

# Application of Phyto-Remediation (Sunflower and Vetiver Grass) on Crude Oil Spilled Soil Cultivated to Jute Mallow (*Corchorus Olitorius* L.)

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**Abstract** This study assessed the potential phytoremedial use of *Tithonia diversifolia* (Mexican sunflower) and *Vetiveria nigrifolia* (Vetiver Grass) on bioremediation of crude oil affected soils as an option that rendered harmless various contaminants using natural biological activity. It relatively low-cost, low techniques with high public acceptance and can often be carried out on site. The experiment consisted of three treatments; non-remediated soil (A), sunflower (B) and vetiver (C) remediated soils. The treatments in three replicates were laid out in a completely randomized design (CRD) layout. Soil physical conditions were significantly best under vetiver treated soils. Nevertheless, heavy metals concentration after the experiment was in the order of treatments A>B>C. Application of sunflower significantly ( $p<0.05$ ) reduced the total hydrocarbon content in the soils by 25.59%, while vetiver grass had removal efficiency greater than 41%. Whereas the results of HUB/THBC ratio is in excess of 1% which tends to indicate recent crude oil input into the environment, the differences among the treatments were not significant. The percentage increment in leaf dry weight from 8 to 16 WAP was significantly lower (11%) in pots grown without treatment than those treated; soils treated with sunflower, the percent increment in biomass dry weight from 8 to 16 WAP was 84% and 139% for those treated with vetiver grass for the same period. However, the degree of toxicity on the tested plant was attributed to the direct contact of the toxic volatiles and water soluble hydrocarbons in spilled soil that might have penetrated the tissues of the tender plant.

**Keywords** Oil spill, Phyto-technology, Vetiver, Cleanup, Sunflower, Hydrocarbon, Plant

## 1. Background

Oil contaminated soils are of environmental concern because they are unsuitable for agricultural and recreational use and are potential sources for surface and ground water contamination[1] Petroleum creates unsatisfactory conditions for plant growth through a number of processes; (i) oil could displace air from soil pore spaces, (ii) an increase in the demand for oxygen brought about by activity of oil-decomposing microorganisms[2] and (iii) petroleum hydrocarbon creates hydrophobic environment, which limits water absorption to plant roots[3].

There are several physical-chemical technologies for the treatment of soils contaminated with organic and hazardous materials such as petroleum hydrocarbons. They include vapour extraction, stabilization, solidification, soil flushing, soil washing, thermal desorption, and incineration[4, 5]. These methods have some drawbacks. Most of these

techniques are expensive to implement at full scale and require continuous monitoring and control for optimum performance. Some of the techniques simply move the contamination elsewhere and may create significant risks in the excavation, handling and transport of hazardous materials. Also, it is very difficult and increasingly expensive to find new landfill sites for the final disposal of the material. Additionally, they do not usually result in a complete destruction of the contaminants.

A better approach than these methods or techniques is to completely destroy the pollutants if possible, or at least to transform them to innocuous substances. Application of Phyto-techniques is alternative methods for the treatment of contaminated soil. These techniques are economically and politically attractive and have shown promising results in the treatment of soil contaminated with organic compounds, particularly with petroleum hydrocarbons[6].

Phyto-remediation is an option that offers the possibility to destroy or render harmless various contaminants using natural biological activity. As such, it uses relatively low-cost, low techniques, which generally have a high public acceptance and can often be carried out on site. Phyto-remediation of Mexican sunflower processes have

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shown to be effective methods that stimulate the biodegradation in contaminated soils[7] Harder[8], estimated that phyto-remediation accounts for 5 to 10 percent of all pollution treatment and has been used successfully in cleaning up the illegal dumping of used engine oil. Lee *et al.*,[9] reported that oil cleanup methods, especially bioremediation strategies, must take into consideration the local fauna, flora and environmental conditions if they are to make a significant impact.

