

# Development of a New Sorption- Spectrophotometric Method for Determining the Ion Fe(III) Using Immobilized Methylthymol Blue

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**Abstract** An accelerated method for sorption-spectroscopic determination of iron ions in natural wastewater using immobilized methylthymol blue with improved metrological indices has been developed. Conditions for immobilization, complexation and detection of iron ions have been optimized. It has been established that the optimal buffer at pH 2.5 is a universal buffer solution. The developed method has been used in the analysis of drinking water with a detection limit of 0.1 µg/l.

**Keywords** Analytical reagent, Iron ion, Methylthymol blue, Immobilization, Asorption-spectrophotometric

## 1. Introduction

Iron is one of the microelements important for the human body. Deficiency of iron in the human body causes various vascular diseases [1]. However, it is worth noting that excess iron in the human body causes damage to the liver, pancreas and myocardium - the development of hemochromatosis [2]. Iron plays an important role in the development of the ecosystem, agriculture and industry, but at the same time it is a pollutant of water bodies [3]. Heavy metals in drinking water pose a threat to human health. Therefore, it is necessary to determine and control the amount of iron ions in environmental objects. A simple and rapid method of solid-phase spectrophotometry for determining free iron (III) in environmental and biological samples is proposed [4]. Disodium salt of 1-nitroso-2-naphthol-3,6-disulfonic acid (Nitroso R salt) and hexadecyltrimethylammonium bromide were used as ligands [5]. A flow-injection spectrophotometric method for determining the amount of dissolved and total iron in tap and natural waters has been developed. The catalytic effect on the oxidation of N,N-dimethyl-p- phenylenediamine was determined in samples of natural waters [6]. The spectrophotometric method for determining iron ions using various ligands does not allow for wide application due to its selectivity and low sensitivity. Iron (III) ion can be determined by atomic absorption spectrometry using immobilized sulfosalicylic acid [7], immobilization of sulfarsazene ligand on polyacrylonitrile-modified polyethylenepolyamine fiber [8]. Inductively coupled plasma mass spectrometry (ICP MS) has been used to determine iron and iodine elements in

watermelon seeds [9], iron and many microelements in the plant, leaves and seeds of the akkhangal plant [10], and 44 macro- and microelements in the flowers of *Tanacetum vulgare* [11]. However, these methods require expensive equipment. Sorption-spectrophotometric methods for determining heavy and toxic metals using immobilized organic reagents have also been developed. For example, immobilized bismuth ion xylene orange [13], mercury(II) ion immobilized on PPA1 fiber [14], immobilized disodium reagent 1,8-dioxonaphthalene -3,6-disulfonic acid for detection of chromium ions [15], manganese ions immobilized with alizarin. Methods of sorption -spectroscopic detection using -3-methylamino-N,N-diacetic acid have been developed [16].

At present, one of the widely developed methods is absorption spectrophotometry. This is an improved version of the spectrophotometric method, since the sensitivity of the spectrophotometric method is  $10^{-6}$ - $10^{-8}$ . If the sample amount is small, it should be evaporated without preliminary concentration. The sorption-spectroscopic method can be used to determine trace amounts of metal directly using an organic reagent immobilized on a fiber. This modified method includes several processes: concentration, determination directly in the solid phase, without separating the metal ion [17].

## 2. Methods and Materials

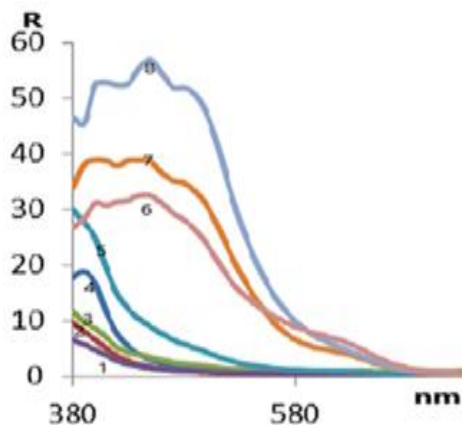
0.5585 g of high-purity metallic iron is dissolved in 10 ml of a mixture of hydrochloric and nitric acids (3:1), diluted to 50 ml with water and boiled until nitrogen oxides and 5585 mg/ml of  $\text{Fe}^{3+}$  chlorides are removed. A standard nitrate solution is formed [18]. Buffer solutions [19] were prepared based on literature data. To prepare a 0.01 M methylthymol

