

Study of Glutathione-S-Transferase, Cancer Embryonic Antigen and Carbohydrate Antigen 19-9 in Ulcerative Colitis and Colorectal Cancer

Fatma Saffeyeldin Mohamed^{1,*}, Mirhan M. Elkady², Eman Fekry Mohamed³,
Boshra E. Hussein⁴, Walid S. H. Elsaied⁵

¹Department of Tropical Medicine, Faculty of Medicine (for girls), Al-Azhar University, Cairo, Egypt

²Department of Clinical Pathology, Faculty of Medicine (for girls), Al-Azhar University, Cairo, Egypt

³Department of Internal Medicine, Faculty of Medicine (for girls), Al-Azhar University, Cairo, Egypt

⁴Department of Tropical Medicine, Faculty of Medicine, Tanta University, Tanta, Egypt

⁵Department of Pathology, Faculty of Medicine, Zagazig University, Zagazig, Egypt

Abstract Background: Ulcerative colitis (UC) is a chronic inflammatory bowel disease (IBD) with a high risk of colorectal cancer (CRC), which is a major cause of cancer morbidity and mortality worldwide. Glutathione-S-transferase (GST) has an essential role within the cells, including the conjugation and detoxification of toxic or carcinogenic compounds, such as reactive oxygen species (ROS). Cancer embryonic antigen (CEA) and carbohydrate cancer antigen (CA19-9) are tumor markers that are used for diagnosis of colorectal cancer. **Aim:** The aim of the study was to assess the total plasma GST activity, CEA and CA 19-9 in ulcerative colitis and colorectal cancer. **Subjects and methods:** The plasma GST activity, CEA and CA 19-9 were assessed in 40 patients with CRC (group I), 40 patients with UC (group II) and 20 apparently healthy individuals as controls (group III). **Results:** The mean GST activity, CEA and CA19-9 were significantly higher in CRC and UC patients than controls with no significant difference between CRC & UC patients in CEA or CA19-9. A significant difference was found between Gs I & II as regards GST activity. **Conclusion:** Measurement of the total plasma GST activity may be helpful as a tumor marker of colorectal cancer.

Keywords Glutathione-S-Transferase (GST), Colorectal cancer (CRC), Ulcerative colitis (UC), CEA & CA19-9

1. Introduction

Inflammatory bowel disease is a chronic inflammatory condition affecting the gastrointestinal tract. It has two main forms, ulcerative colitis (UC) and Crohn's disease [1]. Ulcerative colitis affects only the large bowel, and the inflammatory process is confined to the mucosa. Ulcerative colitis is a well-known risk factor for colorectal cancer [2]. The possibility of UC developing to intestinal cancer has a close and positive relationship with the extent and duration of the disease [1].

Colorectal cancer (CRC) is a major health concern and a leading cause of cancer death in the western world. The estimated life time risk of CRC is 5% to 6%. Worldwide, it is estimated that there was over one million new CRC cases and the annual age-adjusted incidence rate is 57 per 100000 [3]. Survival is directly related to the extent of disease at the

time of diagnosis. Those diagnosed at an advanced stage have an estimated 5-year survival rate of 7%, in contrast with a survival rate of 92% for individuals detected at an early stage, since advanced CRC is largely refractory to conventional therapy and is one of the least curable malignancies [4].

Despite continuing advances in diagnosis and therapy, long-term survival has not improved significantly over the last four decades, and almost 50% of CRC patients will eventually die of their disease [5]. This situation mandates improvement in the early detection of this surgically curable disease and preventative interventions to reduce the incidence of the disease and its morbidities and mortalities [6]. Indeed, given our understanding of the pathogenesis of CRC, current screening technologies, and effective preventative interventions, CRC should be highly preventable [7].

The colon and rectum are constantly challenged by potentially harmful compounds, including mutagens and carcinogens [7]. The large intestine possesses several defense mechanisms to counteract damage of the colorectal mucosa by such reactive compounds [8]. These include the

* Corresponding author:

dr.eltonsy.@gmail.com (Fatma Saffeyeldin Mohamed)

Published online at <http://journal.sapub.org/medicine>

Copyright © 2015 Scientific & Academic Publishing. All Rights Reserved

ability to up-regulate detoxification systems, essentially, the glutathione (GSH) / glutathione-S-transferase (GST) detoxification system [9].

Glutathione-S-transferases (GSTs) are enzymes with detoxifying properties that are ubiquitously expressed, with especially high levels in the liver, gonads and colon [1].

These enzymes are expressed in a wide variety of human tissues, including both normal and malignant colonic mucosa [10]. Most human gastrointestinal tumors contain increased GST enzyme activity which has been suggested as being beneficial to cancer prevention [11]. Previous studies have shown some evidence for a positive association between GST gene mutations and the susceptibility to multiple tumors [1].

