

Refractoriness of Porcine Corpora Lutea to Cloprostenol Sodium and Dinoprost Tromethamine Treatments at Day 7 of Oestrous Cycle

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Abstract This study was carried out to evaluate the refractoriness of porcine *corpora lutea* to exogenous prostaglandins (Cloprostenol sodium and Dinoprost tromethamine). Twenty (n = 20) apparently healthy sows were randomly assigned to 4 treatment groups. Group 1 (n=5) received two injections of Cloprostenol sodium (Synchromate[®]) (500µg) (days 0 and 13). Group 2 (n=5) received three injections of Synchromate[®] (Day 0, 7, 13). Group 3 (n=5) received two injections of Dinoprost tromethamine (Lutalyse[®]) (12.5mg) (days 0 and 13). Group 4 (n=5) received three injections of Lutalyse[®] (Day 0, 7, 13). Five (5) milliliters of blood was collected via the posterior vena cava before PGF_{2α} injections on day 0, 7, and 13 and once weekly afterwards till pregnancy was established. Data on progesterone profile were expressed as mean ± SEM. One-way ANOVA and Tukey's post hoc test were used to compare the mean values between the groups. Graphpad Prism[®] data package for windows (2009) was employed for all statistical analyses. A value of P < 0.05 was considered significant. There were no luteolysis seen following first and second injections of PGF_{2α} in groups 2 and 4 and following first injections in groups 1 and 3. Complete luteolysis was seen with behavioural oestrus after the second injection of PGF_{2α} at day 13 in groups 1 and 3 and after third injection of PGF_{2α} in groups 2 and 4. It was therefore concluded that the corpora lutea was refractory to both Cloprostenol sodium and Dinoprost tromethamine at Day 7. Further work should be done to find out the reason for lack of response of porcine CL to PGF_{2α} between days 7 and 12.

Keywords Refractoriness, Cloprostenol Sodium, Dinoprost tromethamine, Porcine CL, Progesterone

1. Introduction

Pig Production has been advocated as a short term measure towards alleviating the animal protein and calorie deficit, especially where there are no religious edicts preventing their production and consumption, (Ajala *et al.*, 2007). Oestrus synchronization is a valuable management tool for increasing the pregnancy rate in pigs (Brüssow and Wahner, 2011). Several techniques have been developed to induce oestrus. Oestrus synchronization methods in the sow vary and are all based either on controlling events leading to follicular maturation and ovulation or altering luteal lifespan (Estill, 2000). Prostaglandin F_{2α} (PGF_{2α}) is not luteolytic in the sows until about day 12 of the oestrous cycle (De Rensis

et al., 2012; Kouamo and Kanga-Waladjo, 2013; Tur, 2013). Synthetic progestins have been developed and used in sows for oestrus synchronization (Estienne *et al.*, 2001; Van Leeuwen *et al.*, 2011). The explanation for the relative insensitivity of porcine *corpora lutea* (CL) to the luteolytic effect of PGF_{2α} prior to day 12 of the oestrous cycle is unknown (Estill *et al.*, 1993). However, some studies demonstrated a relatively low numbers of specific high-affinity PGF_{2α}-binding sites (receptors) on large luteal cells prior to day 12 (Gadsby *et al.*, 1990; Diaz and Wiltbank, 2004). Others suggested that this lack of luteolytic sensitivity in porcine CL before Day 12 of the oestrous cycle may result from a deficiency in post-PGF_{2α} receptor signaling that is activated within CL (Diaz *et al.*, 2000). In addition, it was reported that the number of high affinity binding sites increased dramatically from day 13 coinciding with the apparent onset of enhanced sensitivity to the luteolytic effects of PGF_{2α} between days 12 and 13 (Gadsby *et al.*, 1990, Estill *et al.*, 2000). The insensitivity of sow's CL to the luteolytic effect of PGF_{2α} before day 12 is considered to preclude the use of prostaglandin in oestrus synchronization

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programmes for swine (Przygodzka *et al.*, 2015). This study was therefore designed to evaluate the refractoriness of porcine corpora lutea to Cloprostenol sodium and Dinoprost tromethamine at day 7 of the sow's oestrous cycle.

2. Materials and Methods

Study Location

The study was carried out at the Swine and Rabbit Research Programme of the National Animal Production Research Institute (NAPRI), Shika, Ahmadu Bello University, Zaria.

