and Technology, Satya Vihar, Raipur, 492101, India


#### Abstract

The mixtures of polyvinyl alcohol (PVA) and polyethylene glycol (PEG) with calcium chloride $\left(\mathrm{CaCl}_{2}\right)$ were exposed to gamma-irradiation to synthesize transparent hydrogels for wound dressing. In this system, $\mathrm{CaCl}_{2}$ was used not only as a gelling material but also as a plasticizer. The presence of $\mathrm{CaCl}_{2}$ enhances the effects of PEG, this synergistic effects give births a new hydrogel with improved characteristics. The water absorption capacity of the hydrogel was in the range of $350-375 \%$ after immersion in normal saline for 72 h . In vitro weight loss experiments were conducted at $37^{\circ} \mathrm{C}$ in order to simulate the use of dressing on wounded skin. The enhanced thermal stability of the hydrogel was confirmed by thermogravimetric analysis (TGA). FTIR studies confirmed the cross-linking between PVA and PEG in the hydrogel. Furthermore, the microbe penetration test of the hydrogel was carried out to confirm its impermeability to bacteria. The effect of hydrogel on cell proliferation was also evaluated to ensure that the hydrogel does not release materials which might be deleterious to cell proliferation. The results showed that the hydrogel could be considered as a potential wound-dressing material.


Keywords Hydrogel, Radiation, Polyvinyl Alcohol, Polyethylene Glycol, Calcium Chloride, Wound Dressing

## 1. Introduction

In the recent past, hydrogels have received increasing attention due to their potential application in wound healing [1-4]. According to the old concept of wound healing, the use of cotton and gauze was quite significant to keep the wound dry. It has been observed by Winter[5] that a scab formation occurs when a wound is left open without any dressing thereby decreasing the rate of wound epithelization. He showed that a moist wound dressing can remarkably increase the rate of epithelization by preventing the scab formation. The hydrogels are suitable for absorbing exudates and serve as temporary physiological covers. It also provides efficient shielding as well as protects the wound from mechanical trauma.

PVA is an important material because it is non-toxic, non-carcinogenic, biodegradable, biocompatible, water soluble, and non-expensive polymer[6,7]. It has been extensively commercialized and studied in the chemical and medical industries[8] for the productions of fibers, films, coatings, cosmetics, pharmaceuticals, etc. PVA-based hydrogels are promising wound dressing materials because of their permeability to small molecules, impermeability to bacteria, soft-consistency, low interfacial tension, high water content and transparency[9].

[^0]Hydrogels can be prepared in various ways. Motoyama et al.[10] demonstrated the preparation of PVA hydrogel by using covalent cross-linking agent such as glutaraldehyde into aqueous solution of PVA. In another study, Singh et al. [11] reported a PVA hydrogel chemically cross-linked with an aldehyde such as formaldehyde, glutaraldehyde, terephthalaldehyde, and hexamethylenediamine to increase the gel strength. The disadvantages of such hydrogels include presence of residual cross-linking agents in the hydrogel which could be toxic to the tissues. Further, the elimination of the residuals increases the production cost. Varshney[12] demonstrated the synthesis of PVA-based hydrogel by gamma-irradiation technique. Besides, the above hydrogel also contains agar and carrageenan as other components, and is used for treating burns, non-healing ulcers of diabetes, and other external wounds etc. The use of agar as a mechanical strength enhancing agent in hydrogel synthesis makes it susceptible to microbial contamination. In another study, Mirzan et al.[13] reported the synthesis of gamma-irradiated polyvinyl alcohol-polyvinyl pyrrolidone (PVA-PVP) blended hydrogel. They investigated the gel fraction, mechanical properties, water content and water absorption capacity of the hydrogel for wound dressing. Many of the researchers have reported the synthesis of PVA-based hydrogels by using freeze-thaw method[14-16]. Altering process parameters, such hydrogels can be made transparent by using this method. In a study, Kim et al.[17] reported the development of polyvinyl alcohol-alginate gel-matrix-based wound dressing containing nitrofurazone by using freeze-thaw method. In another study, Nho et al.[18]
reported a novel PVA/PVP/glycerin/antibacterial agent hydrogel for wound dressing by using gamma-irradiation followed by freeze-thaw method. Freeze-thaw is a very expensive and time-consuming method whereas gamma- irradiation method accomplishes gel formation and sterilization simultaneously in one step.