The most important currently available markers in CRC that provide prognostic or predictive information are serum markers such as CEA and CA19-9, expressed by tumor tissue [12]. Carcinoma embryonic antigen (CEA) is an oncofetal tumor marker in 70% of cases. It is significant in the diagnosis of colorectal cancer. Increase of its concentration for a period of a few months after the surgery help in prediction and diagnosis of recurrence [13]. Its concentration is also correlated with the tumor size. Thus, tumors of smaller size have normal serum concentrations of CEA antigen. Only tumors greater than 3 cm are accompanied with high concentration of CEA antigen [14]. Carbohydrate antigen (CA19-9) is a cancer antigen whose elevated serum concentration is also detected in the case of colorectal cancer. It is a tumor marker that is observed to be in elevated serum concentration with metastatic colon cancer [12, 15].

2. Subjects and Methods

The study was conducted on 40 patients with CRC (group I), 40 with UC (group II) and 20 apparently healthy controls (group III). Mean ages of participants were 50.05 ± 11.43 , 49.25 ± 9.42 and 48.25 ± 15.68 in groups I, II & III, respectively.

Patients included in this study were selected from outpatient clinics and inpatients of Internal and Tropical Medicine Departments, Alzahraa University Hospital, Cairo, Egypt. Group (I) included 27 males and 13 females with CRC, group (II) included 18 males and 22 females with UC and group (III) included 10 males and 10 females as controls. Patients with liver disease or high bilirubin were excluded as both affect GST enzymatic activity, thus decreasing the clinical sensitivity of the test [16]. A verbal consent was taken from all patients and controls.

All participants were subjected to: thorough history taking, full clinical examination and the following routine laboratory investigations; Liver function tests (Alanine transaminase ALT, aspartate transaminase AST, alkaline phosphatase ALP, total and direct bilirubin, albumin and total proteins), kidney function tests (Urea and Creatinine).

Estimation the total plasma GST enzyme activity, CEA

and CA19-9 were assayed in serum samples using enzyme-linked immunosorbent assay (ELISA) in all patients and controls. Abdomino-pelvic ultrasound was also done. CT was done for patients with CRC. Colonoscopy, biopsy and histopathological examination were done for all patients.

2.1. Measuring the total GST Activity in Plasma

Plasma total GST activity was measured in all patients and controls using enzyme-linked immunosorbent assay (ELISA). The Cayman Chemical Glutathione S-transferase assay Kit (Catalog No. 703302) was supplied from Cayman Chemical company, USA. Plasma total GST activity kit measures the conjugation of 1-chloro-2, 4-dinitrobenzene (CDNB) with reduced glutathione. The conjugation is accompanied by an increase in absorbance at 340nm. The rate of increase is directly proportional to the GST activity in the sample. The Cayman GST Assay Kit can be used to measure GST activity in plasma.

2.2. Quantitative Measurement of CEA

The CEA enzyme immunoassay test kits (Catalogue No. EK-310-11) were supplied from Phoenix Pharmaceuticals, Inc., CA, USA. The CEA ELISA test is based on the principle of a solid phase enzyme linked immunosorbent assay. The assay system utilizes a monoclonal antibody directed against a distinct antigenic determinant on the intact CEA molecule is used for solid phase immobilization (on the microtiter wells). A goat anti-CEA antibody conjugated to horseradish peroxidase (HRP) is in the antibody-enzyme conjugate solution. The test sample is allowed to react simultaneously with the two antibodies, resulting in the CEA molecules being sandwiched between the solid phase and enzyme-linked antibodies. After a 1 hour incubation at room temperature, the wells are washed with water to remove unbound labeled antibodies. A solution of TMB Reagent is added and incubated for 20 minutes, resulting in the development of a blue color. The color development is stopped with the addition of stop solution changing the color to yellow. Absorbance is measured at 450 nm.

2.3. Quantitative Measurement of CA19-9

The CA19-9 enzyme immunoassay test kits (Catalogue No. TM E-4500) were supplied from LDN Labor Diagnostika Nord GmbH Co. KG, Germany. The TM-CA19.9 ELISA Kit is a solid phase enzyme-linked immunosorbent assay (ELISA) based on the sandwich principle. The microtiter wells are coated with a monoclonal antibody directed towards a unique antigenic site of the CA 19-9 molecule. An aliquot of patient sample containing endogenous CA 19-9 is incubated in the coated well with assay buffer. After a washing step a second incubation follows with enzyme conjugate, which is an anti-CA 19-9 antibody conjugated with horseradish peroxidase. After incubation the unbound conjugate is washed off. The amount of bound peroxidase is proportional to the concentration of CA 19-9 in the sample. Having added the substrate solution,

the intensity of color developed is proportional to the concentration of CA 19-9 in the patient sample.

2.4. Statistical Methods

The collected data were coded, fed to a computer, organized and statistically analyzed. Statistical analyses were used computer programs: Microsoft excel version 10 and Statistical Package for Social Science (SPSS) for windows version 20.0 [17].

A) Descriptive statistics:

1. Qualitative (categorical) data were presented by frequency and percentage.
2. Quantitative data were presented by mean \pm SD.

B) Analytical statistics: Comparing groups was done using

1. Student "t"- test: was used for comparison between mean of two groups as regard quantitative variables.
2. Analysis of variance [ANOVA (F)] test: was used for comparison of quantitative data of more than 2 groups.

