

Present Susceptibility Status of *Culex Quinquefasciatus*, Say to Four Insecticides at Mysore, India

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Abstract The current susceptibility status of *Culex quinquefasciatus* at Mysore, India to four insecticides was determined using standard WHO larval bioassay method. *Culex quinquefasciatus* is the primary vector of lymphatic filariasis, an endemic disease in India in 17 states and 6 union territories with about 553 million people at risk of infection. So, an effort was made to determine the susceptibility of this mosquito to 3 chemical insecticides viz Deltamethrin, Lambdacyhalothrin, Propoxur and a biocide, Spinosad. Among these Spinosad was found to be the most effective insecticide with LC₅₀ of 0.000002 ppm followed by Lambdacyhalothrin, Propoxur and Deltamethrin with LC₅₀ values of 0.00001, 0.00013 and 0.00062 ppm respectively. The results revealed the importance of a biocide explored from nature maybe employed to prevent pollution and health hazards.

Keywords *Culex quinquefasciatus*, Bioassay, Spinosad, Biocide

1. Introduction

Mosquitoes account for majority of the transmissions among the vector-borne diseases. They are the most important of the disease vectors that number over 350 species worldwide out of about 3500 species. Mosquito-transmitted diseases are the major cause of loss of human life worldwide, with over 700 million people suffering annually^[1]. Such diseases vectored by mosquitoes in India are malaria, lymphatic filariasis, dengue, chikungunya and Japanese encephalitis. *Culex quinquefasciatus* is the main vector which transmits microfilariae of *Wuchereria bancrofti* that causes about 98% of the lymphatic filariasis cases, a major neglected tropical disease. A district-level endemicity map created for India in 2000 shows that of the 289 districts surveyed up to 1995 (62% of all districts), as many as 257 were found to be endemic. Seventeen states and six Union Territories were identified to be endemic with about 553 million people exposed to the risk of infection; and of them, about 146 million live in urban and the remaining in rural areas. About 31 million people are estimated to be the carriers of mf and over 23 million suffer from filarial disease manifestations in India^[2]. Due to the absence of an effective vaccine and considerable side effects of the available chemotherapy, the main option available for controlling and preventing filariasis is the control of *Culex quinquefasciatus*^[3]. Chemical control is one of the several methods used in integrated vector control. Since the discovery of DDT as a

potent insecticide, chemical control has become the method of choice which necessitated the use of other types of insecticides as well^[4]. But due to reasons such as indiscriminate use of insecticides and natural adaptation, the vector has developed resistance to many of the available chemical insecticides in many places all over the world resulting in repeated disease outbreaks. This negative impact of chemical insecticides has prompted researchers to develop new environmentally compatible vector control methods using biocides.

Spinosad is a relatively new bioinsecticide, the active ingredient of which is derived from *Saccharopolyspora spinosa*, proving to be highly effective against mosquitoes. It kills susceptible species by causing rapid excitation of the insect nervous system. The discovery and characterization of this soil Actinomycete has presented a novel opportunity to develop a portfolio of progressive insect management tools. It has been used to control a variety of insect pests including fruitflies, sawflies, spider mites, fire ants and leaf miners^{[5][6]}. Along with Spinosad three other chemical insecticides were also used for the present investigation in order to determine the current susceptibility status of *Culex quinquefasciatus* as the susceptibility status varies from region to region and from time to time. It is in this context the present bioassay was carried out in the Vector Biology Research Lab of Zoology Department at University of Mysore.

2. Materials and Methods

2.1. Collection of Mosquito Larvae

Culex quinquefasciatus larvae were collected from breeding source of the University campus in Manasagangotri,

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with the help of a dipper. They were transferred to trays containing fresh water in order to get acclimatized to laboratory conditions. The late 3rd or early 4th instar larvae were separated and transferred to fresh water using Pasteur pipette just before the bioassay.

2.2. Larval Bioassay

The standard WHO procedure was followed for determining the larval susceptibility[7]. Accordingly the larvae were subjected to different concentrations of insecticides whose stock solutions were prepared by using distilled water as the solvent. Deltamethrin stock solutions were prepared in absolute alcohol and subsequent dilutions were made as per requirements. Test concentrations were prepared by adding 1ml of insecticide containing solution to 249ml of dechlorinated water in 500ml capacity beaker. It was stirred vigorously for 30 seconds with a glass rod. For the control, 1ml of distilled water or absolute alcohol as required was added to 249ml of dechlorinated water instead of insecticide. To each of the beakers containing different test and control, 25 late 3rd or early 4th instar larvae were released with the help of a strainer. Mortality was observed after 24 hours and the data was established using Abbott's formula[8]. All the tests were carried out at room temperature of 26±2°C and relative humidity of 70±5%. At least two replicates were maintained for both test and control.