Bearing in mind that gamma-irradiation reduces the production cost as well as to avoid chemical cross-linkers, we have developed a novel hydrogel which is composed of polyvinyl alcohol, polyethylene glycol, and calcium chloride. To the best of our knowledge, the development of novel $\mathrm{PVA} / \mathrm{PEG} / \mathrm{CaCl}_{2}$ hydrogel has not been reported so far. This hydrogel as a wound dressing material has been demonstrated by its good water absorption capacity, high water content and minimum weight loss. Its impermeability to microbial contamination and its non-cytotoxic nature reflect its advantages as a suitable wound dressing material.

## 2. Experimental

### 2.1. Preparation of Gel Systems

First of all, $\mathrm{CaCl}_{2}$ (Sigma, Germany) was dissolved in de-ionized water system followed by the addition of approximate amounts of PEG 4000 (Thomas Baker, Mumbai, India) to it. Once, the PEG 4000 got into the solution, 20\% w/w of PVA M. W. 14,000 (West Coast Laboratories, Mumbai, India) was directly added to the system. Then the entire mixture was subjected to autoclave for 1 h for complete dissolution. After autoclaving, the solution was deaerated and poured into trays. The trays were then inserted into plastic pouches and sealed. Finally, the trays containing solution were exposed to gamma-irradiation at 25 kGy for the formation of hydrogels.

### 2.2. Characterization

For swelling experiments, the hydrogel samples were immersed in excess normal saline solution at $37^{\circ} \mathrm{C}$. At predetermined time point, the hydrogels were taken out and weighed after removal of surface water with cotton cloth. Swelling was calculated as follows

$$
\begin{equation*}
\text { Swelling }(\%)=\frac{W_{S}-W_{i}}{W_{i}} \times 100 \tag{1}
\end{equation*}
$$

where $\mathrm{W}_{\mathrm{i}}$ is the initial weight of the prepared hydrogel and $\mathrm{W}_{\mathrm{s}}$ is the weight of the hydrogel in swollen state.

To examine the relative degree of cross-linking in the hydrogels, the swelling ratio of each hydrogel was also determined. The equilibrium swelling ratio is inversely related to the extent of cross-linking. The procedure for this experiment is mentioned elsewhere[19].

The water content in the hydrogel was estimated by drying it at $110^{\circ} \mathrm{C}$ for 6 h to thoroughly remove the water contained therein. The water content was calculated as follows

$$
\begin{equation*}
\text { Water content }(\%)=\frac{W_{0}-W_{1}}{W_{0}} \times 100 \tag{2}
\end{equation*}
$$

where $\mathrm{W}_{0}$ and $\mathrm{W}_{1}$ are, respectively, the weights of hydrogel
before and after drying.
The weight loss was determined by putting the hydrogel samples on each $10 \times 10 \mathrm{~cm}^{2}$ glass plate, followed by placing them in the oven at $37^{\circ} \mathrm{C}$. At predetermined time point, the hydrogels were taken out and weighed. The percentage weight loss was calculated according to the following equation

$$
\begin{equation*}
\text { Weight loss }(\%)=\frac{W_{i}-W_{r}}{W_{i}} \times 100 \tag{3}
\end{equation*}
$$

where $\mathrm{W}_{\mathrm{r}}$ is the reduced weight of the hydrogel after water loss.