blue solution, 0.76085 g of the reagent powder is measured out on an analytical balance and distilled water is added to a 100 ml flask. To immobilize the selected fiber, 0.2 g is weighed on an analytical balance and activated with a 0.1 N HCl solution (24 hours). After activation, it is washed with distilled water to pH = 7. The fiber is kept wet in a Petri dish and stirred with a stirrer by adding 10 ml of 0.01 M solution of our reagent [17]. ABS 120-4N analytical scales (China) were used to measure the samples. The reagent environment was measured using a pH meter "PHS-3E" (China). The reflectance of immobilized fibers was measured using an X-Rite Eye one pro spectrophotometer (380-730 nm), and the light absorption of reactive solutions was measured using an EMC-30PC-UV Spectrophotometer and "UV-5100 UV VIS".

### 3. Results and Discussion

A sorbent for immobilization of an organic reagent was selected and the order of adding components to the immobilization process was determined. Optimal conditions for the formation of a complex of the immobilized organic reagent methylthymol blue (MTB) with  $\text{Fe}^{3+}$  were established. The research work studied the time dependences of complex formation, optimal pH values and optimal buffer solutions, the order of adding buffer solutions during the formation of complex compounds and other features.

When choosing the optimal conditions for immobilization of organic reagents on fibrous sorbents, 0.2000 g were taken from several types of sorbents on an analytical scale and activated with a standard solution of 0.1 M hydrochloric acid for 24 hours, then washed with distilled water and stored in a special Petri dish (in a wet state). For the analysis, CMA1, PPD, PPF and PPA1 fibers were selected, 0.2000 g of the selected fibrous sorbents were placed in a beaker, 10 ml of 0.1% solution of our methylthymol blue reagent was added and stirred for 5-10 minutes. The reflection spectrum of the immobilized fibers was measured on an X-rite one spectrophotometer (Switzerland).



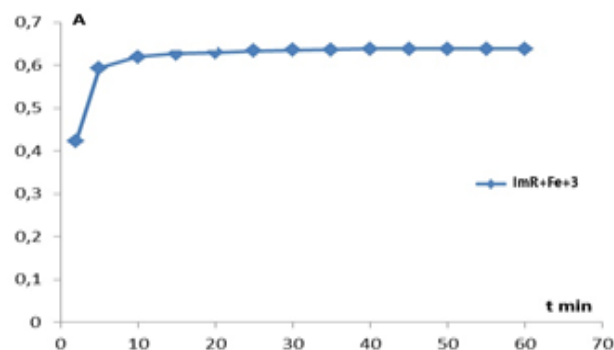
**Figure 1.** Reflectance spectrum of fibers immobilized with methylthymol blue reagent. (1-CMA; 2-PPD; 3-PPF; 4-PPA; 5-Imm.CMA; 6-Imm.PPD; 7-Imm.PPF; 8-Imm.PPA)

From the above figures, it can be concluded that the highest analytical signal was observed for the immobilized PPA1 fiber, which was chosen as the sorbent in the research work.

