

# Application of Two Level Factorial Experimental Design in Assessing the *in vitro* Effect of Nutrient Supplementation on Growth of *Ceratocystis paradoxa* Isolated from Coconut Palm

Okogbenin E. A.<sup>1,\*</sup>, Okogbenin O. B.<sup>2</sup>, Obibuzor J. U.<sup>1</sup>, Emoghene A. O.<sup>3</sup>

<sup>1</sup>Technology and Product Development Department, Nigerian Institute for Oil palm Research (NIFOR). Edo State, Nigeria

<sup>2</sup>Plant pathology Division, Nigerian Institute for Oil palm Research (NIFOR). Edo State, Nigeria

<sup>3</sup>Department of Microbiology, Faculty of Life sciences, University of Benin, Benin City, Edo State, Nigeria

**Abstract** Mycelia growth of *C. paradoxa* was evaluated using two nutrient factors; Carbon source (glucose and fructose) and Nitrogen source (peptone and asparagine). A two by two level factorial design was used to ascertain the most important nutrient factor and any possible interaction between the factors. Half normal plot revealed that Nitrogen source was the most important ( $P < 0.05$ ) nutrient factor affecting the growth of *C. paradoxa* followed by the carbon source ( $P < 0.05$ ). It also revealed significant ( $P < 0.05$ ) interaction between both factors. A model was generated using Design- expert Software and statistical assumptions were validated by diagnosing residuals using normal plot for residuals and residuals versus predicted level plot. A combination of glucose and asparagine yielded the highest mycelia growth of *C. paradoxa* (this compared favorably with the industrially prepared PDA) while a combination of glucose and peptone yielded the least radial growth.

**Keywords** *Ceratocystis paradoxa*, Carbon source, Nitrogen source, Factorial Design, Design-expert

## 1. Introduction

The fungus, *Ceratocystis paradoxa* belongs to the Class *Ascomycota*, Order *Microascales* and is widely distributed around the world. The fungus is highly parasitic on many economic important palms (Elliott *et al.*, 2004) amongst which are *Cocos nucifera*, *Elaeis guineensis*, and *Phoenix dactylifera*. *C. paradoxa* is one of the major causes of black fruit rot in these palms (Wingfield *et al.*, 1993). This pathogen has been reported by Scot Nelson, 2005 to cause stem bleeding disease of coconut and in Nigeria, the fungus has been reported by Ojomo *et al.*, 1987 to be associated with bole rot coconut (*Cocos nucifera*) palms where it spreads rapidly through the soil between palms by root contact and probably by air-borne basidiospores. *In vitro* study of an organism is greatly influenced by the nutrient composition of the media as well as other factors. A wide range of media are used for isolation of different groups of fungi and these media are capable of influencing the vegetative growth, colony morphology and pigmentation of the organism (Kuhn and Ghannoum, 2003; Kumara and Rawal, 2008). In

laboratories, most fungi including *C. paradoxa* are isolated on specific culture medium for cultivation; preservation and microscopical examination depending upon the nutrient composition of the culture medium (Meletiadiis *et al.*, 2001). However, the requirements for mycelia growth of these fungi are generally less controlled. Nutrient type and availability has a pronounced effect on diagnostic characters as well as rapid mycelia growth of fungi. Hence, it is often necessary to use different media containing different nutrients while attempting to carry out *in vitro* studies on a fungus in culture since mycelia growth rate and sporulation on artificial media are important biological characteristics for fungi identification (St-Germain and Summerbell, 1996). Using a simple design of experiment (DOE) technique, this study was therefore undertaken to assess the effect of two nutrient factors; Carbon source and Nitrogen source on mycelia growth rate of *C. paradoxa* isolated from coconut palm showing symptoms of bole rot to ascertain the growth effects of the nutrient sources on mycelia growth in comparison to routinely used commercial Potato dextrose agar (PDA).

Two-level factorial design is an ideal statistically-based method used in identifying the vital few variables that significantly affect your process or product, leading to breakthrough improvements. It involves simultaneous adjustment of experimental factors at only two levels: high and low (Robert, 2009). The two-level design approach

\* Corresponding author:

okogbeninemma@yahoo.com (Okogbenin E. A.)