Tensile strength and elongation at break of the hydrogel specimen were determined using LLOYD (Model: LRX Plus) instrument at a speed of $300 \mathrm{~mm} / \mathrm{min}$ maintaining a distance 50 mm between the jaws. In this case, a hydrogel strip with a length of 6 cm and a width of 2 cm was cut from $6 \times 6 \mathrm{~cm}^{2}$ hydrogel sheet of 3 mm thickness, the upper and the lower portion of the hydrogel was wrapped with plain paper and finally, placed in between two clamps.

Thermogravimetric analyses (TGAs) were carried out with Mettler TG-50 thermoanalyser. In this study, about 20.0 to 20.5 mg of the hydrogel samples were weighed individually in an alumina crucible. Samples were heated from $35^{\circ} \mathrm{C}$ to $500^{\circ} \mathrm{C}$ under nitrogen atmosphere at a rate of heating, $10^{\circ} \mathrm{C} / \mathrm{min}$. The weight change of each sample was continuously monitored. The weight loss was plotted against the temperature.

Spectra were recorded on FTIR Spectrophotometer (Make: Perkin Elmer, model: Spectrum 100) using Horizontal ATR assembly placing the hydrogel sample on ZnSe flat plate and covered with rubber coated cover plate in the range from $4000-650 \mathrm{~cm}^{-1}$.

The microbe penetration test was carried out to evaluate the resistance of the hydrogel wound dressings against microbe transmission from environment to the top surface of the wound. Briefly, bacterial culture lawns (Staphylococcus aureus, Escherichia coli and Pseudomonas aeruginosa) were prepared on Blood agar and MacConkey agar plates. Small pieces of hydrogel sheets were placed on the culture lawn. After overnight incubation the upper surfaces of hydrogels were exposed to sterile media contained in Petri dishes by Agar-overlay method. The exposed media in the Petri dishes were incubated at $37^{\circ} \mathrm{C}$ for 48 h and examined for any bacterial growth.

The effect of hydrogels on skin cells was evaluated to ensure that the hydrogels do not release materials that might be detrimental to the growth of skin cells. For this, the hydrogels were incubated with cell specific culture media (Gibco, USA) for different time points. The contact media was added to cultures of keratinocytes and fibroblasts. The effect of hydrogels on cell growth was monitored for 24, 48, 72 and 96 h.

Cell proliferation was analyzed indirectly by the MTT method[20]. Briefly, MTT ( $0.5 \mathrm{mg} / \mathrm{ml}$ ) was added to the cells in triplicate dishes after 24, 48, 72 and 96 h exposure to the contact media. Viable cells were indirectly determined
by their ability to convert soluble MTT to insoluble formazan crystals. The crystals were solubilized and the absorbance was determined as difference in optical density measured at a test wavelength of 570 nm and a reference wavelength of 650 nm (Shimadzu UV-VIS Spectrophotometer, Japan). At each end point, the absorbance was recorded and the optical density was compared to control.

## 3. Results and Discussion

Introduction of PEG into the macromolecular chain of PVA in presence of $\mathrm{CaCl}_{2}$ improved some of the characteristics of the final hydrogel (Scheme 1). The mechanism for the formation of PVA hydrogel using gamma-irradiation is already known[21]. Irradiation of aqueous PVA solution results in a chemical cross-linking of the polymer chains by forming covalent bonds.


Scheme 1. Interaction between PVA and PEG in presence of $\mathrm{CaCl}_{2}$

### 3.1. Swelling Study of Hydrogels

The gel thickness was optimized by performing swelling measurements only for PVA hydrogels of different thicknesses namely $3 \mathrm{~mm}, 4 \mathrm{~mm}$ and 5 mm (Fig. 1). As can be seen from the Fig. 1, there is no marked difference of the swelling capabilities of the hydrogels of varying thicknesses upto 24 h . After that the degree of swelling of the hydrogel of 3 mm thickness remarkably increases over time and reaches equilibrium at 72 h . On equilibration, the swelling increased to more or less $300 \%$. On the basis of the degree of swelling, 3 mm thickness of the hydrogel was optimized for wound dressing.