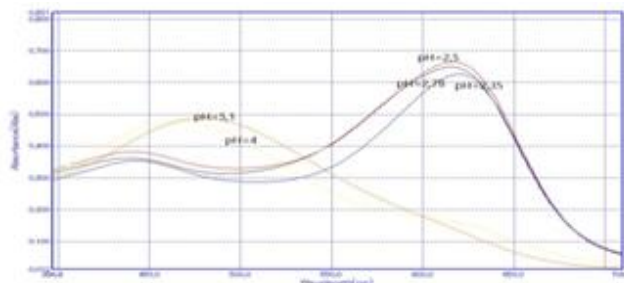
**Table 1.** Immobilization procedure

№	The order of the immobilization process	Optical density difference
1	PPA1+ $\text{Fe}^{3+}$ +MTB	1,958
2	PPA1+ MTB+ $\text{Fe}^{3+}$	2,523
3	Complex + $\text{Fe}^{3+}$	1,756

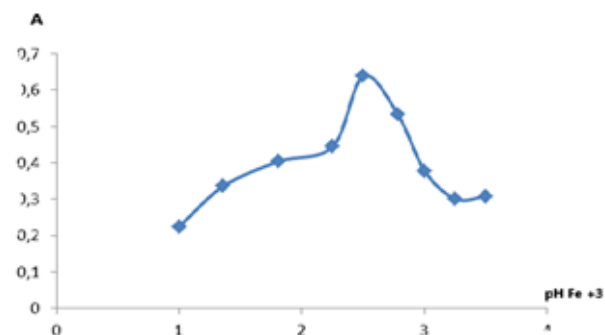
Based on the analysis results, the second method of the immobilization process was chosen.



**Figure 2.** Dependence of complex formation of immobilized MTB with iron (III) ion on time



**Figure 3.** Dependence of the formation of a complex of immobilized MTB with the  $\text{Fe}^{3+}$  ion on the pH of the environment



**Figure 4.** Optical density of iron complex formation at different pH values

The obtained results show that the complex compound is formed in 1 minute, has a maximum optical density in 7 minutes and forms a complete complex. Since the complex

compound is quite stable, it can be easily used in analytical processes. The formation of a complex of immobilized MTB with the  $\text{Fe}^{3+}$  ion depends on the acidity of the medium. As follows from Figure 2, it was found that the optimal medium for complex formation is  $\text{pH} = 2.5$ , and at  $\text{pH} = 4$  and above, the complex compound is not formed and only the reagent itself (MTB) gave an analytical signal.

**Table-2.** Selecting the optimal buffer for complexation

R+Me <sup>3+</sup>	pH	1	2	2,5	2,8	3	4
$\Delta A$							
MTB+Fe <sup>3+</sup>	glycol	0,470	0,476	0,490	0,500	0,493	-
	universal	0,460	0,518	0,520	0,535	0,534	-

According to Table 2, a universal buffer solution was chosen as the optimal buffer solution for complex formation.

## 4. Conclusions

In this research work, the optimal sorbent PPA1-fiber for immobilization of the methylthymol blue reagent was selected. A sorption-spectroscopic method for determining the iron (III) ion with immobilized MTS was developed. The optimal conditions for the formation of the complex compound were studied: dependence on time, pH of the medium, buffer solution and the order of adding the components.