Published online at <http://journal.sapub.org/statistics>

Copyright © 2014 Scientific & Academic Publishing. All Rights Reserved

offers a parallel testing scheme that's much more efficient than one-factor-at-a-time. By restricting the tests to only two levels, you minimize the number of experiments and consequently save cost. The contrast between levels gives you the necessary driving force for product and process improvement (Robert, 2009). Aside from the issue of efficiency, two-level factorial designs can uncover critical interactions that wouldn't be revealed using one-factor-at-time experimentation or any other way (Robert, 2009).

## 2. Materials and Methods

### 2.1. Isolation and Identification of the Pathogen

In the laboratory, under aseptic conditions, a sterile scalpel was used to scrap necrotic tissues from the point of attachment of diseased coconut fruits showing characteristic symptoms of bole rot into sterile Petri dishes. The diseased tissues were cut into bits using a sterile blade to make thin films of about 5×5 mm in size and placed in sterile Petri dishes. Isolation of fungi was performed by first surface sterilizing the tissues following the method of Kinkel and Andrews, 1988. The bits of samples were surface sterilized in 70% sodium hypochlorite solution for one minute after which they were rinsed carefully in three changes of sterilized water and further blotted with sterile filter paper and allowed to dry in a sterile laminar flow chamber. Commercially available Potato dextrose agar (CMA Oxoid England) was prepared according to manufacturer's instructions and allowed to cool to a temperature of 28°C after which it was supplemented with 250mg/250ml of chloramphenicol so as to eliminate any bacterial contamination in the sample. Ten (10) mls each of the cooled PDA was dispensed into labeled petri plates and allowed to set after which the blotted tissues were transferred aseptically using a sterile wire loop into the PDA medium in plates. The plates were incubated at ambient conditions of light and temperature ( $30 \pm 2^\circ\text{C}$ ) and monitored on a daily basis until growth was observed (Abou-Zeid *et al.*, 2008). The pathogen isolated was identified by morphological and microscopic observations using Texts from Common wealth Mycological Institute (CMI) as *Ceratocystis sp.* The Culture was further sent to CABI laboratories, Surrey, London where it was confirmed to be *Ceratocystis paradoxa* (*Chalara* anamorph) with IMI NO. 501888. The identification was carried out using ITS DNA sequencing analysis. The sequence obtained showed 99.8-100% identity to multiple ITS sequences described from *Ceratocystis paradoxa*, including sequences reported in peer-reviewed literature. The isolate was stored in PDA slants and stored for further use.

### 2.2. Preparation of Culture Media

The laboratory prepared potato medium was prepared as thus; One hundred (100g) grams of fresh Irish potato was weighed after peeling and placed in a 500ml Erlenmeyer

flasks and boiled for one hour (). The cooked potatoes were mashed and squeezed and passed through muslin cloth to get a fine juice which was made up to 500ml using sterile distilled water and 10grams of agar powder (CMA Oxoid England) was added to the solution as a solidifying agent.

The carbon and nitrogen supplemented media were prepared by weighing 0.4grams each of the following combination into four 100ml Erlenmeyer flasks labeled; Glucose + Peptone (G+P), Glucose + Asparagine (G+A), Fructose + Peptone (F+P) and Fructose + Asparagine (F+A). After weighing, 40mls each of extracted potato juice containing agar was poured into each of the flask; while a fifth 100ml Erlenmeyer flasks contained 40mls of the extracted potato juice without any nitrogen or carbon source and labeled PDA(−) and a sixth 100ml Erlenmeyer flasks contained 40mls of the commercially prepared PDA labeled (PDA). The flasks were then plugged with sterile cotton wool and covered with foil paper. All the media were sterilized using a Gallenkamp autoclave for 15 minutes at a temperature of  $121^\circ\text{C}$  at a pressure of 15 Psi. After sterilization, each medium was allowed to cool after which, six (6) 10ml sterile syringe were labeled accordingly and used to dispense each medium unto the sterile, labeled Petri dishes in the laminar flow chamber and allowed to solidify.