Figure 1. Time course of swelling change in normal saline solution at $37^{\circ} \mathrm{C}$ for only PVA hydrogels of various thicknesses

The fluid uptake efficiency of the hydrogel is an important criterion for providing a moist environment over wound bed [22]. Keeping in mind, the PVA was blended with the poly-
ethylene glycol in water alone or aqueous calcium chloride solution. The degree of swelling of hydrogels of different compositions namely PVA/water $\left(\mathrm{A}_{1}\right), \mathrm{PVA} / \mathrm{CaCl}_{2}\left(\mathrm{~A}_{2}\right)$, PVA/PEG $\left(\mathrm{A}_{3}\right)$ and PVA/PEG/ $\mathrm{CaCl}_{2}\left(\mathrm{~A}_{4}\right)$ was plotted as a function of time. Kinetic curves of swelling of the hydrogels are displayed in Fig. 2. The result showed that the degree of swelling of all the hydrogels increased with time. Apart from this, the relative degree of cross-linking was determined from the swelling ratio (data has not shown) of each sample. It was found that the cross-linking density of the $\mathrm{A}_{2}$ hydrogel was relatively higher as compared to other hydrogels, resulting in the absorption of less water. On the other hand, formation of the $\mathrm{A}_{4}$ hydrogel with low cross-linking density, showed the highest degree of swelling ranging between $350 \%$ and $375 \%$ after immersion in normal saline for 72 h compared to other hydrogels that showed $275-300 \%$. In a study, Bencherif et al.[19] had also demonstrated that lower the cross-linking density of the hydrogel, higher will be the water absorption capacity. This characteristic of the $\mathrm{A}_{4}$ hydrogel could be the positive indication for treating fullthickness exuding wounds.


Figure 2. Swelling behaviour of hydrogels of different compositions


Figure 3. Experimental data of water content of hydrogels of different compositions

### 3.2. Water Content Analysis

As shown in Fig. 3, it was observed that the water content of the hydrogels of different compositions to be in the range between $76 \%$ and $79 \%$. Mirzan et al.[13] showed that higher the water content in the PVA-PVP hydrogel, lesser is the ability to absorb more water. Here, the $\mathrm{A}_{4}$ hydrogel pos-
sesses the least water content i.e. $76.65 \pm 0.66 \%$ but the highest water absorption i.e. $359.78 \pm 13.33 \%$ after 72 h immersion in normal saline compared to other hydrogels which agrees the above statement.

### 3.3. Weight Loss Study

Weight loss experiments were conducted at $37^{\circ} \mathrm{C}$ in order to simulate the use of dressing on wounded skin. Fig. 4 shows the relative weight loss of the hydrogels of different compositions. However, the weight loss of the $\mathrm{A}_{4}$ hydrogel is $17.64 \pm 0.79 \%$ at 72 h whereas in other hydrogels the weight loss is in the range of $22-25 \%$. This phenomenon can be explained as follows: 1) due to hydrophilic nature, PEG attracts more water molecules towards itself as compared to PVA, and 2) due to hygroscopic nature of $\mathrm{CaCl}_{2}$, and it holds maximum numbers of water molecules and does not allow them to escape from the surface of the $\mathrm{A}_{4}$ hydrogel. Surprisingly, the only presence of PEG or $\mathrm{CaCl}_{2}$ in the hydrogel does not exhibit the said phenomenon. Therefore, it is assumed that this interesting characteristic of the $\mathrm{A}_{4}$ hydrogel may accelerate the healing process by providing a moist environment that leads to rapid granulation and re- epithelization; and concurrently, may prevent the wound from scab formation. Here, it is quite noteworthy to mention that the moist environment is conducive for faster wound- healing.


Figure 4. Weight loss of hydrogels of different compositions
On the basis of swelling, water content and weight loss study; the $\mathrm{A}_{4}$ hydrogel was chosen for further characterization.