2.3. Data analysis

Larval counts were adjusted for the mortality in control, if present, employing Abbot's formula.

$$\frac{\% \text{ of corrected mortality} = \frac{\% \text{test mortality} - \% \text{control mortality} \times 100}{100 - \% \text{control mortality}}}$$

The LC₅₀ and LC₉₀ values for each insecticide were calculated by dosage mortality regression line using Probit analysis^[9].

3. Results

Data showing the LC₅₀ and LC₉₀ values along with the fiducial limits of all the insecticides employed is presented in Table 1. Deltamethrin in the concentration of 0.0002, 0.0003, 0.0005, 0.0009 and 0.0014 ppm produced an experimental mortality of 16.0%, 32.0%, 48.0%, 56.0% and 74.0% respectively with zero mortality in control. The LC₅₀ and LC₉₀ values were 0.00062 and 0.00338 ppm respectively. Similarly lambdacyhalothrin in the concentration of 0.000004, 0.000006, 0.000008, 0.00001 and 0.00004 ppm produced an experimental mortality of 16.0%, 30.0%, 41.3%, 62.08% and 98% respectively with zero mortality in control. The LC₅₀ and LC₉₀ values for this insecticide were 0.00001 and 0.00002 ppm respectively. Propoxur in the concentration of 0.00006, 0.00010, 0.00014, 0.00018 and 0.0002 ppm produced an experimental mortality of 8.0%, 16.0%, 56.0%, 72.0% and 92.0% respectively with zero mortality in control.

The LC₅₀ and LC₉₀ values here were 0.00013 and 0.00023 ppm respectively. Similarly Spinosad in the concentration of 0.0000009, 0.000001, 0.000002, 0.000003 and 0.000004 ppm produced an experimental mortality of 8.33%, 32.0%, 40.0%, 76.0% and 100% respectively with zero mortality in control. The LC₅₀ and LC₉₀ values were 0.000002 and 0.000004 ppm respectively. Figure 1 depicts the percentage mortality of *Cx. quinquefasciatus* larvae after treating with deltamethrin, lambdacyhalothrin, propoxur and spinosad shows a linear exponential curve.

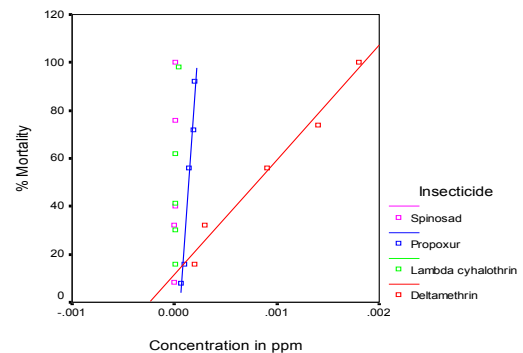


Figure 1. Regression profile indicating % mortality of *Cx. quinquefasciatus* larvae after treating with Deltamethrin, Lambdacyhalothrin, Propoxur and Spinosad

4. Discussion

Depending on the LC₅₀ and LC₉₀ values of insecticides employed for the present study it can be inferred that bioinsecticide, Spinosad is highly effective as mosquito larvicide. It has the lowest LC₅₀ and LC₉₀ values compared to that of other insecticides which shows that Spinosad is highly effective at low concentration (Table 1). The present results are in agreement with that of Darriet *et al*[2005] who have observed that Spinosad had lethal action on the larvae of mosquito species *Aedes aegypti*, *Culex quinquefasciatus* and *Anopheles gambiae* that were susceptible or resistant to pyrethroids, carbamates and organophosphates at Montpellier, France[10]. Liu *et al*[2004] have demonstrated that 3 strains of larvae of *Culex quinquefasciatus* which had great ability to develop resistance to different insecticides such as permethrin, deltamethrin, chlorpyrifos, fipronil and imidacloprid were highly susceptible to *Bacillus thuringiensis* variety *israelensis* and Spinosad in USA^[11]. Spinosad kills susceptible species by causing rapid excitation of the insect nervous system. As spinosad is ingested by the insect, it has very little effect on non-target predatory insects. No case of bioaccumulation in water or soil has been recorded so far.

The second most effective insecticide among the four was found to be Lambdacyhalothrin as per the LC₅₀ and LC₉₀ values (Table 1). Though lambdacyhalothrin, a synthetic pyrethroid, is efficient as a mosquito larvicide, mosquitoes have been known to develop resistance against this pyrethroid[12][13][14]. It is extremely poisonous to aquatic organisms and when used indiscriminately it affects the aquatic ecosystem.