### 2.3. Inoculation of Media

Using a 5mm sterile cork borer, a mycelia disc cut was made on a three day-old culture and the organism was aseptically inoculated unto the centre of the petriplates containing the solidified medium in an inverted position in order to achieve maximum contact of the mycelium with the culture medium. The position of the disc was marked on the base of the dish with a marker pen and two perpendicular lines passing through the centre of the disc were marked to be used as references for recording growth. The cultures were incubated at room temperature ( $28 \pm 2^\circ\text{C}$ ) and the experiment was carried out in triplicates. The commercial available PDA (CMA Oxoid England) and the PDA (−) were used as control for comparing the growth rate of the fungus with respect to those containing the nitrogen source. The experiment was monitored for a three day period as the organism grows rapidly.

### Determination of mycelia growth:

Mycelial growth was determined for each medium by measuring the radial growth of the fungus from the perpendicular lines drawn on the reverse side of the plate at the end of three days using a pair of divider and a meter rule. The results were noted and recorded.

## 3. Results and Discussion

A 2 by 2 full factorial design in triplicate, resulting in a total of 12 experimental runs, was carried out with the mycelia growth of *C. paradoxa* measured randomly as shown in table 1. Both positive control (commercial media,

PDA+) and negative control (Potato media with no supplementation) was set up alongside.

**Table 1.** Random measurement of radial growth of *C. paradoxa* in supplemented media

S/N	Carbon Source	Nitrogen Source	Average Mycelia Growth (mm)
1	Fructose	asparagine	77.00
2	glucose	peptone	62.80
3	Glucose	peptone	62.80
4	fructose	asparagine	77.00
5	fructose	asparagine	82.30
6	glucose	peptone	62.00
7	fructose	peptone	69.60
8	glucose	asparagine	84.30
9	glucose	asparagine	83.300
10	fructose	peptone	66.30
11	glucose	asparagine	86.00
12	fructose	peptone	65.30

**Table 2.** Mean radial growth of *C. paradoxa* in commercial and formulated media preparation

Media	Radial growth (mm)
PDA-	52.43 $\pm$ 1.37 <sup>b</sup>
PDA+	86.87 $\pm$ 0.57 <sup>a</sup>
G+A	84.53 $\pm$ 0.79 <sup>a</sup>
G+P	62.53 $\pm$ 0.27 <sup>d</sup>
F+A	78.77 $\pm$ 1.77 <sup>c</sup>
F+P	67.07 $\pm$ 1.30 <sup>c</sup>

Radial growth are mean values of triplicates  $\pm$  SEM. same alphabet are not significantly different at  $P > 0.05$

Table 2 shows mean values of radial growth of Potato supplemented media (G+A, G+P, F+A and F+P),

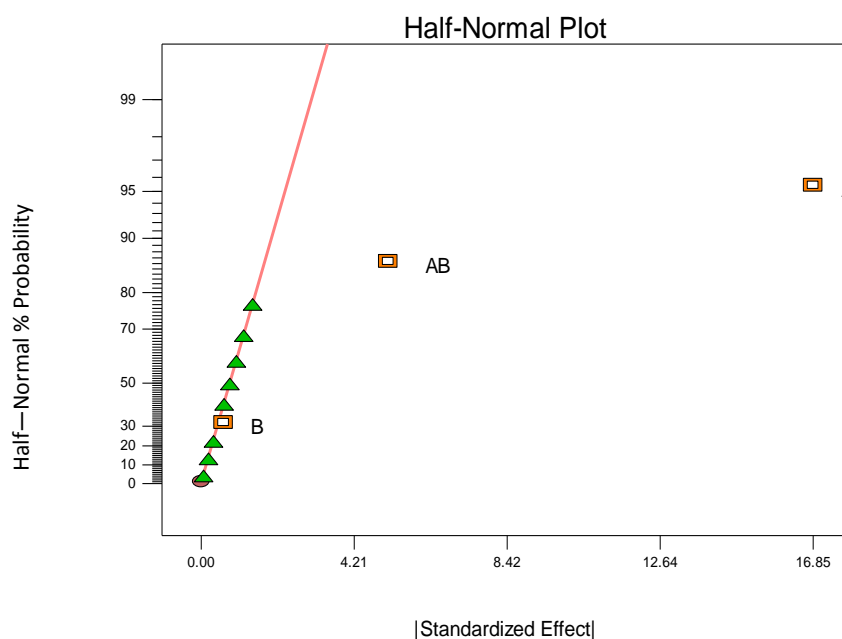
commercially prepared Potato dextrose agar (PDA+) and non supplemented potato (PDA-). It was clearly observed that supplementation of PDA-improved radial growth significantly ( $p < 0.05$ ), while mycelia growth on G+A supplemented medium differed non-significantly ( $p > 0.05$ ) from mycelia growth on PDA+. Both media supported the radial growth better than G+P, F+A and F+P supplemented media.