C) Mann-whitney (Z test) and Kruskal Wallis Test were used for non-parametric data.

D) Pearson linear correlation coefficient (r) was estimated to show the relationship between quantitative parameters [17].

E) Receiver operating characteristic (ROC) curves to determine a cutoff value. The optimal value of the cut-off point is thus obtained when the sum of sensitivity and specificity is at its maximum [18].

3. Results

Patients with CRC (group I) included 40 patients [27 males (67.4%) and 13 females (32.6%)], group II (UC) included 40 patients [18 males (45%) and 22 females (55%)] and group III included 20 apparently healthy controls [10 males (50%) and 10 females (50%)]. Mean ages of participants were 50.05 ± 11.43 , 49.25 ± 9.42 and 48.25 ± 15.68 in groups I, II & III, respectively (Table 1 and Figures 1 & 2).

Table (1). Age and sex distribution among the studied groups

Characteristic	Group I CRC (n=40)		Group II UC (n=40)		Group III control (n=20)		Significant test	P-value	
	N	%	N	%	N	%			
Age (years)	50.05 ± 11.43 a		49.25 ± 9.42 a		48.25 ± 15.68 a		F = 2.662	0.098 NS	
Sex	Male	27	67.4	18	45	10	50	X ² = 4.635	0.099 NS
	Female	13	32.6	22	55	10	50		

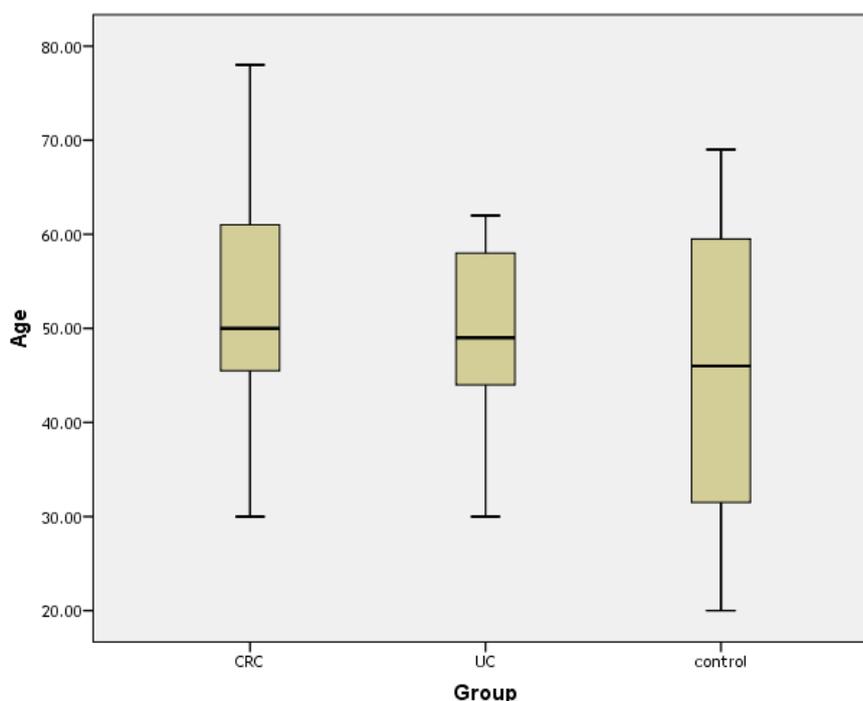


Figure (1). Box plot of groups as regards age and their statistical difference

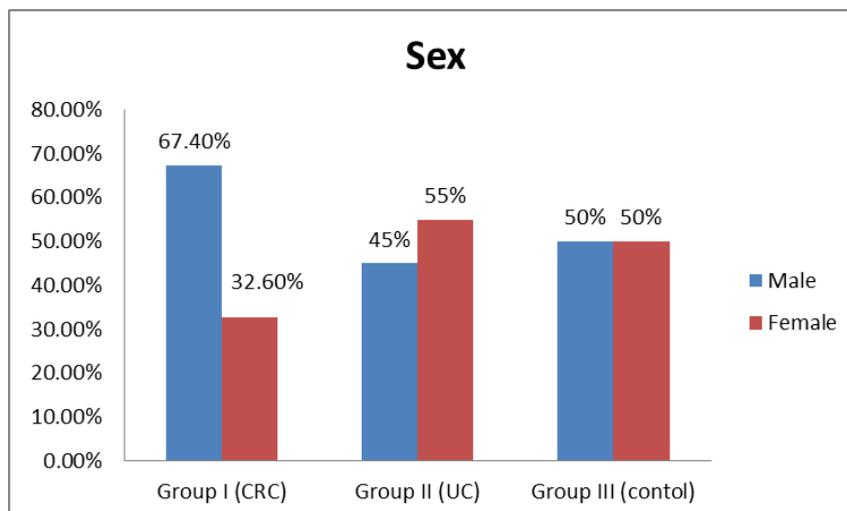


Figure (2). Distribution of sex among the studied groups

As regards site of CRC, it was 42.5%, 27.5%, 20% and 10% in the rectum (and/or sigmoid colon) followed by descending, ascending and transverse colon respectively (Table 2).