Experimental Animals and Herd Management

Twenty ($n = 20$) apparently healthy cross bred sows belonging to the Swine and Rabbit Research Programme of the NAPRI Shika, Zaria were used for this study. The sows were between 2 - 3 years of age weighing between 120 and 150 kg and were identified with ear tag numbers. The cross bred sows were fed with diet containing 16% crude protein. The ration was formulated to meet the minimum nutrient requirements for breeding sows and boars as recommended by National Research Council (NRC) (1998). The ingredients for the diet were sourced in NAPRI feed store and the ration was mixed in the feed mill in NAPRI, Shika, Zaria. Water was given *ad libitum*.

Experimental Design

A total of twenty ($n = 20$) cross bred sows were randomly divided into four groups and each of the group consists of 5 sows with different treatment protocol.

Group 1 (n=5) – Double intramuscular injection of Cloprostenol sodium (Synchromate®).

Each of the sows received a dose of 500 µg (2 ml) Cloprostenol sodium injection on days 0 (day of first injection) and 13 (day of second injection). The sows were then monitored for signs of oestrus. Those found in oestrus were bred using natural breeding.

Group 2 (n=5) – Triple intramuscular injection of Cloprostenol sodium (Synchromate®).

Each of the sows received a dose of 500 µg (2 ml) of Cloprostenol sodium on days 0 (day of first injection), 7 (day of second injection) and 13 (day of third injection). The sows were also monitored for signs of oestrus. Those found exhibiting signs of oestrus were bred using natural breeding.

Group 3 (n=5) – Double injection of Dinoprost tromethamine (Lutalyse®).

Each of the sows received a dose of 12.5mg (2.5 ml) dinoprost tromethamine on days 0 (day of first injection) and 13 (day of second injection). The sows were then monitored for signs of oestrus. Those found in oestrus were bred using natural breeding.

Group 4 (n=5) – Triple intramuscular injection of Dinoprost tromethamine (Lutalyse®).

Each of the sows received a dose of 12.5mg (2.5 ml) of

dinoprost tromethamine on days 0 (day of first injection), 7 (day of second injection) and 13 (day of third injection). The sows were also monitored for signs of oestrus. Those found exhibiting signs of oestrus were bred using natural breeding.

Oestrus Detection and Mating

The cross bred sows were observed visually for behavioural oestrus manifestation twice (0700-1000 and 1500-1800 h) daily from commencement of the study for 21 days. Sows were considered to be in oestrus when they stood to be mounted by females (homosexual mount) or male (heterosexual mount).

Blood Sampling

Five (5) milliliters of blood was collected via the posterior vena cava, using a 10-ml hypodermic syringe, fitted with 18 gauge needle, from the sows on days 0, 7, and 13 (just before prostaglandin injection), and once weekly afterwards until confirmation of pregnancy based on non-return rate to oestrus. Blood samples collected in vacutainers without anticoagulant were quickly transported to the laboratory. Serum samples were separated by centrifugation of the blood at 2500G for 15 minutes. The serum samples in vials were appropriately labeled and stored at -20°C until hormone analysis.

Progesterone assay

Serum P₄ was determined by using Competitive ELISA kits (AccuBind® ELISA, Monobind Inc. 100 North Pointe Drive Lake Forest, CA 92630, USA), intended for a quantitative determination of P₄ concentration in serum or plasma using ELISA microplate reader (ELx800). The sensitivity of the assay was 0.105 ng/ml. Within assay precision, coefficients of variation for low, normal and high pooled controlled serum samples were 9.9%, 3.1% and 2.9% respectively.

Data Analyses

Data on progesterone profile during the oestrous cycle were expressed as mean ± SEM. One-way ANOVA and Tukey's post hoc test were used to compare the mean values between the groups. Graphpad Prism® data package for windows (2009) was employed for all statistical analyses. A value of $P < 0.05$ was considered significant.