## REFERENCES

- [1] Iminova I.M., Abdulboriyeva D.Yo., Abdulazizov A.A., Isakov A.A. Medicines used in iron deficiency anemia "Economics and society" №3(118)-1 2024 179-181.
- [2] Indu Khurana, Arushi Khurana: // Textbook of medical physiology // 2nd edition, 2009.
- [3] Arthur Guyton, John Hall: // Textbook of medical physiology // 14th edition, 2020.
- [4] Alberti, G., Emma, G., Colleoni, R., Nurchi, V. M., Pesavento, M., & Biesuz, R. (2019b). // Simple solid-phase spectrophotometric method for free iron (III) determination // Arabian Journal of Chemistry, 12(4), 573-579. <https://doi.org/10.1016/j.arabjc.2014.08.017>.
- [5] Cheng, F., Zhang, T., Sun, T., Wang, Y., Zhou, C., Zhu, H., & Li, Y. (2021). //A simple, sensitive and selective spectrophotometric method for determining iron in water samples // MicrochemicalJournal, 165, 106154. <https://doi.org/10.1016/j.microc.2021.106154>.
- [6] Lunvongsa, S., Oshima, M., & Motomizu, S. (2006). // Determination of total and dissolved amount of iron in water samples using catalytic spectrophotometric flow injection analysis. // Talanta, 68(3), 969-973. <https://doi.org/10.1016/j.talanta.2005.06.067>.
- [7] Adsorption-Atomic-Absorption Determination of Cu(II) Ions in Technogenic Waters. Ruzmetov, U.U., Jumayeva, E.S., Smanova, Z.A. Journal of Analytical Chemistry, 2024, 79(5), 578-584.
- [8] Ruzmetov, U.U., Jumaeva, E.Sh., Orziqulov, B.T., Smanova, // Determination of iron in water by flame atomic absorption spectrometry with sorption preconcentration // Industrial Laboratory. Materials Diagnostics, 2023, 89(12), 22-30.
- [9] Atakulova, N. B. Q., Askarov, I. R., & Anvarova, M. M. Q. (2022). // Determination of macro and microelements of "Shirin" watermelon seeds by inductively coupled plasma mass spectrometry method (icp-ms) // Journal of Chemistry of Goods and Traditional Medicine. 1, 1(5), 114-122. <https://doi.org/10.55475/jcgtm/vol1.iss5.2022.108>.
- [10] Ibrokhimov, A., Ibrokhimova, N., & Jalolov, I. (2024). // Study of the composition of macro and microelements of the leaves and seeds of the okchangel (nitraria sp) plant by icp-ms method // Fergana State University, 30(3), 142. Retrieved from <https://journal.fdu.uz/index.php/sjfsu/article/view/3963>.
- [11] Ibragimov, A., & Odilova, N. (2023). // Study of quantitative elemental composition of tanacetum vulgare L. Plant. // Scientific Journal of the Fergana State University, 29(1). [https://doi.org/10.56292/sjfsu/vol29\\_iss1/a113](https://doi.org/10.56292/sjfsu/vol29_iss1/a113).
- [12] Smanova, Z., Normatov, B., Ahmadjonov, U., Baigenzhenov, O., Hosseini-Bandegharai, A. (2024) // Sorption-spectrophotometric determination of bismuth ion (III) using immobilized xylenol orange on modified polyacrylonitrile. // International Journal of Environmental Analytical Chemistry. 195-203.
- [13] Ashirov, M.A., Yusupova, M.R., Akhmadzhanov, U.G., Baigenzhenov, O., Berdimurodov, E.T // Sulfarsazen-immobilized PPA Matrix as a New Efficient Analytical Reagent for Hg(II) Determination // Analytical and Bioanalytical Chemistry Research, 2023 , 10(2), 135-148.
- [14] Madatov, U., Rakhimov, S., Normatov, B., Khalilova, L., Smanova, Z. // Immobilized Reagent Disodium Salt of 1, 8-Dioxynaphthalene-3,6-Disulfonic Acids for the Determination of Chromium Ions // AIP Conference Proceedings, 2022, 2432, 050022.
- [15] Madatov, U., Rakhimov, S., Shahidova, D., Basant, L., Berdimurodov, E. // A new, green, highly effective procedure for manganese determination using alizarin-3-methylamino -N,N- diacetic acid immobilized on a polymer matrix // international Journal of Environmental Analytical Chemistry, 2022.
- [16] Zaporozhets O.A., Haver O.M., Sukhan V.V. // Immobilization of organic reagents on the surface of carriers. Advances in Chemistry. – 1997. – Vol. 66, №7. 701-712.
- [17] Amelin V.G., Tretyakov A.V. // Fabrics from artificial and natural fibers with immobilized reagents in chemical test methods of analysis. Journal of Chemistry Analyst.. – 2006. -V. 61, №4. – P. 430-435.
- [18] Ya. Lazarev, I.P. Kharlamov, P.D. Yakovlev, E.F. Yakovleva Handbook of a chemist-analyst Moscow. METALLURGY // 1976 p. 122.
- [19] Lurye Yu.Yu. // Handbook of analytical chemistry. Moscow: Chemistry. 1989. P. 446-445.