### 3.1. Half Normal Plots

The half normal plot is a graphical tool used to identify which experiment factors have significant effect on the response (Shari Kraber, 2013). This plot shows the magnitude of the experiment's effects as "Standardized Effects", ordered in increasing magnitude, along the x-axis. The Standardized Effect for a factor is the difference of the average response variable over "high" factor levels minus the average response over the "low" factor levels. The values on the y-axis are given by the idealized expected values for the number of effect, ranked by increasing value, if drawn from a half-normal distribution. Figure 1 shows the half normal plot for the effect of nutrient sources on the *in vitro* growth of *C. paradoxa*. It clearly shows that Nitrogen sources used in this study as well as the interaction (only detected by factorial experiments) between the carbon and nitrogen sources used, are significant with the former being the most important factor. These single factor and interaction obviously failed the "pencil test" as they do not fall in line with the other points. For factor B (carbon source), it appears that the switch from fructose to glucose does not have any significant effect on the growth of *C. paradoxa* as it passes the pencil test with the other points resulting from the error from replicates.

Design-Ease® Software  
Radii Diameter

▲ Error from replicates  
A: Nitrogen Source  
B: Carbon Source  
■ Positive Effects  
■ Negative Effects

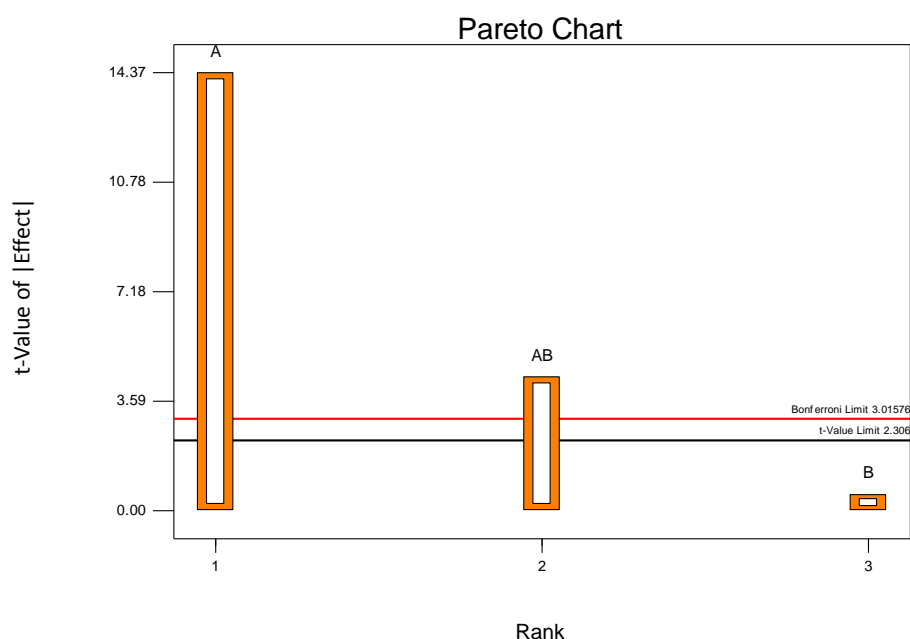


**Figure 1.