The focus of this study was to assess the effectiveness of sunflower and vetiver remediation technology in cleaning up crude oil contaminated soil for optimum plant production.

## 2. Materials and Methods

### 2.1. Site Description

The study was conducted near the Department of Forestry Arboretum, University of Uyo, Akwa Ibom State, Nigeria. Uyo and its environs, is located between latitudes  $4^{\circ} 30'$  and  $5^{\circ} 31'N$  and longitudes  $7^{\circ} 31'$  and  $8^{\circ} 20' E$ [10]. The state has an estimated area of 8, 412 km<sup>3</sup>. It is characterized by two seasons, a wet season that last for nine (9) months (April –October) and dry season (November-March). The annual rainfall ranges from 2000-3000 mm, while annual temperature varies between 26 °C and 28 °C . Relative humidity is high varying from 75-95% with the highest and lowest values in July and January respectively.

### 2.2. Experiment Layout and Greenhouse Study



**Figure 1.** Showing plants grown on the crude oil spilled contaminated soils (A), cured soil with sunflower (B) and cured soils with vetiver (C) soils

The green house experiment consists of three treatments (non-remediated soil, remediated with sunflower and remediated with vetiver) (Fig. 1) replicated three times within completely randomized design (CRD) layout under Jute mallow cultivation. The crude contaminated soil was evenly mixed separately with freshly cut and finely chopped shoots of sunflower and vetiver grass, equivalent to 2.5 t ha<sup>-1</sup>, were tied in bags and allowed to cure for three (3) months. One (1) kilogram of crude oil contaminated and cured loamy soils were packed into perforated plastic buckets where *Corchorus olitorius* plant used as test plant was transplanted after three (3) weeks in the nursery. The treatments were designated A, B and C for contaminated soil, and cured soils with sunflower and cured soil with vetiver grass respectively. The perforated plastic buckets containing the treatments were subsequently watered every 24 hours until the plant was fully established (10 days after transplanting). This experiment was repeated for three consecutive times in order to validate the results.

### 2.3. Measurement of Growth Characteristics

When the plants were sufficiently established, six plants were randomly selected from each treatment and tagged for the measurement of growth characteristics at 4, 8 and 16 weeks after planting (WAP) as follows:

#### 2.3.1. Plant Height (cm)

This was taken as the height of the *Corchorus olitorius* plant measured to the nearest centimeter from the base to top. The mean height from the randomly selected plants was taken as the score for each treatment.

#### 2.3.2. Number of Leaves Per Plant

The number of leaves per plant was determined by counting the leaves from plants used to compute the score for each treatment.

#### 2.3.3. Biomass (dry matter) Yield Per Treatment (g)

Biomass was determined by harvesting the leaf and stem materials after full establishment and oven-dried at 70 °C for 3 days.

### 2.4. Soil Sample Preparation and Analyses

All samples were air dried and passed through a 2 mm sieve for routine analyses and 0.5 mm sieve for Organic C and Total N determinations.

#### 2.4.1. Determination of Physico- chemical Properties

Particle-size distribution was determined by hydrometer method[11]. Soil pH was measured in distilled water (1:2.5 soil:water) with the use of glass electrode pH meter. Available phosphorus was determined using bicarbonate extraction, with acid reductant. The exchangeable cations (Calcium, Ca; Magnesium, Mg; Potassium, K; and Sodium, Na) in the soil were determined by first extracting the soil sediment with 1M NH<sub>4</sub> OAc (ammonium acetate)

solution[12]. Soil Organic Carbon (SOC) was determined by loss-on-ignition and the standard Van Bemmelen factor (1.724) was used for conversion of SOC into organic matter content[13]. Nitrogen was determined in form of ammonium ( $\text{NH}_4\text{-N}$ ) and nitrate ( $\text{NO}_3\text{-N}$ ) by absorbance measurement as described by Nelson[14]. Base saturation (BS) was expressed as the fraction of the negative binding sites occupied by exchangeable cations. It was calculated by summing together the levels of Ca, Mg, K, and Na found in the soil, then expressing this sum as a percentage of the ECEC value as follows:

$$\text{BS} = \frac{100 (\text{Ca} + \text{Mg} + \text{K} + \text{Na})}{\text{ECEC}}$$