Table 1. Data of tensile strength and elongation at break

| Maximum <br> load (Kgf) | PVA/PEG/CaCl $\left(\mathrm{A}_{4}\right)$ | Elongation (\%) |
| :---: | :---: | :---: |
|  | Tensile strength $\left(\mathrm{Kgf} / \mathrm{cm}^{2}\right)$ |  |
| 0.09 | 0.16 | 232 |

### 3.4. Tensile Strength and Elongation

As a consequence of their high water content, hydrogels lose mechanical strength which makes it difficult to handle. Tensile strength and elongation at break of the $A_{4}$ hydrogel were measured to find its suitability as a wound dressing material. The results are shown in Table 1. It shows that the tensile strength and elongation at break of the $\mathrm{A}_{4}$ hydrogel
are $16 \times 10^{-2} \mathrm{Kgf} / \mathrm{cm}^{2}$ and approximate $233 \%$ respectively. Thus, the $\mathrm{A}_{4}$ hydrogel possesses sufficient mechanical strength for its easy handling during its application on the wound.

### 3.5. Thermogravimetric Analysis (TGA)

Thermal decomposition behaviour of the $\mathrm{A}_{4}$ hydrogel was determined by thermogravimetric analysis. Fig. 5 shows that initial decomposition starts for (a) PVA at $243^{\circ} \mathrm{C}$ and (b) for PEG at $328^{\circ} \mathrm{C}$. In Fig. 5 (c), it was found that weight suddenly dropped to $76 \%$ was due to loss of water content in the $\mathrm{A}_{4}$ hydrogel. The initial decomposition temperatures of both PVA and PEG were higher than those which were shown in their individual therograms, indicating that PEG in presence of $\mathrm{CaCl}_{2}$ improved the thermal stability of the hydrogel, because the hydroxyl groups of PEG formed hydrogen bond cross-linking with hydroxyl groups of PVA. This restricted the chain movement during thermal treatment which is confirmed by FTIR spectroscopy as discussed below.


Figure 5. Thermograms of (a) PVA, (b) PEG, and PVA/PEG/ $\mathrm{CaCl}_{2}$ hydrogel

### 3.6. Fourier Transform Infrared Spectroscopy (FTIR)

In Fig. 6 (c), the FTIR spectrum of the $\mathrm{A}_{4}$ hydrogel exhibits several characteristic bands of stretching and bending vibrations of $\mathrm{O}-\mathrm{H}, \mathrm{C}-\mathrm{H}, \mathrm{C}=\mathrm{C}$ and $\mathrm{C}-\mathrm{O}$ groups. Two broad and strong bands were observed at 3318 and $657 \mathrm{~cm}^{-1}$ corresponding to $\mathrm{O}-\mathrm{H}$ stretching frequency. Typically, strong hydroxyl bands for free alcohol (nonbonded -OH stretching) and hydrogen bonded bands were found in the respective region between $3600-3650 \mathrm{~cm}^{-1}$ and 3200-3500 $\mathrm{cm}^{-1}$ [23-25]. The respective characteristic absorption bands at 3408 and $3391 \mathrm{~cm}^{-1}$ for hydroxyl groups were found in the FTIR spectra of PVA and PEG shown in the Fig. 6 (a) and 6 (b). Therefore, shifting of the band to a lower wave number i.e. at $3318 \mathrm{~cm}^{-1}$ as well as band widening clearly indicates the presence of hydrogen bond in the $\mathrm{A}_{4}$ hydrogel. Again, in Fig. 6 (c), the band observed at $2952 \mathrm{~cm}^{-1}$ indicates an asymmetry in stretching mode of $\mathrm{CH}_{2}$ group. A weak band was observed at $2137 \mathrm{~cm}^{-1}$ and had been assigned to the combination frequency of $(\mathrm{CH}+\mathrm{CC})$. The band at $1639 \mathrm{~cm}^{-1}$ has been assigned to $\mathrm{C}=\mathrm{C}$ stretching mode. Two bands observed at 1417 and $1380 \mathrm{~cm}^{-1}$ have been attributed to bending modes of $\mathrm{CH}_{2}$
groups. A weak band at $1326 \mathrm{~cm}^{-1}$ was assigned to the combination frequency of $(\mathrm{CH}+\mathrm{OH})$ group. The FTIR spectrum for the system $\mathrm{PVA} / \mathrm{PEG} / \mathrm{CaCl}_{2}$ showed a characteristic absorption band for - C-O-C- stretching vibration at $1094 \mathrm{~cm}^{-1}$. This again confirms the cross-linking between PVA and PEG in this hydrogel.