** Half normal plots showing effects of carbon and nitrogen sources on *C. paradoxa*

Figure 2 below offers a simpler view of the relative effect via an ordered bar graph called a “Pareto chart”. The Pareto chart is useful for showing the relative size of effects especially to non-statisticians. The Pareto chart for this study, also clearly shows that the Nitrogen factor (factor A) and its interaction with the carbon (AB interaction) exceeds the t- value limit of 2.306 (the P-value was set at 0.05). Also it exceeds another limit called the Bonferroni limits which cuts into half the p value of 0.05 to a much stricter level of 0.025.

Design-Ease® Software  
Radii Diameter

A: Nitrogen Source  
B: Carbon Source  
■ Positive Effects  
■ Negative Effects



**Figure 2.** Pareto chart showing the effect of carbon and nitrogen media supplementation on growth of *C. paradoxa*

### 3.2. Interaction Plots

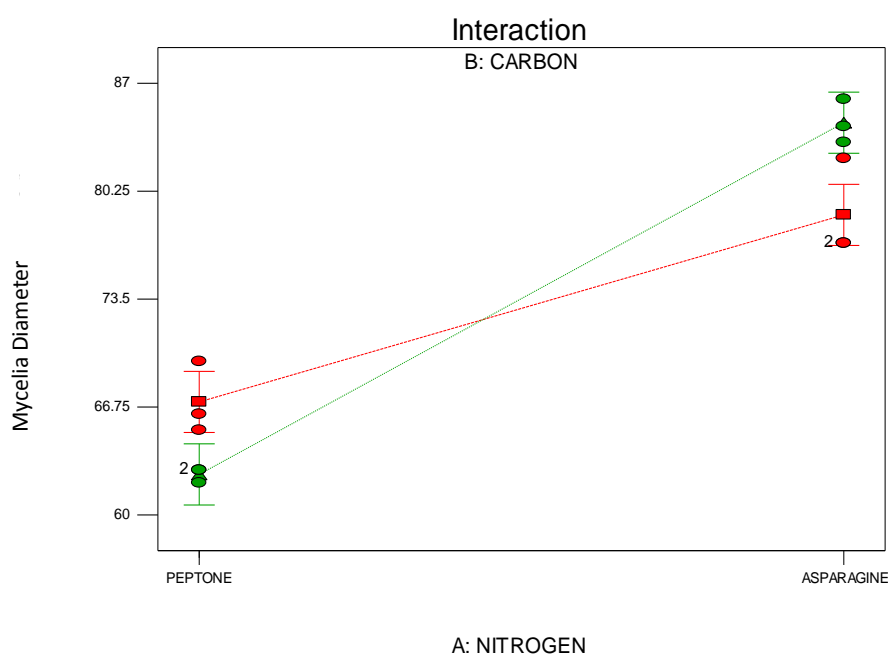
An interaction occurs when one factor affects the results differently depending on the second factor. It is detected using an interaction plot. The presence of interaction in the plot is usually associated with non-parallel lines (unequal slopes) as shown in Figure 3. The interaction plot shown below reveals that when the carbon source is fructose the optimum radial growth is when the nitrogen source is peptone while when the carbon source is glucose the optimum radial growth is when the nitrogen source is asparagine.

Design-Ease® Software

Mycelia Diameter

● Design Points  
■ B1 FRUCTOSE  
▲ B2 GLUCOSE

X1 = A: NITROGEN  
X2 = B: CARBON



**Figure 3.** Interaction plot showing the interactive effect between sources of carbon and sources nitrogen on growth of *C. paradoxa*

**Table 3.** Analysis of variance (ANOVA) table

Source	Sum of squares	DF	Mean Square	F value	p-value prob>F	
Model	932.48	3	310.83	75.34	<0.001	significant
A (nitrogen source)	851.77	1	851.77	206.45	<0.001	Significant
B (Carbon source)	1.14	1	1.14	0.28	0.6133	
AB	79.57	1	79.57	19.29	0.0023	Significant
Pure error	33.01	8	4.13			
Cor total	965.48	11				