## 2.5. Microbial Enumeration of Bacteria and Fungi

Numbers of viable bacteria and fungi were estimated by the plate count technique. Ten grammes (10g) of 2-mm sieved soil was added to 90ml of sterile distilled water and shaken for 30 minutes. Serial dilutions were prepared up to  $10^6$ . Total viable counts of culturable aerobic heterotrophic bacteria were obtained by surface plating 1ml of the serial dilution on to sterile nutrient agar while fungi counts were obtained by surface plating 1ml of the serial dilution on to potato dextrose agar (PDA). Culture plates were incubated at room temperature ( $28 \pm 2^\circ\text{C}$ ) for 48 hours. Plates yielding counts of 30 - 300 colonies were chosen and the counts obtained were multiplied by the dilution factor and expressed as CFU/g of soil. The morphological characterization (colour and size) of each bacteria isolates and colony characteristics (shape, elevation, edge, surface and pigmentation) were done. Gram staining was also done to determine the reactions of the bacteria to Gram reaction. Fungi identification test was also carried out.

## 2.6. Statistical Analyses

All data collected were statistically analysed using analysis of variance (ANOVA) procedure to assess the effect of phyto technology on contaminated soil remediation and plant growth using GLM procedure (SPSS soft ware, Version 17). The means were compared using Fisher's least significant difference (LSD) test. All tests of significance were made with probability value of 0.05.

# 3. Results and Discussion

## 3.1. Effects of Phyto-remediation on Soil Properties

### 3.1.1. Soil Physical Conditions

Soil texture which is considered to have profound effect on many other properties was considered among the physical properties. As shown in Table 1, application of phyto-technology had no effect on the textural class (loamy sand) of the soil, rather the distribution of clay was significantly improved with in the treatment containing vetiver grass (treatment C).

### 3.1.2. Soil pH (soil reaction)

Soil pH is one of the most common and important measurements in standard soil analyses. Many soil chemical and biological reactions are controlled by the pH of soil solution in equilibrium with the soil particle surfaces. Soil acidity is a major problem in the production of arable crops in the humid tropical soil of which this location is one. Yields of many crops are highly reduced when the pH values are low (strongly acidic  $< 5.0$ )[15]. The results of soil reaction (Table 1) revealed only slight differences in pH among the treatments. The acidity level of the contaminated soil changed from slightly acidic (6.3) to moderately acidic (5.6) when the soil was amended with treatment C, but remained slightly acidic with treatment B (6.5). Obviously, phyto technology reduced the acidic level by 11%, and the soils under this condition can support the optimum performance of most of the arable crops.

**Table 1.** Means of some physico-chemical properties of the experimental soils

Parameters	Units	A	B	C	LSD (0.05)
Texture		Loamy sand	Loamy sand	Loamy sand	
Sand	g kg <sup>-1</sup>	790	790	710	46.19
Silt	g kg	108	88	148	30.55
Clay	g kg	102	122	142	20.00
pH		6.3	6.5	5.6	0.47
THC	mg/kg	21	16.80	13.40	1.30
Oil & grease	mg/kg	1.20	0.27	0.59	0.0646
Base saturation	%	97	98	98	0.58
Total N	mgkg <sup>-1</sup>	132.40	97.30	70.70	30.95
phosphorus	mgkg <sup>-1</sup>	1.31	1.00	1.25	0.16
<b>Exchangeable cations (c mol<sup>-1</sup>)</b>					
K		155.00	100.10	97.50	32.47
Mg		1.00	1.23	2.45	0.78
Ca		3.70	4.68	5.31	0.81
Na		3.89	13.58	21.30	8.72
CEC		167.19	128.76	120.99	24.74