There was a statistically significant increase in GST activity, CEA and CA19-9 in CRC & UC patients in comparison to controls, with no significant difference between groups I & II in CEA or CA19-9. However, GST activity was significantly higher in CRC than UC patients (Table 3 and Figures 3, 4 & 5).

Correlation analysis showed that CA19-9 had a significant positive correlation with GST and CEA, while there was no significant correlation between GST and CEA (Table 4 and Figures 6 & 7).

ROC curves were drawn to look at maximum sensitivity and specificity for studied markers and also to see which one gave maximum area under curve (AUC), used for diagnosis of CRC. The maximum area under the curve for GST, CEA, CA19-9 was (0.84, 0.68 and 0.77, respectively). Cut-off was >44.85 nmol/min/ml, 7.55 ng/ml and 47.25 U/ml for GST, CEA and CA19-9, respectively, cut off can predict CRC. Sensitivity and specificity of GST were 65% and 95%, respectively, while CEA showed 68% sensitivity and 97% specificity. Sensitivity and specificity of CA19-9 were 55% and 90%, respectively (Figures 8).

Table (2). Site of cancer in patients with CRC (G I)

Site of Cancer	N	%
Ascending colon	8	20
Transverse colon	4	10
Descending colon	11	27.5
Rectum and/or sigmoid colon	17	42.5

Table (3). Mean level of plasma GST activity, CEA and CA19-9 among studied groups

TEST	Group I CRC	Group II UC	Group III Control	Significant test	P-value
GST (nmol/min/ml)	43.33±10.03 a	37.78±5.03 b	18.77±3.47 c	F= 65.463	0.000 HS
CEA(ng/ml)	5.75±3.59 a	4.88±1.67 a	2.02±1.36 b	Kruskal Wallis = 22.266	0.000 HS
CA19-9 (U/ml)	46.31±18.91 a	37.26±11.93 a	21.15±7.81 b	F = 17.872	0.000 HS

Groups sharing the same initials (letters a, b & c) indicate no statistically significant difference, while groups with different initials indicate a statistically significant difference.

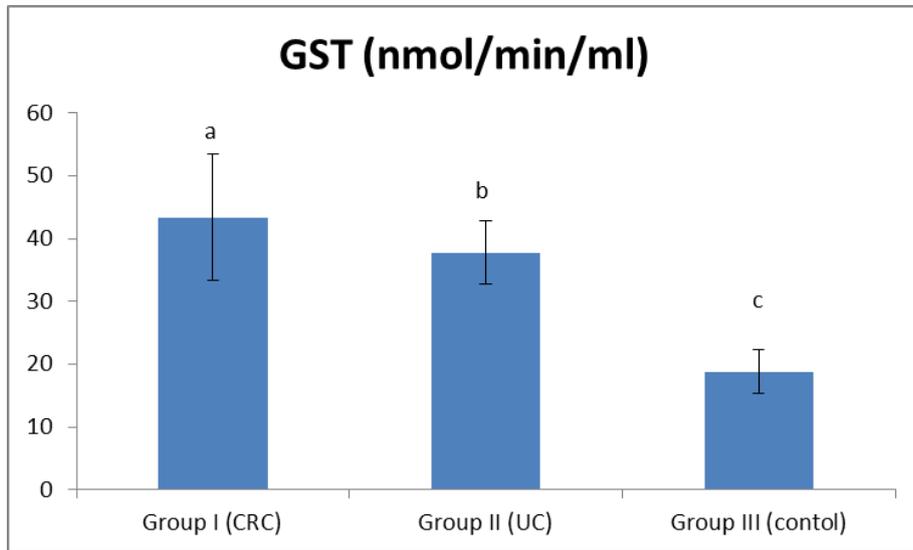


Figure (3). Comparison between both groups as GST and their statistical significance

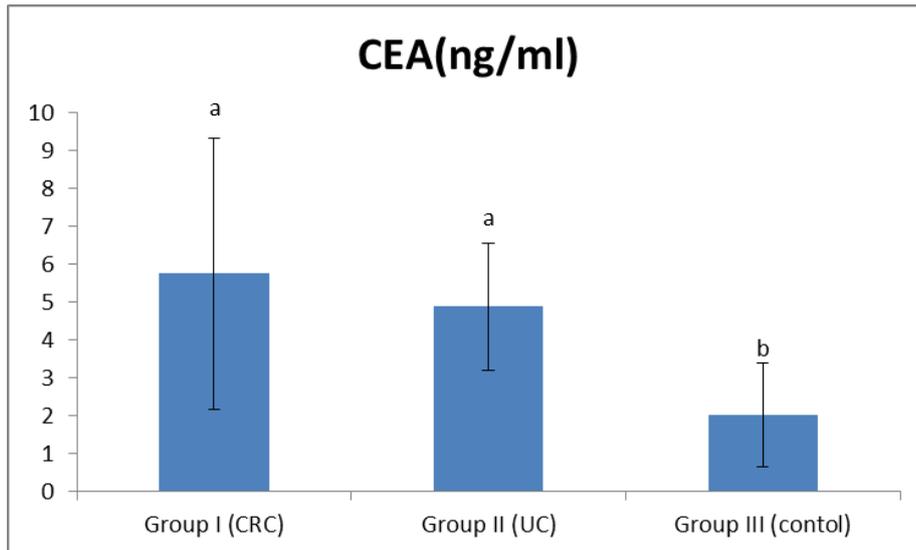


Figure (4). Comparison between both groups as CEA and their statistical significance