Table 1. Differential efficacy of the insecticides tested against fourth instar larvae of *Cx. quinquefasciatus*

Insecticides	LC50 ppm (LCL-UCL)	LC90 ppm (LCL-UCL)	Slope±SE	Heterogeneity	Regression equation
Deltamethrin	0.00062 (0.00053-0.00074)	0.00338 (0.00235- 0.00590)	1.7446	1	1.7446 X ± 10.5920
Lambdacyhalothrin	0.00001 (0.00001-0.00001)	0.00002 (0.00002-0.00004)	3.1630	1	3.1630 X ± 20.9734
Propoxur	0.00013 (0.00009- 0.00018)	0.00023 (0.00017- 0.00074)	5.3765	7.66	5.3765 X ± 25.8699
Spinosad	0.000002 (0.000001- 0.000004)	0.000004 (0.000002- 0.000071)	3.7189	13.57	3.7189 X ± 26.3807

The third most effective insecticide among the four was found to be Propoxur (Table 1). This carbamate was initially found to be very effective and even after 10 years of house-spraying programme in Southern Iran in 1976-86 time period, mosquitoes had not developed resistance against this insecticide[15]. However it was found later that, mosquitoes had developed resistance against propoxur in several countries[16]. During an earlier study of response of *Culex fuscocephala* to six different insecticides at Mysore, it was found that the adult population of *Cx. fuscocephala* expressed least susceptibility to both propoxur (0.1%) and DDT (4%) in terms of LT₅₀ when compared to other insecticides used in this study. The LT₅₀ value for propoxur was found to be 110.6254 minutes while that of cyfluthrin, deltamethrin, permethrin, dieldrin and DDT were found to be 4.6999, 16.0118, 12.4859, 25.9535 and 69.2131 minutes respectively[17]. It is moderately toxic to fishes and aquatic birds. It acts as a carcinogen, blood toxicant, and reproductive toxicant and due to its cholinesterase inhibiting properties, a neurotoxicant as well. However it is not known to be bioaccumulative[18].

The least effective insecticide among the four was found to be Deltamethrin as it had registered higher LC₅₀ and LC₉₀ values (Table 1). Earlier reports indicate that deltamethrin, a synthetic pyrethroid, was among the most effective pyrethroids to control the mosquito population, but mosquitoes soon developed resistance towards it[19][20][21][22][23]. In an earlier report regarding the susceptibility of five species of mosquitoes to deltamethrin at Mysore, an LC₅₀ value of 0.00078 was observed for *Cx. quinquefasciatus*[24]. So according to the present study regarding the susceptibility of *Cx. quinquefasciatus* to deltamethrin (LC₅₀ value- 0.00062), it can be said that *Cx. quinquefasciatus* is more become susceptible to deltamethrin when compared to the earlier value of 0.00078 ppm in 2007. Deltamethrin is not mobile in the environment because of its strong adsorption on particles, its insolubility in water, and very low rates of application. However, it still presents risks to the ecosystem in which it is applied^[25].

5. Conclusions

Therefore the present investigation leads to the point that, Spinosad is a highly effective bioinsecticide against the larvae of mosquitoes and it may be valuable for the man-

agement of *Culex quinquefasciatus* especially in situations where local strains are highly resistant to other insecticides. Further experiments should be conducted in order to determine whether *Cx. quinquefasciatus* could develop resistance to this insecticide. Experiments should also be conducted regarding the susceptibility of other groups of mosquitoes against this insecticide so that the control of vectors will successfully help in the reduction of incidence of vector-borne diseases.

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REFERENCES

- [1] Taubes G, 1997. A mosquito bites back, New York Times Magazine. 40-46
- [2] Sabesan S, Vanamail P, Raju KHK, Jambulingam P, 2010. Lymphatic filariasis in India: Epidemiology and control measures. *Journal of Postgraduate Medicine*. 2010; 56(3):232-238
- [3] Maizels RM and Denham DA, 1992. Diethyl carbamazine immune-pharmacological interactions of an anti- filarial drug. *Parasitology*. 49-60
- [4] Anonymous, 1996. Operational manual on the application of insecticides for control of the mosquito vectors of malaria and other diseases. Geneva, WHO. 1996; 1000:20-37
- [5] Thompson WG and Trumbel JT, 1997. Effect of insecticides on Celery Insects. *Arthropoda management test*. 117
- [6] Kerns DL, 1996. Control of Lepidoptera larvae and leaf miner in lettuce, 1995. *Arthropod Management tests*. 117-118
- [7] Brown A W A and Pal R, 1971. Detection and measurement of resistance, *Insecticide resistance in Arthropods*, Second Edition, WHO, Geneva. 1971; 56-63