Where, A = untreated soil, B = treated soil with sunflower, C = treated soil with vetiver grass

### 3.1.3. Total Extractable Hydrocarbon Content (THC) and Oil and Grease in Experimental Soil

Organic compounds are complex and valuable mixture of compounds of which hydrocarbon is one. An occurrence of THC resulting from crude spilled is alien to soil and their presence affects exchange of oxygen and stains surface soil[16]. The mean concentration of THC was of 21 mgkg<sup>-1</sup> in contaminated soil (treatment A), 19.80 and 18.40 mgkg<sup>-1</sup> (treatment B and C respectively). Surprisingly, the concentration of THC was significantly higher in the treatment soils. These concentrations could be accounted for mainly from biogenic sources of wax and suberins from decay plants that became prominent when submerge in water during watering[17]. Therefore soils receiving treatments B and C are not contaminated by petroleum hydrocarbon.

Furthermore, amendment of crude affected soil with sunflower and vetiver reduced the stain THC values by 6 and 14% (removal efficiency of B and C treatments respectively). This shows that the soil was heavily polluted, as evident in the poor growth of *Corchorus olitorius* especially on the untreated soils. The values of oil and grease showed concentrations that varied from 0.07 to 1.28 mg kg<sup>-1</sup> with means values of 1.20, 0.27 and 0.59 mg kg<sup>-1</sup> for the respective treatments. Sunflower demonstrated superior reduction efficiency of 344% compared to control and vetiver treated soils.

### 3.1.4. Total Nitrogen (NH<sub>4</sub><sup>+</sup>, and NH<sub>3</sub>)

Total Nitrogen content of a soil gives an indication of the organic nitrogen present in the soil. Most of the Nitrogen in soils is in the organic form as only relatively small quantities occur in NH<sub>4</sub><sup>+</sup>, and NH<sub>3</sub>; the more available forms. The amount of available nitrogen in the soil is an indication of how suitable conditions in the soils are. A value below 0.1% total Nitrogen is considered low for soils (Isirimah *et al.*, 2003). Between the control and remediated soils, the concentration of Total N in the soil was very high (A=132.40, B=93.3 and C=70.70 mgkg<sup>-1</sup> respectively) The Total Nitrogen values of 0.15, 0.15 - 0.20, and greater than 2.0 mgkg<sup>-1</sup> are classified as low, medium, and high respectively [18]. Therefore, the results indicated very high concentration of total N for these soils.

### 3.1.5. Exchangeable Cations

The basic cations values in this experiment is not in consonance with the decreasing cation magnitude observed by of Oputa and Udo (1980), that is Ca<sup>2++</sup> > Mg<sup>2++</sup> > K<sup>+</sup> > Na<sup>+</sup>. The Potassium (K) and cation exchange capacity (CEC) level in the soil followed the decreasing order of Treatment A > B > C. The reverse was true for magnesium, calcium and sodium. In comparing the treatments, it was assumed that Mg, Ca and Na will increase significantly with incremental rate of the phyto application. At 0 t/ha (treatment A), Mg, Ca and Na contents were, 1.00, 3.70, and 3.89 cmolkg<sup>-1</sup>, respectively. Treatment B (sunflower), increased these cations to 1.23, 4.68 and 13.58 cmolkg<sup>-1</sup> respectively. In treatment C (vetiver), the cations further increased respectively to 2.45, 5.31 and 21.30 cmolkg<sup>-1</sup>. Although, highest CEC was observed in the control soil, the actual exchanges of cations in the soils' sorption site were hindered due to the presence of stain substances resulting from crude oil spillage. Hendrickx *et al.*[19], noted that it is crucial to understand the concentration of exchangeable cations as they occur in the area of environmental management as this affects the quality of soil resources. Most of the cations in soils that are available for plant uptake are induced by microbial activities in the soil. Thus CEC availability was also affected by soil environmental factors that affect microbial activities. Apart from stain substances in the contaminated soil, immobilization of magnesium and sodium was largely responsible for the development of

acidity in the soils.