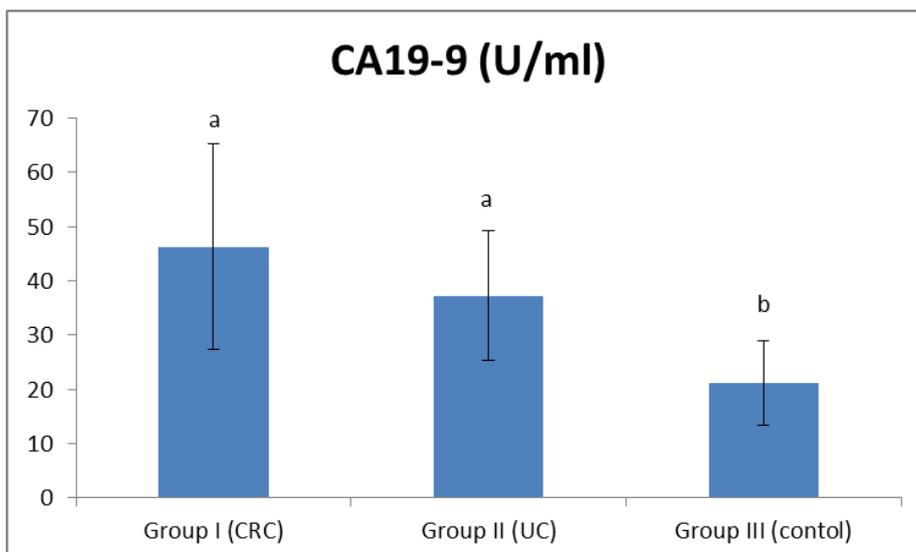


Figure (5). Comparison between both groups as CA19-9 and their statistical significance

Table (4). Correlation between CEA, GST and CA19-9 among the studied groups

Correlations			
		GST	CEA
CEA	r	0.079	
	p	0.55	
CA19-9	r	.257*	.299*
	p	0.048	0.02

*. Correlation is significant at the 0.05 level

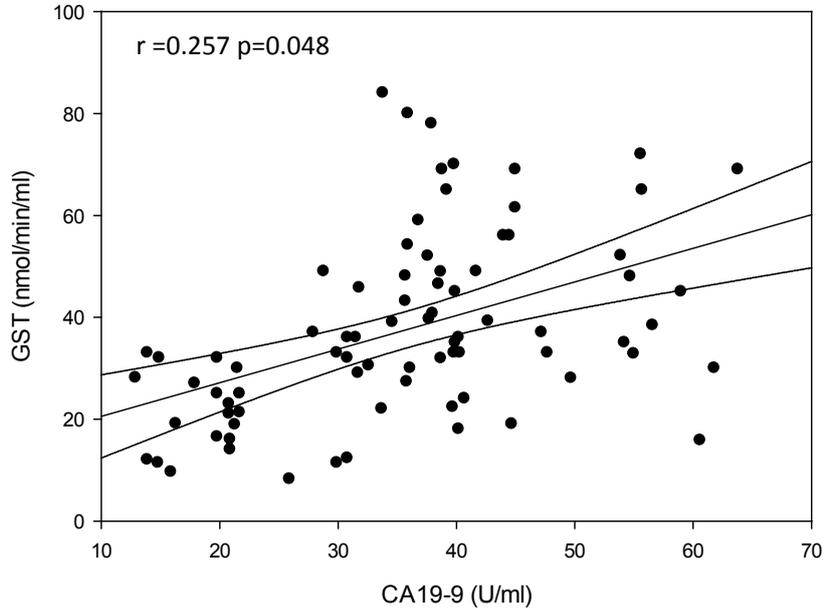


Figure (6). Linear correlation between GST and CA19-9 and their statistical significance