However, to protect against spurious outcomes, analysis of variance was carried out to verify conclusion drawn from the interaction plot, half normal plots and Pareto charts. Table three shows ANOVA for Radii growth of *C. paradoxa*. It is observed that a highly significant model ( $P < 0.0001$ ) was generated with factor A (Nitrogen source) contributing about 88.22% to the model while it interaction (AB) contributes about 8.24% to the model. The change from glucose to fructose had an insignificant effect on microbial growth of *C. paradoxa* and this factor contributes 0.12% to the model generated.

R-Squared            0.9658  
 Adj R-Squared    0.9530  
 Pred R-Squared   0.9231  
 Adeq Precision   18.760

The coefficient determination, R-squared approached 1 (0.9658) and "Predicted R-Squared" of 0.9231 is in reasonable agreement with the "Adjusted R-Squared" of 0.9530 (both should not differ from each other by more than 0.2) (Shari Kraber, 2013). Adeq. Precision measures the signal to noise ratio and a ratio greater than 4 is desirable (Anderson and Patrick, 2000) The ratio achieved for this work was 18.760 which indicates an adequate signal and as such the model can be used to navigate the design space. The model generated in both coded and actual factors from the analysis is as thus:

#### Final Equation in Terms of Coded Factors:

Radii Diameter = + 73.23 + 8.42 \* A + 0.31 \* B + 2.58 \* A \* B

#### Final Equation in Terms of Actual Factors:

Nitrogen Source Peptone  
 Carbon Source      Fructose  
 Radii Diameter      =  
 +67.06667

Nitrogen Source    Asparagine  
 Carbon Source      Fructose  
 Radii Diameter      =  
 +78.76667

Nitrogen Source    Peptone  
 Carbon Source      Glucose  
 Radii Diameter      =  
 +62.53333

Nitrogen Source    Asparagine  
 Carbon Source      Glucose  
 Radii Diameter      =  
 +84.53333

### 3.3. Model Validation Plots

The main aim of a model is to develop a good predictive relationship between the dependent variable (response; mycelia growth) and the independent variable (effect; carbon and nitrogen sources). Diagnosing the residuals, which is the difference between predicted and actual values, plays a vital role in finding and validating such a relationship. Carefully looking at the residuals can tell us whether our assumptions are reasonable and our choice of model is appropriate. If assumptions are satisfied, residuals should vary randomly around zero and the spread of the residuals should be about the same throughout the plot (no systematic patterns.) (Anderson and Patrick, 2007).

This model generated is generally fitted under two important assumptions:

- Normal distribution of residuals
- Constant variance of residuals (homoscedasticity)

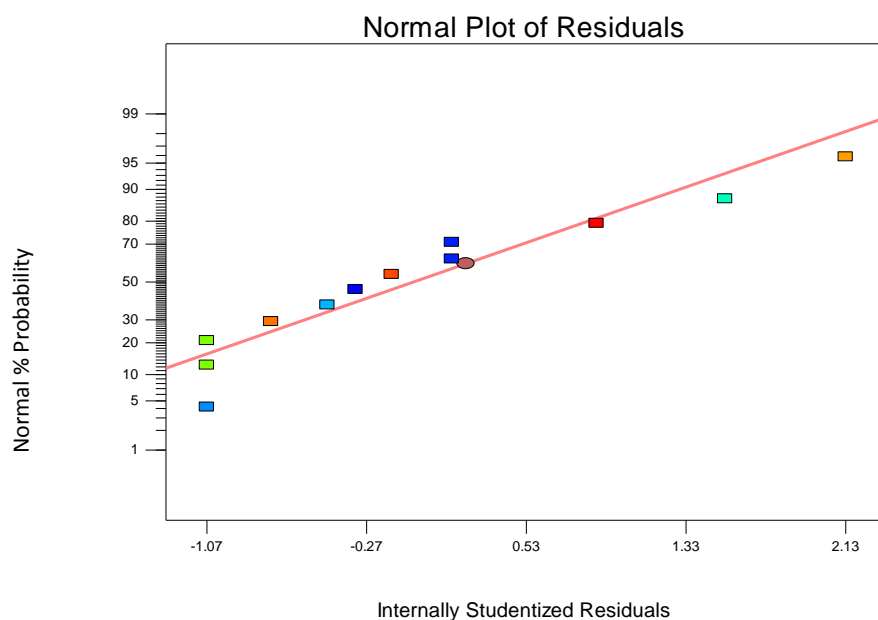
Departure from these assumptions usually means that the residuals contain structure that is not accounted for in the model (Anderson and Patrick, 2000). Consequently, certain inferences and predictions from the analysis may be unreliable and inaccurate. In order to validate these statistical assumptions of normal distribution of residuals, (the difference between predicted and actual values) as well as their independence with constant variance, the two plots were generated.