This soils' pH varied from slightly to moderately acidic, an indication of low level of Na and Mg (treatment A) which may favour the solubility of Al and Mn thus reducing plant yield. Although Na deficiency has not been identified as a limitation in most of the arable crops production, but magnesium deficiency is common source. This effect of Na and magnesium noticed in the control soil may be due to the rise in the level of exchangeable Al. This according to Rolling *et al.*[20] usually occurs at low pH and affects crop growth and yield.

### 3.2. Heavy Metals (Traced elements)

The mineral elements originate from soil and are dissolved in water for plant roots' absorption, but those required in small quantity for optimum performance are regarded as traced elements. The concentration of these metals can however be increased to become potential pollutants if heavy metals containing waste products from industrial or domestic activities are introduced into the environment. Concern over the presence of heavy metals in an environment arises from the fact that they cannot be broken down into non toxic forms. Thus once aquatic ecosystems are contaminated by heavy metals; they remain a potential threat for many years[21]. These heavy metals include: Cu, Pb, Zn, Cd, Ba and Fe.

**Table 2.** Means of heavy metals, biological counts and THC of the experimental soils

Parameters	Treatments			
	A	B	C	LDS (0.05)
Heavy metals (mg/kg)				
<b>Copper</b>	3.12	2.13	1.14	0.990
<b>Lead</b>	0.11	0.10	<0.01	0.055
<b>Zinc</b>	25.30	15.00	8.60	8.426
<b>Cadmium</b>	0.01	<0.01	<0.01	0.058
<b>Barium</b>	0.82	0.54	0.43	0.201
<b>Iron</b>	2122	1810	831	673.605
microbial group				
(cfu/g)				
<b>THBC (x 10<sup>3</sup>)</b>	2.00	8.00	12.00	5.033
<b>THFC(x 10<sup>2</sup>)</b>	0.00	1.00	0.00	0.577
<b>HUB(x 10<sup>1</sup>)</b>	1.00	5.00	7.00	3.055
<b>HUF(x 10<sup>1</sup>)</b>	0.00	0.00	1.00	0.577
<b>HUB/THBC</b>	0.5	0.3	0.5	-

Copper is important for the reproductive growth and root metabolism. A major property affecting their availability is soil pH, a measure of soil acidity or alkalinity. Cu tends to be less available in soils with high pH. In this experiment, (Table 2), Copper concentrations averaged 3.12, 2.13 and 1.14 mg kg<sup>-1</sup> (treatments A, B and C respectively). The study showed 47.47 and 174% reduction of Cu concentration in soils treated with B and C respectively. The significantly high reduction with phyto technology suggests that large amount of copper remain relatively unstable even in the vetiver treated soil (treatment C). In this experiment, lead concentrations were 0.11, 0.10 and <0.01 mg kg<sup>-1</sup> for treatments A, B and C respectively. Increased in the

remediation materials to  $5 \text{ t ah}^{-1}$  reduced Pb to below detectable limit. Similarly, Zn, Cd, Ba and Fe were significantly reduced between the ranges of 90 to 155%.