Figure 6 (a). FTIR spectrum of PVA


Figure 6 (b). FTIR spectrum of PEG


Figure 6 (c). FTIR spectrum of PVA/PEG/ $\mathrm{CaCl}_{2}$

### 3.7. Microbe Penetration Test

Based on the microbe penetration test it was found that no bacteria passed through the $\mathrm{A}_{4}$ hydrogel into sterile media even after 48 h . This indicated that the 3 mm thickness of the hydrogel dressing could protect the wound from getting
infected. In another study, Kokabi et al.[26] also reported that 3 mm thickness of PVA-clay nanocomposite hydrogel protected wound from the same, while Purna et al.[27] had shown that the use of 64 layers of gauze was inefficient to prevent microbial penetration. This ability of the hydrogel would protect the wound from infection.

### 3.8. Cytotoxicity Studies

Cytotoxicity of $\mathrm{A}_{4}$ hydrogel was indirectly assessed by studying the effect of hydrogel exposed media on keratinocytes and fibroblasts proliferation using the MTT method which is shown in Fig. 7. No significant difference in the proliferation of cells treated with either control media or hydrogels exposed media was observed. Similar rate of proliferation in keratinocytes and fibroblasts indicated that the hydrogel was non-toxic and its application on the wound bed would not interfere with cell proliferation. In another study, Sirousazar et al.[28] also reported that polyvinyl alcohol/clay nanocomposite hydrogel is non-toxic and biocompatible wound dressing. Therefore, it can be said that in vitro cytotoxicity studies give the preliminary authentication of the material to be used as wound dressing is non-toxic and biocompatible as well.


Figure 7 (a). The effect of hydrogel exposed media on keratinocytes proliferation


Figure 7 (b). The effect of hydrogel exposed media on fibroblasts proliferation

## 4. Conclusions

The above studies demonstrated that the PVA/PEG/ $\mathrm{CaCl}_{2}$ $\left(\mathrm{A}_{4}\right)$ hydrogel of 3 mm thickness was suitable for providing a moist environment over the wound bed because the percent weight loss of the $\mathrm{A}_{4}$ hydrogel was found to be less than $19 \%$ at 72 h whereas in other hydrogels the weight loss was between $22 \%$ and $25 \%$. In addition, suitable fluid uptake efficiency (350-375\% after immersion in normal saline for 72 h ) would enable the said hydrogel to provide an adequate moist environment that leads to rapid granulation and re- epithelization to wound closure. The enhanced thermal stability of the hydrogel indicated that both PVA and PEG were cross-linked together through hydrogen bond. This was again confirmed by FTIR study. Microbial penetration test revealed that the $\mathrm{A}_{4}$ hydrogel could be considered as a good barrier against microbes. The cytotoxicity studies showed no inhibition on cell proliferation. Furthermore, the transparent appearance of the hydrogel would obviate the need for frequent dressing changes to monitor the wound from outside. Finally, the above properties of the $\mathrm{A}_{4}$ hydrogel suggest that it can be used as a wound dressing in practical wound management.