- Normal probability plot of the studentized residuals to check for normality of residuals.
- Studentized residuals versus predicted values to check for constant error.

As shown in figure 4, the deviations from linear are very minor, so it supports the assumption of normality. When clearly non-linear patterns such as an "S" shape is observed, a response transformation should be considered. However response transformation is dependent on the ratio of maximum response to minimum response. Transformation becomes necessary once this ratio reaches 10 and above. For generated data, the maximum response is 86 while minimum is 62 a ratio of 1.3871, hence data transformation is not necessary.

Design-Ease® Software  
Mycelia Diameter

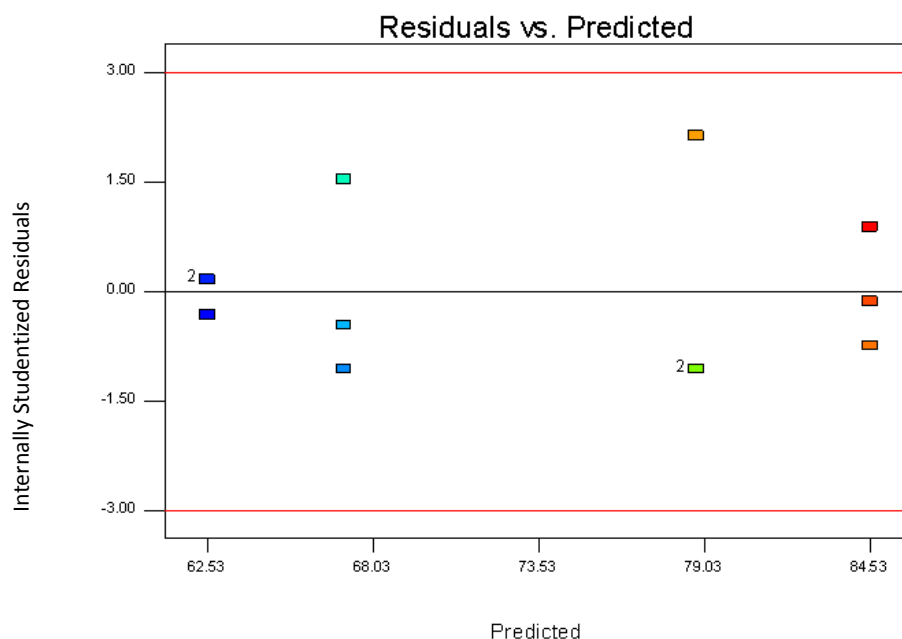
Color points by value of  
Mycelia Diameter:



**Figure 4.** Normal plot of residual for mycelia growth of *C. paradoxa*

Design-Ease® Software  
Mycelia Diameter

Color points by value of  
Mycelia Diameter:



**Figure 5.** Residual versus Predicted response plot for mycelia growth of *C. paradoxa*

In the residuals versus predicted response plot shown in figure 5 above, it is safe to say that there is no definite increase in residuals with predicted level, which supports the underlying statistical assumption of constant variance. Ideally, the vertical spread of data will be approximately the same from left to right. Nonconstant variance is often associated with a megaphone (<) pattern, where the residuals increase with predicted level.