### 3.3. Biological Characteristics

The microbial community in soil is important because of its relationship to soil fertility and the biochemical cycling of elements. Thus the need to enumerate and isolate major and minor members of the microbial community in soils. Micro organisms are predictable first component of biota in an ecosystem to demonstrate the effect of environmental pollution resulting from any contamination. As shown in Table 2, the soil microbes consisting mainly of fungi and bacteria substantially contribute to the recycling of nutrients within the system. Total heterotrophic bacterial counts (THBC) in untreated treatment (A) averaged  $2.0 \times 10^3 \text{ cfu/g}$  soil, but on amending the soil with treatment B (sunflower) it averaged  $8.0 \times 10^3 \text{ cfu/g}$ . In the vetiver treated soils (treatment C), THBC value was  $12.0 \times 10^3 \text{ cfu/g}$  was identified. Conversely, there were no heterotrophic fungi counts in the soils except in treatment B, with THFC of  $1.0 \times 10^2 \text{ cfu/g}$  soil. During the periods under investigation, the Counts for the Hydrocarbon utilizing bacteria (HUB) and fungi (HUF) were equally low among the treatments. Interestingly, the same counts in THFC were found in the HUF, only that  $1.0 \times 10^1 \text{ cfu/g}$  soil isolated in treatment B was also noticed in treatment C. From the results, HUB/THBC ratio of the treatments is in excess of 1% which is an indication of recent crude oil input into the environment. Also the significant growth differences observed in the treated soils could be attributed to favourable environmental condition consequence upon amendment of soil with plant residues. Soil fauna thrives in relative large number and subsequently brake down complex compounds to simple form that is released into the soil. The bacterial isolates of these soils belongs to the genera *Acinetobacter*, *Alcaligenes*,

*Arthrobacter*, *Bacillus*, and *Serratia*, while *Actinimycetes* were of the genera *Norcadia*. These isolates are typical from soils of the Niger Delta region[22].

### 3.4. Effects of Crude Oil Polluted Soil on Some Growth Parameters of Jute Mallow Plant

Inventory of best performed Jute mallow plant in each of the treatments for three experiments are presented in Table 3. Jute mallow plant vegetative growth parameters assessed at 4 WAP showed that averaged plant height and number of leaves per plant varied among experiments and increased significantly among the treatments. At  $0 \text{ t ha}^{-1}$  (treatment A), the plant heights were 6,4 and 6 cm (averaged, 5.33 cm), while in treatment B, plants height were 33, 35 and 35cm with mean height of 34.33cm. Also treatment C recorded 35, 36, and 36 cm with average height of 35.67 cm among the respective experiments/treatments. Similarly, number of leaves steadily increased in all the experiments but higher in the treated soils relative to control, indicating that phyto-remediation generally enhanced vigorous plant growth. However, there was no significant difference between treatments B and C.

Furthermore, the vegetative growth characteristics measured at 8 and 16 WAP among the three treatments were also significantly better in all pots receiving plant treatment than the control treatment where plant treatment was not applied. The tallest plant height of 152.67 and 173.67 cm were obtained in pots treated with sunflower and vetiver grass respectively. However, there was no significant change in the plant heights among the number of experiments conducted to validate the results in the respective treatments. Although the use of plant substantially promoted plant height, there was no significant increase in the number of leaves at 4 and 8 WAP among the treatments except treatment C at 8 weeks.

**Table 3.** Effects of crude oil polluted soil on some growth parameters of Jute mallow plant

Treatments	WAP	Plant height (cm)				No. of leaves per plant				Leaf dry weight per plant (g)			
		Experiments				Experiments				Experiments			
		1	2	3	Mean	1	2	3	Mean	1	2	3	Mean
A	4	6	4	6	5.33	2	2.5	3.1	2.53				
B	4	33	35	35	34.33	3	3.1	3	3.03				
C	4	35	36	36	35.67	3.1	3.1	3.2	3.13				
	LSD 0.05	16.197	17.039	17.039		0.608	0.635	0.643					
A	8	10	8	9	9.00	2	2	2	2.00	27.2	20.6	25.9	24.57
B	8	53	55	64	57.33	4.2	4.4	5.7	4.77	45.3	53.1	54.4	50.93
C	8	65	73	87	75.00	5.1	5.8	6.4	5.77	53.6	54.5	58.7	55.60
	LSD 0.05	28.919	32.45	39.526		1.595	1.922	2.364		13.5	15.373	17.081	
A	16	13	12	11	12.00	6	5	3	4.67	29.6	27.3	25	27.30
B	16	133	151	174	152.67	14.1	13.6	16.3	14.67	89.3	96.2	95.8	93.77
C	16	171	175	175	173.67	13.3	14	15.3	14.20	108.2	135.2	155.3	132.90
	LSD 0.05	82.47	87.43	93.243		4.464	4.508	5.68		41.027	53.398	63.389	