- [8] Abbott W.S, 1925. A method of computing the effectiveness of an insecticide. *J Econ Entomol.* 1925; 18:265
- [9] Finney .D.J., 1971. Probit analysis. Cambridge University Press. 58
- [10] Darriet F, Duchon S, Hougard JM 2005. Spinosad: a new larvicide against insecticide-resistant mosquito larvae. *J Am Mosq Control Assoc.* 2005 Dec; 21(4):495-496
- [11] Liu H, Cupp EW, Micher KM, Guo A, Liu N. 2004. Insecticide resistance and cross-resistance in Alabama and Florida strains of *Culex quinquefasciatus*. *J Med Entomol.* 2004 May; 41(3):408- 413
- [12] González T, Bisset JA, Díaz C, Rodríguez MM, Diéguez L, 1996. The evolution of resistance in a *Culex quinquefasciatus* strain starting from selection with the pyrethroid insecticide lambda-cyhalothrin. *Rev Cubana Med Trop.* 1996; 48(3):218-223
- [13] Bisset J, Rodriguez M, Soca A, Pasteur N, Raymond M. 1997. Cross-resistance to pyrethroid and organophosphorus insecticides in the southern house mosquito (Diptera:Culicidae) from Cuba. . *J Med Entomol.* 1997 Mar; 34(2):244-246
- [14] Santacoloma Varón L, Chaves Córdoba B, Brochero HL, 2010. Susceptibility of *Aedes aegypti* to DDT, deltamethrin, and lambda-cyhalothrin in Colombia. *Rev Panam Salud Publica.* 2010 Jan; 27(1):66-73
- [15] Manouchehri AV, Yaghoobi-Ershadi MR. 1988. Propoxur susceptibility test of *Anopheles stephensi* in southern Islamic Republic of Iran (1976-86). *J Am Mosq Control Assoc.* 1988 Jun; 4(2):159- 162
- [16] Elissa N, Mouchet J, Rivière F, Meunier JY, Yao K., 1994. Susceptibility of *Anopheles gambiae* to insecticides in the Ivory Coast. *Sante.* 1994 Mar-Apr; 4(2):95-99
- [17] Pushpalatha N, Vijayan VA. 1994. Adult population differences in response to six insecticides in *Culex fuscocephala* Theobald. *Southeast J Trop Ed Public Health.* 1994; 25(3):532-553
- [18] Hudson, R. H., Tucker, R. K. and Haegele, M. A., 1984. Handbook of Toxicity of Pesticides to Wildlife, U.S. Department of the Interior, Fish and Wildlife Service, Washington, DC. 1984; 153:3-48
- [19] Elissa N, Mouchet J, Rivière F, Meunier JY, Yao K., 1994. Susceptibility of *Anopheles gambiae* to insecticides in the Ivory Coast. *Sante.* 1994 Mar-Apr; 4(2):95-99
- [20] Urmila J, Vijayan VA, Ganesh KN, Gopalan N, Prakash S. 2001. Deltamethrin tolerance & associated cross resistance in *Aedes aegypti* from Mysore. *Indian J Med Res.* 2001 Mar; 113:103- 107
- [21] Gayathri V and Murthy PB, 2006. Reduced susceptibility to deltamethrin and kdr mutation in *Anopheles stephensi* Liston, a malaria vector in India. *J Am Mosq Control Assoc.* 2006 Dec; 22(4):678- 688
- [22] Urmila J and Vijayan VA, 2009. Biochemical characterization of deltamethrin resistance in a laboratory selected strain of *Aedes aegypti*. *Parasitol Res.* 2009; 10:1-5
- [23] Abate A, Hadis M. 2011. Susceptibility of *Anopheles gambiae* s.l. to DDT, malathion, permethrin and deltamethrin in Ethiopia. *Trop Med Int Health.* 2010Apr; 16(4):486- 491
- [24] Vijayan V. A., Sathish Kumar B. Y., Ganesh K. N., Urmila J., Fakoorziba M. R. and Makkapati A. K., 2007. Efficacy of piperonyl butoxide as a synergist with deltamethrin on five species of mosquitoes. *J. Commun. Dis.* 2007; 39 (3):159-163
- [25] Aldridge, W.N. 1990. An assessment of the Toxicological Properties of Pyrethroids and their Neurotoxicity. *Toxicology.* 1990; 21(2): 89-104