## REFERENCES

[1] M. Wu, B. Bao, F. Yoshii and K. Makuuchi,Nucl. Sci. Tech., 11, 59 (2000).
[2] Y. C. Nho and K. R. Park, J. Appl. Polym. Sci., 85, 1787 (2002).
[3] M. Zhai, F. Yoshii, T. Kume and K. Hashim,Carbohydr. Polym., 50, 295 (2002).
[4] H. L. A. El-Mohdy and E. A. Hegazy, J. Macromol. Sci., Part A: Pure and Applied Chemistry, 45, 995 (2008).
[5] G. D. Winter, J. Wound Care, 4, 366 (1962).
[6] P. Giusti, L. Lazzeri, N. Barbani, P. Narducci, A. Bonaretti, M. Palla and L. Lelli, J. Mater. Sci.: Materials in Medicine, 4, 538 (1993).
[7] C. Valenta and B. G. Anver,Eur. J. Pharm. Biopharm., 58, 279 (2004).
[8] M. R. Fechner and M. Kon, Eur. J. Plast. Surg., 13, 43 (1990).
[9] M. Kita, Y. Ogura, Y. Honda, S. H. Hyon, W. I. Cha and Y. Ikada, Grafe's Arch. Clin. Exp. Opthalmol., 228, 533 (1990).
[10] T. Motoyama and S. Okamura, Kobunshi Kagaku, 11, 23 (1954).
[11] H. Singh, P. Vasudevan and A. R. Ray, J. Sci. Ind. Res., 39, 162 (1980).
[12] L. Varshney, Nucl. Instrum. Methods Phys. Res. Sec B: Beam Interactions with Materials and Atoms, 255, 343 (2007).
[13] T. R. Mirzan, D. Darmawan and S. Zainuddin, Radiat. Phys. Chem., 62, 107 (2001).
[14] M. Velazco-Diaz, F. A. Ruiz, M. C. Doria-Serrano, A. Gonzàlez-Montiel and M. Zolotukin, Ind. Eng. Chem. Res., 44, 7092 (2005).
[15] M. J. Mc Gann, C. L. Higginbotham, L. M. Geever and M. J. D. Nugent, Int. J. Pharm., 372, 154 (2009).
[16] M. Sirousazar and M. Yari, Chinese J. Polym. Sci., 28, 573 (2010).
[17] J. O. Kim, J. K. Park, J. H. Kim, S. Giujin, C. S. Yong, D. X. Li, J. Y. Choi, J. S. Woo, B. K. Yoo, W. S. Lyoo, K. Jung-Ae and C. Han-Gon, Int, J, Pharm,. 359, 79 (2008).
[18] Y. C. Nho, Y. M. Lim, H. Gwon and E. K. Choi, Korean J. Chem. Eng. , 26, 1675 (2009).
[19] S. A. Bencherif, A. Srinivasan, F. Horkay, J. O. Hollinger, K. Matyjaszewsky and N. R. Washburn, Biomaterials, 29, 1739 (2008).
$[20]$ B. L. Molinari, D. R. Tasat, M. A. Palmieri, S. E. O'Connor and R. L. Cabrini, Eur. Polym. J., 25, 254 (2003).
[21] B. Wang, S. Mukatak, E. Kokufuta and M. Kodama, Radiat. Phys. Chem.,59, 91 (2000).
[22] B. Balakrishnan, M. Mohanty, P. R. Umashankar and A. Jayakrishna, Biomaterials, 26, 6335 (2005).
[23] N. A. Peppas, Polymer, 18, 403 (1977).
[24] N. A. Peppas, Makromol. Chem., 178, 595 (1977).
[25] N. A. Peppas and L. Wright, Macromolecules, 29, 8798 (1996).
[26] M. Kokabi, M. Sirousazar and Z. M. Hassan, Eur. Polym. J., 43, 773 (2007).
[27] S. K. Purna and M. Babu, Burns, 26, 54 (2000).
[28] M. Sirousazar. M. Kokabi and Z. M. Hassan, J. Biomed. Sci. Polym. Ed., 22, 1023 (2011).


[^0]:    * Corresponding author:
    dutta_joy@yahoo.co.in (Joydeep Dutta)
    Published online at http://journal.sapub.org/chemistry
    Copyright © 2012 Scientific \& Academic Publishing. All Rights Reserved