At 8 WAP, leaf dry weight significantly varied from 20.6 to 27.2 g at 0 t ha<sup>-1</sup> up to 53.1 to 54.4g; and 54.5 to 58.7 g for treatments B and C respectively, with mean values of 24.57g (treatment A), 50.93g (treatment B) and 55.60g (treatment C). At 16 WAP, the mean leaf dry weights were 27.30, 93.77, and 132.9g for the respective treatments A, B, and C, with a range of 25.0 to 29.6g for treatment A, 89.3 to 96.2g for treatment B, and 108.2 to 135.2g for treatment C.

The percentage increment in leaf dry weight from 8 to 16 WAP was significantly lower (11%) in pots without treatment than those with treatments; in soils treated with sunflower, the percent increment in leaf dry weight from 8 to 16 WAP was 84% and 139% for those treated with vetiver grass for the same period. However, the general higher percent increments of Jute mallow plant parameters grown on treated soils probably suggest that phyto remediation is effective and it enables plant to attain maximum growth.

#### 4. Findings and Recommendations

The degree of toxicity on crops was attributed to the direct or contact toxicity of the volatile and water soluble hydrocarbons in spilled soil that may have penetrated the tissues of the tender plant. There were alterations in the physico-chemical properties of the polluted soils treated with sunflower and vetiver grass compared with the control. Application of sunflower significantly ( $p < 0.05$ ) reduced the total hydrocarbon content of the soils by 25.59%, while vetiver grass had removal efficiency greater than 41%. From the above results, it could be concluded that crude oil polluted soils can be reclaimed with the application of phyto technology sourced from sunflower and vetiver grass. Albeit at 2.5 t ha<sup>-1</sup>, the growth parameters of the cultivated Jute mallow plant gained advantage on soil treated with vetiver grass. With the present poor fertility status of acid sand soil, coupled with scarcity of arable land resource due to oil spillage from exploration of petroleum in the Niger Delta Region of Nigeria, the yield potential of farmers in this study area can be successfully maximized with application of phyto technology of vetiver and sunflower. Therefore, no special management tool is required for this soil when it is contaminated with crude oil. It relatively low-cost, low techniques with high public acceptance and can often be carried out on site.