## 4. Conclusions

The application of two level full factorial (2FL) design in

this study was able to reveal that the change from asparagine to peptone is the most important factor that affected the growth of *C. paradoxa* and a change from glucose to fructose had a non significant effect at  $P > 0.05$ . This was achieved with both factors varied simultaneously and with minimum amount of experimental runs when compared with one factor at a time (OFAT) experiment. Also, 2FL design was able to reveal that an interaction between the carbon sources and nitrogen exist which would have not been possible to detect using OFAT. Several validation plots were generated which are simple to interpret even to non-statisticians. Consequently, the use of this statistical experimental tool should be

encouraged by all those interested in improving any biological and chemical processes.

## REFERENCES

- [1] Abou-Zeid A.M., Altalhi, A. D. and Abd El-Fattah, R.I. (2008). Fungal Control of Pathogenic Fungi Isolated From Some Wild Plants in Taif Governorate, Saudi Arabia. *Mal .J. Microbiol.* 4(1): 30-39.
- [2] Anderson Mark and Patrick Whitcomb (2007). Using Graphical Diagnostics to Deal with Bad Experimental Data. *Quality Engineering.* 19: 2.
- [3] Anderson Mark and Patrick Whitcomb. DOE Simplified-Practical Tools for Effective Experimentation second edition. Pp 49-56. New York: Productivity, Inc 2000.
- [4] Elliott, M.L., T.K. Broschat, J.Y. Uchida, and G.W. Simone (eds.). 2004. Compendium of Ornamental palm diseases and disorders. *American Phytopathological Society, Minneapolis, Minn.* 71 p.
- [5] Ojomo, E.E. and E.N. Ekpo. (1987). Identification of Major Diseases and Pests of Coconut and their Control Measures. Nigerian Institute for Oil Palm Research (NIFOR) *Annual Report* 24:107-108.
- [6] Kinkel, L.L and Andrews, J.H. (1988). Disinfestation of living leaves by hydrogen peroxide. *Trans. Br. Mycol. Soc.* 91: 523-528.
- [7] Robert, W.M (2009) Introduction to Full Factorial Designs with Two-Level Factors. A Comprehensive Guide to Factorial Two-Level Experimentation. *Springer sci.* Pp 3-26.
- [8] Shari Kraber. Introduction to Response surface methods; Part-1-Central composite design. Presentation posted at [www.statease.com/webinar.html](http://www.statease.com/webinar.html). Accessed november 2013.
- [9] Wingfield, M. J., Seifert K.A., Webber, J.F. (1993). *Ceratocystis* and *Ophiostoma*: Taxonomy, Ecology and pathogenicity. APS press, St. Pal, M.N. 293 pp.
- [10] Scot Nelson, (2005). Stem bleeding of Coconut Palm. Cooperate extension service, College of Tropical Agriculture and Human Resources, University of Hawai'i, Manoa. *Plant Disease.* PD-30.
- [11] Kuhn, D.M and Ghonnoumm, M.A. (2003). Indoor mold, toxigenic fungi, and *Stachybotrys chartarum*: Infectious disease perspective. *Clin. Microbiol. Rev.*, 16(1): 144-172.
- [12] Kumara, K.L.W and Rawal, R.D. (2008). Influence of carbon, nitrogen, temperature and pH on the growth and sporulation of some Indian isolates of *Colletotrichum gloeosporioides* causing anthracnose disease of papaya (*Carrica papaya* L). *Trop. Agric. Res. Ext.*, 11: 7- 12.
- [13] Meletiadiis, J., Meis, JFGM., Mouton, J.W. and Verweij, P.E. (2001). Analysis of growth characteristics of filamentous fungi in different nutrient media. *J. Clin. Microbiol.*, 39(2): 478-484.
- [14] G. Sharma and R. R. Pandey. (2010). Influence of culture media on growth, colony character and sporulation of fungi isolated from decaying vegetable wastes. *Journal of Yeast and Fungal Research.* 1(8): 157 – 164.