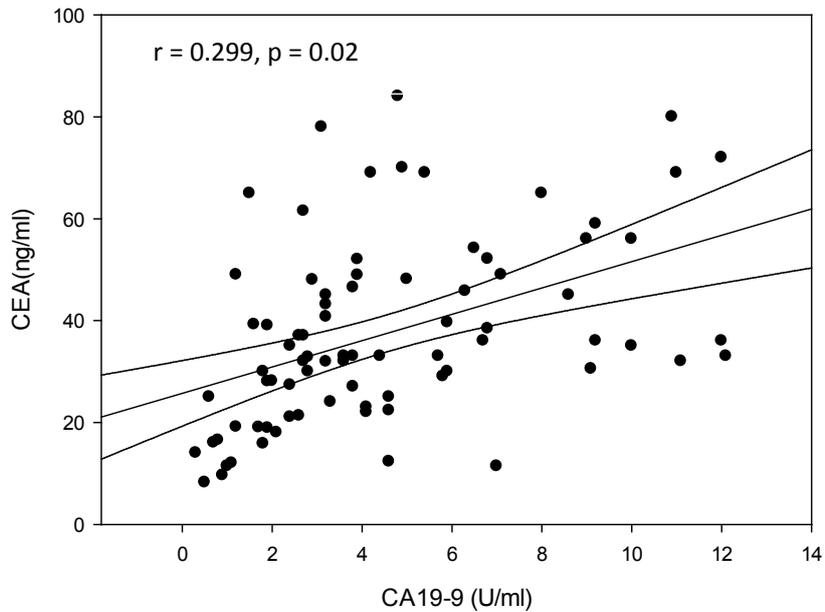


Figure (7). Linear correlation between CEA and CA19-9 and their statistical significance

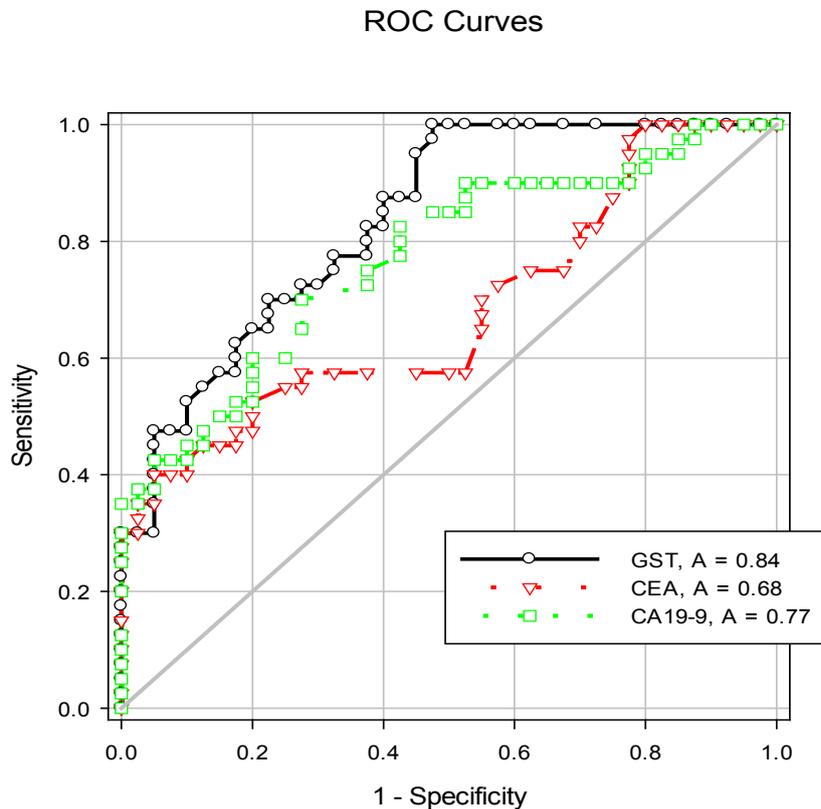


Figure (8). Sensitivity and specificity of CEA and plasma GST activity in the CRC patients shown in the ROC curve

4. Discussion

Colorectal cancer (CRC) is a major health concern and a leading cause of cancer death in the western world [3]. Ulcerative colitis is a well known risk factor for colorectal cancer [2].

The most important currently available markers in CRC that provide prognostic or predictive information are serum markers such as CEA and CA19-9, expressed by tumor tissue [12]. Previous studies have shown some evidence for a positive association between GST gene mutations and the susceptibility to multiple tumors [1].

The present study had demonstrated a significant elevation in the plasma GST activity in CRC patients compared to UC and the control group. This was in agreement with Naidu *et al.* [19] and Nomani *et al.* [20], they compared GST activity levels in paired samples of colorectal cancer, adenoma and adjacent normal mucosa.

The results of the present study were also in agreement with Severini [21], who found that the mean value of the GST activity was significantly elevated in patients with CRC as compared with the reference population. He suggested that GST measurement may be useful as a tumor marker in CRC. Moreover, the combined determination of GST and other markers increase the sensitivity for cancer detection.

Butler *et al.* [22] measured the glutathione (GSH) concentrations and GST activity in adenomatous polyps, cancer and normal mucosa. They found that concentrations

of GSH were significantly higher in adenomas and cancer than in uninvolved mucosa while GST activity was significantly higher in cancer and these results were in agreement with our results.

In contrary to the results of the present study, Upadhyya *et al.* [23] and Ozdemirler *et al.* [24] demonstrated that plasma GST did not show any significant change between CRC patients and controls.

A significant elevation was also found in plasma GST activity in cases with UC, compared to the control group, this was in agreement with Nieto *et al.* [25], they performed the study on experimental rats. However, our results were not in agreement with Bhaskar *et al.* [26], they found a significant decrease in the activity of GST activity in patients with UC compared to controls. They speculated that the reduced activity of GST in UC may be an additional factor in the pathogenesis of mucosal damage in this disease.