#### REFERENCES

- [1] Swanell R.P., Lee, K., McDonagh, M 1996. Field evaluation of marine oil spill bioremediation. *Microb. Rev.* (June): 342-365.
- [2] Mishra, S.J, Jyot, R.C., Kuhad, B., Lal, R. 2001. Evaluation of inoculum addition to stimulate *in situ* Bioremediation of oily-sludge-contaminated soil. *Appl. Environ. Microbiol.* 67(4): 1675-1681.
- [3] Schnoor, J.L. 1997. Phytoremediation. Ground water technology Analysis Center: Technology. Am. Chem. Soc. Agron. Madison, WI. pp. 23-26.
- [4] Balba, M.T., Al-Awadhi, N. and Al-Daher, R. 1998. Bioremediation of Oil-Contaminated Soil: Microbial Methods for Feasibility Assessment and Field Evaluation. *J. Microbiol. Meth.* 32:155-164.
- [5] Zappi, M.E., Rogers, B.A., Teeter, C.L., Gunnison, D. and Bajpai, R. 1996. Bioslurry Treatment of a Soil Contaminated with Low Concentrations of Total Petroleum Hydrocarbons. *J. Haz. Mat.* 46:1-12.
- [6] Cho, J.S., DiGiulio, D.C. and Wilson, J.T. 1997. *In-Situ* Air Injection, Soil Vacuum Extraction and Enhanced Biodegradation: A Case Study in a JP-4 Jet Fuel Contamination Site. *Env. Prog.* 16(1):35-42.
- [7] McLaughlin, B. 2001. Soil Remediation. *Enginr. Sci. Rev.* 2:69-77.
- [8] Harder, E. 2004. Bioremediation of engine oil. Little Flower Academy. Dallas, Texas.
- [9] Lee, K., A.D. Venosa, G. Wohlgeschaffen, S.E. Cobanli, and J. Gauthier. 2002. Monitoring habitat recovery and toxicity reduction in an oiled freshwater wetland to determine remediation success. *Biorem. J.* in preparation.
- [10] Edem, I.D., Usoroh, A.D. and Ijah, C. J. 2013. Soil Properties Dynamics Induced by Passage. of Fire During Agricultural Burning. *International Journal of Plant & Soil Science* 2(1): 111-126, 2013; Article no. IJPSS.2013.009.
- [11] Gee, G. W. and Or, D. 2002. Particle size analysis. In: J. H. Dane and G. C. Topp (eds.) *Methods of soil analysis. Part 4, Physical Methods*, SSSA, Incorporated, Madison, 255-294.
- [12] Thomas, G. W. 1982. Exchangeable cation. In: Page A.L., Miller, R. H. and Keeny, D. R. (Edition), *Method of Soil Analysis, II. Chemical and Microbiological Properties*, Agronomy Monogram 9, Second Edition. Soil Science Society of America, Madison, Wisconsin. Pp 296-301.
- [13] Cabardella, C.A., Galda, A.M., Doran, J.W., Wienhold, B.J. and Kettler, T. A. 2001. Estimation of particulate and total organic carbon by weight loss-on-ignition In; Lal R., Kimbe, J.M., Follet R. F., Stewart, B. A. (Editions), *Assessment method for Soil carbon*. Lewis publishers, Boca Raton, FL, pp 349-359.
- [14] Nelson, D.W. 1983. Determination of ammonium in KCl extracts of soils by the salicylate method. *Communication in Soil Science and Plant Analysis*, 14:1051-1062.
- [15] Special Programme for Food Security (NPSF). 2005. Critical values for nutrient elements.
- [16] Nunez-Delgado, B., Morrison L, Gulson, L. 2002. Threshold of concern: a technique for evaluating environmental impact and amenity values. *Journal of forestry* 84-86.
- [17] Udo-Inyang, U. C. and Edem I. D. 2012. Oil Producing and Agro-Ecological Area in the Niger Delta, Nigeria: A Case of Oil Spillage. *International Journal of Agriculture Innovations and Research*. 1: 6, ISSN (Online) 2319-1473.
- [18] Enwezor W.O., and Sabulo R.A. 1981. Fertility Status and Productivity of the Acid Sands. In E. J. And Udo and R. Sabulo A (Eds) *Acid Sands of Southern Nigeria*. SSS Special

Publication Monograph No.1 Pp 165-167.

- [19] Hendrickx, J. M. H., Wraith, J. M., Corwin, D. L., and Kachanoski, R. G. 2002. Solute content and concentration, In: Dane, J. H., and Topp, G. C., eds. iMethods of Soil Analysis, Part 4: Physical Methods. in Soil Science Society of America, Madison, WI.
- [20] Rolling W.F.M, Milner, M.G, Jones, D. M., Lee, K., Daniel, F. Swannell, R. J. P, and Head, I.M. 2002. Robust hydrocarbon degradation and dynamics of bacterial communities during nutrient-enhanced oil spill bioremediation. Appl. Environ. Microbiol. 68 (11): 5537-5548.
- [21] Akinrinde, E.A. 2006. Issues of optimum nutrient supply for sustainable crop production in tropical developing countries. Pakistan journal of nutrition 5 (4): 377-387.
- [22] Udotong, I. R., Akpanekong, O. J. 2007. Microbial and physico-chemical studies of wetland soils in Itu, Nigeria. Nigeria Journal of microbiology. 21:1623-1635.