3. Results

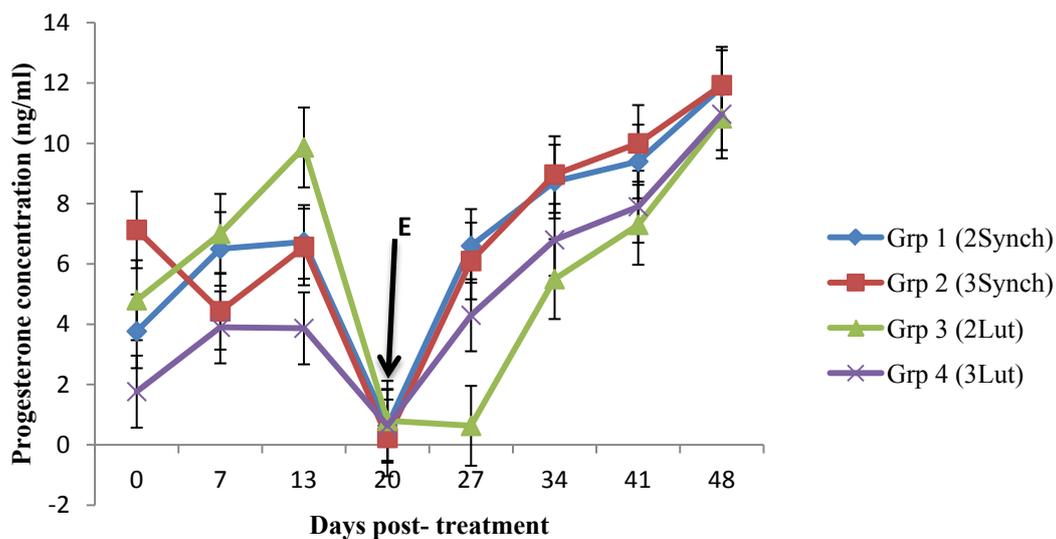
The serum P₄ concentrations of the samples collected from the first day of PGF_{2α} to 48 days or 26th day post-mating ranged from 0.00 to 11.87 ng/ml. For sows in group 1 (Figure 1), mean P₄ concentration increased from 3.77 ± 1.53 ng/ml at first PGF_{2α} (Synchromate®) to 6.73 ± 0.29 ng/ml on day 13th of the second dose of PGF_{2α} (Synchromate®) injection the concentration. There was a decrease following the injection of the second dose of the Synchromate® to 0.63 ± 0.09 ng/ml at day 20 and an increase was seen at day 27 (6.60 ± 1.41 ng/ml) which continues to increase till 11.87 ±

0.20 ng/ml at day 48 of the treatment and 23rd day post mating. For sows in group 2 (n=5) (Figure 1), mean P₄ concentration decreased from 7.13 ± 0.64 ng/ml at first PGF_{2α} (Synchromate[®]) to 4.43 ± 1.72 ng/ml on day 7th of the second dose of PGF_{2α} (Synchromate[®]) injection and increased to 6.57 ± 0.23 on day 13th before the third dose of PGF_{2α} (Synchromate[®]) injection. There was a decrease following the injection of the third dose of the Synchromate[®] to 0.23 ± 0.09 ng/ml at day 20 and an increase was seen at day 27 (6.10 ± 1.01 ng/ml) which continues to increase till 11.93 ± 0.87 ng/ml at day 48 of the treatment and 24th day post mating. For sows in group 3 (Figure 1), mean P₄ concentration increased from 4.8 ± 1.86 ng/ml at first PGF_{2α} (Lutalyse[®]) to 7.00 ± 0.95 ng/ml on day 13th of the second dose of PGF_{2α} (Lutalyse[®]) injection the concentration. There was a decrease following the injection of the second dose of the Lutalyse[®] to 9.87 ± 1.20 ng/ml at day 20 and 0.80 ± 0.36 ng/ml at day 27. An increase was seen at day 34 (5.50 ± 0.29 ng/ml) which continues to increase till 10.83 ± 0.75 ng/ml at day 48 of the treatment and 26th day post mating. For sows in group 4 (Figure 1), mean P₄ concentration increased from 1.77 ± 1.23 ng/ml at first PGF_{2α} (Lutalyse[®]) to 3.90 ± 0.38 ng/ml on day 7th of the second dose of PGF_{2α} (Lutalyse[®]) injection and slightly decreased to 3.87 ± 1.86 ng/ml (though decreased but no luteolysis) on day 13th of the third dose of PGF_{2α} (Lutalyse[®]) injection. There was a decrease following the injection of the third dose of the Lutalyse[®] to 0.63 ± 0.20 ng/ml at day 20 and an increase was seen at day 27 (4.30 ±

1.23 ng/ml) which continues to increase till 10.97 ± 1.01 ng/ml at day 48 of the treatment and 26th day post mating. Overall mean showed no statistical significant difference (P > 0.05) between the groups.