In the present study there was a significant increase in serum CEA in CRC and UC patients than controls, with no significant difference between CRC and UC patients. This was in agreement with Vukobrat-Bijedic *et al.* [27], they found a significantly elevated serum concentration of CEA in cases of colon cancer with already developed metastases.

The results of the present study, as regards CEA, were also in agreement with Carpelan-Holmstrom *et al.* [28], they evaluated and compared serum tumor markers, CEA & CA19-9, and their value in the diagnosis of malignant colorectal disease and concluded that only CEA provided a

significant diagnostic information. In their a study ROC curve was constructed, and the area under the curve (AUC) was determined for the probability of cancer. Of the individual markers, the highest AUC was observed for CEA (AUC = 0.746). In the present study, According to the ROC curve, the highest Area Under the Curve (AUC) was for GST (AUC = 0.846).

Also, Zhao *et al.* [29] concluded that CEA levels in CRC patients were significantly higher than those in controls and that was in agreement with our results. In the present study, a significant increase was found in CA19-9 in CRC and UC than controls, with no significant difference between the first two groups (Gs I & II). This was in agreement with Basbug *et al.* [30], they found that the mean values of CA19-9 were elevated in CRC than controls. They concluded that CA19-9 should be used for the study of this kind of malignancy. However, Carpelan-Holmstrom *et al.* [28], Cerda *et al.* [31] and Morita *et al.* [32] found no significant difference in CA 19-9 values between cancer and control groups and this was not in agreement with our results. Also Morita *et al.* [32] did not recommend routine use of CA19-9 in the staging and surveillance of colorectal cancer patients.

5. Conclusions

GST activity, CEA and CA19-9 were significantly elevated in CRC and UC with no significant difference between both groups in CEA or CA19-9. While, GST activity showed a statistically significant increase in CRC than UC patients.

6. Recommendations

Further studies for evaluation of GST in UC and its usefulness as a marker for CRC.

REFERENCES

- [1] Ye X, Jiang Y, Wang H, Chen L, Yuan S and Xia B (2011). Genetic polymorphisms of glutathione S-transferases are associated with ulcerative colitis in central China. *Cell Biochemistry and Biophysics*, 60(3): 323–328.
- [2] Riegler G, Bossa F, Caserta L, Pera A, Tonelli F, Sturniolo GC, Oliva L, Contessini Avesani E, Poggioli G, IG-IBD Group (2003). Colorectal cancer and high grade dysplasia complicating ulcerative colitis in Italy. A retrospective co-operative IG-IBD study. *Dig Liver Dis.*, 35(9): 628-634.
- [3] Siegel R, Ma J, Zou Z and Jemal A (2014). Cancer statistics, *CA Cancer. J Clin*, 64: 9–29.
- [4] Edna TH, Karisen V, Julamstre E and Lydersen S (2012). Prevalence of anemia at diagnosis of colorectal cancer: assessment of associated risk factors. *Hepatogastroenterology*, 59(115): 713-716.
- [5] Mitchell E (2013). Targeted therapy for metastatic colorectal cancer: role of aflibercept. *Clin Colorectal Cancer*, 12: 73–85.
- [6] Jemal A, Bray F, Center MM, Ferlay J and Ward E (2011). Global cancer statistics. *CA Cancer J Clin*, 61: 69-90.
- [7] Panczyk M (2014). Pharmacogenetics research on chemotherapy resistance in colorectal cancer over the last 20 years. *World J Gastroenterol*, 20: 9775–9827.
- [8] Kolahdoozan S, Sadjadi A and Radmard A (2010). Five Common Cancers in Iran. *Arch Iran Med*, 13: 143-146.
- [9] Monireh AN, Mojtaba P, Siamak MS, Farzaneh R and Ahmad M (2011). Glutathione S-transferase mu Gene Variants and Colorectal Cancer Development - Use of Sequence-specific Probes for an Iranian Population. *Asian Pacific Journal of Cancer Prevention*, 12: 1511-1515.
- [10] Josephy P (2012). Genetic variations in human glutathione. *Journal of Medical Investigation*, 59(3-4): 280-283.
- [11] Rui YY, Zhang D, Zhou ZG, Wang C and Yang L (2013). Can K-ras Gene Mutation Be Utilized as Prognostic Biomarker for Colorectal Cancer Patients Receiving Chemotherapy? A Meta-Analysis and Systematic Review. *PLoS ONE* 8(10): e77901.
- [12] Vukobrat-Bijedic Z, Husic-Selimovic A, Bijedic N, Mujkic A, Sofic A, Gogov B, Mehmedovic A, Bjelogric I, Glavas S and Djuran, A. (2014). Sensitivity of Symptomatology Versus Diagnostic Procedures and Concentration of CEA and CA19–9 in the Early Detection of Colorectal Cancer. *Acta Informatica Medica*, 22(2): 89–93.
- [13] Jurgensmeier J, Schmoll H, Robertson J, Brooks L, Taboada M and Morgan S (2013). Prognostic and predictive value of VEGF, SVEGFR-2 and CEA in mCRC studies comparing cediranib, bevacizumab and chemotherapy. *Br J Cancer*, 108: 1316–1323.
- [14] Selcukbiricik F, Bilici A, Tural D, Erdamar S, Soyuk O, Buyukunal E, Demirelli F and Serdengecti S (2013). Are high initial CEA and CA19-9 levels associated with the presence of K-ras mutation in patients with metastatic colorectal cancer? *Tumor Biol*, 34(4): 2233-2239.
- [15] Nahatoni H, Kumon T, Kumon M, Hamada S, Okanoue T, Kawamura A, nakatiani K, Hiroi M and Hanazaki K (2012). High serum levels of both carcinoembryonic antigen and carbohydrate antigen 19-9 in a patient with sigmoid colon cancer without metastasis. *J Med Invest*, 59(3-4): 280-283.
- [16] Hayes PC, Bouchler IA and Beckett GJ (1991). Glutathione-S-transferase in humans in health and disease. *Gut*, 32: 813-818.
- [17] Snedecor GM and Cochran WG (1982). *Statistical methods-7th edition*, Iowa state Univ., Press, Ames., Iowa, USA; 325-330.
- [18] Härdle W and Simar L (2007). *Applied Multivariate Statistical Analysis*. 2nd ed, Springer; 420pp.
- [19] Naidu KA, Nasir A, Pinkas H, Kaiser HE, Brady P and Coppola D (2003). Glutathione-S-transferase pi expression and activity is increased in colonic neoplasia. *In Vivo*, 17(5): 479-482.
- [20] Nomani H, Ghobadloo M, Yaghmaei B, Rezvanie NA and Yaghmaei K (2005). Glutathione S-transferases activity in

- patients with colorectal cancer. *Clinical Biochemistry*, 38(7): 621-624.
- [21] Severini G (1993). Glutathione S-transferase activity in patients with cancer of the digestive tract. *J Cancer Res Clin Oncol*, 120(1-2): 112-114.
- [22] Butler RN, Butler WJ, Moraby Z, Fettman MJ, Khoo KK and Roberts-Thomson IC (1994). Glutathione concentrations and glutathione S-transferase activity in human colonic neoplasms. *J Gastroenterol Hepatol*, 9(1): 60-63.
- [23] Upadhyaya S, Upadhyaya S, Mohan SK, Vanajakshamma K, Kunder M and Mathias S (2004). Oxidant-antioxidant status in colorectal cancer patients before and after treatment. *Indian J Clin Biochem*, 19(2): 80-83.
- [24] Ozdemirler G, Pabuccuoglu H, Bulut T, Bugra D, Uysal M and Toker G (1998). Increased lipid peroxide levels and antioxidant system in colorectal cancer. *J Cancer Res Clin Oncol*, 124: 555-559.
- [25] Nieto N, Torres MI, Fernandez MI, Giron MD, Rios A, Suarez MD and Gil A (2000). Experimental ulcerative colitis impairs antioxidant defense system in rat intestine. *Dig Dis Sci*, 45(9): 1820-1827.
- [26] Bhaskar L, Ramakrishna BS and Balasubramanian KA (1995). Colonic mucosal antioxidant enzymes and lipid peroxide levels in normal subjects and patients with ulcerative colitis. *J Gastroenterol Hepatol*, 10(2): 140-143.
- [27] Vukobrat-Bijedic Z, Husic-Selimovic A, Sofic A, Bijedic N, Bjelogrljic I, Gogov B and Mehmedovic A (2013). Cancer Antigens (CEA and CA 19-9) as Markers of Advanced Stage of Colorectal Carcinoma. *Med Arch*, 67(6): 397-401.
- [28] Carpelan-Holmstrom M, Louhimo J, Stenman UH, Alftan H, Jarvinen H and Haglund C (2004). Estimating the probability of cancer with several tumor markers patients with colorectal disease. *Oncology*, 66(4): 296-302.
- [29] Zhao XW, Jiang B, Han CZ and Jing JX (2005). Detection and clinical study of serum tumor markers in patients with colorectal cancer. *Zhonghua Zhong Liu Za Zhi*, 27(5): 286-288.
- [30] Basbug M, Arikanoglu Z, Bulbuller N, Cetinkaya Z, Aygen E, Akbulut S and Satici O (2011). Prognostic value of preoperative CEA and CA 19-9 levels in patients with colorectal cancer. *Hepatogastroenterology*, 58(106): 400-405.
- [31] Cerda SR, Bissonnette M, Scaglione-Sewell B, Lyons MR, Khare S, Mustafi R and Brasitus TA (2001). PKC-delta inhibits anchorage-dependent and -independent growth, enhances differentiation, and increases apoptosis in CaCo-2 cells. *Gastroenterology*, 120: 1700-1712.
- [32] Morita S, Nomura T, Fukushima Y, Morimoto T, Hiraoka N and Shibata N (2004). Does serum CA19-9 play a practical role in the management of patients with colorectal cancer?. *Dis Colon Rectum*, 47(2): 227-232.