4. Discussion

The results of this study have shown the luteolytic effect of Cloprostenol sodium (Synchromate[®]) and Dinoprost tromethamine (Lutalyse[®]) in porcine *corpora lutea*. In group 1, the basal concentration of P₄ (3.77 ± 1.53 ng/ml) after the injection of PGF_{2α} was an increased (6.50 ± 0.20 ng/ml) after one week of the injection this shows that most of the animals in group 1 were in their early stage of luteal phase making the CL unresponsive to the injected PGF_{2α} as porcine CL is unresponsive to exogenous PGF_{2α} before day 12 which is in agreement with Guthrie and Polge, 1976, Estill *et al.*, 1993, Zannoni *et al.*, 2007, De Rensis *et al.*, 2012 and Kouamo and Kamga, 2013. There was luteolysis following second dose of the PGF_{2α} at day 13 (0.63 ± 0.09 ng/ml) with oestrus signs which is in corroboration with the luteolytic effect of PGF_{2α} on porcine CL as reported by De Rensis *et al.*, 2012. Pregnancy was established following increased P₄ concentration (11.87 ± 0.20 ng/ml) which is in agreement with Boma and Bilkei (2008) who reported P₄ concentration of pregnant sows to be greater than 5ng/ml.



Key:

E = Luteolysis

Day 0 = day of first PGF_{2α} injection

Day 7 = day of second dose PGF_{2α} injection (Groups 2 and 4)

Day 13 = day of second and third dose PGF_{2α} injection (Groups 1 and 3, 2 and 4 respectively)

2Synch = double injection of Synchromate[®] (Cloprostenol sodium)

3Synch = triple injection of Synchromate[®] (Cloprostenol sodium)

2Lut = double injection of Lutalyse[®] (Dinoprost tromethamine)

3Lut = triple injection of Lutalyse[®] (Dinoprost tromethamine)

Figure 1. Progesterone profile of cross bred sows treated with double and triple injections of PGF_{2α} (Synchromate[®] and Lutalyse[®])

In group 2 with triple doses of synchromate®, the basal p4 level (7.13 ± 0.64 ng/ml) decreased to (4.43 ± 1.72 ng/ml) which indicate that the most of the animals were in their late luteal phase on the day of first PGF_{2α} injection which was able to cause decrease in P4 concentration following the first dose of the injection but the second dose given on day 7 (4.43 ± 1.72 ng/ml) there was no reduction in the p4 rather an increased P4 was seen (6.57 ± 0.23 ng/ml) then a decrease in the p4 (0.23 ± 0.09 ng/ml) following the third dose of PGF_{2α} due to the CL responsiveness to PGF_{2α} from day 12 as reported by Estill *et al.*, 1993 and Kouamo and Kamga, 2013 and oestrus was observed in the animals hence complete luteolysis occurred after the third injection of PGF_{2α}. Following oestrus and natural breeding there was significant increase in the p4 concentration from mating till day 48 (11.93 ± 0.87 ng/ml) which indicates pregnancy as reported by (Boma and Bilkei, 2008).

In group 3 the basal p4 concentration (4.8 ± 1.86 ng/ml) was elevated (7.00 ± 0.95 ng/ml) after the first dose of PGF_{2α} which indicates that luteolysis has not taken place and that the animals were in their early luteal phase and reports has shown non responsiveness of porcine CL to exogenous PGF_{2α} as reported by De Rensis *et al.* 2012 and Kouamo and Kamga, 2013. Luteolysis (0.80 ± 0.36 ng/ml) was reported following the second injection of PGF_{2α} which is in agreement with that reported by De Rensis *et al.* 2012. Pregnancy was established (10.83 ± 0.75 ng/ml) which was also in agreement with that reported by Boma and Bilkei (2008).

In group 4 (triple injection of Lutalyse®) the basal p4 concentration (1.77 ± 1.23 ng/ml) indicates that the animals were in their proestrus because no observable signs of oestrus was seen but following the first dose of PGF_{2α} there was a slight though not significant decline in the p4 concentration (3.90 ± 0.38 ng/ml). Luteolysis with observable oestrus signs were reported in this study following the third injection of PGF_{2α} (0.63 ± 0.20 ng/ml) which is in agreement with the works of De Rensis *et al.* 2012, Kouamo and Kamga, 2013 and Przygodzka *et al.* 2015. Pregnancy was established (10.97 ± 1.01 ng/ml) which was also in agreement with that reported by Boma and Bilkei (2008).

5. Conclusions and Recommendations

It is therefore concluded that porcine corpora lutea is refractory to exogenous PGF_{2α} at day 7 of the oestrus cycle but was responsive following injection at day 13. Further work should be done to find out the reason for lack of response of porcine CL to PGF_{2α} between days 7 and 12 and appropriate pregnancy diagnosis should be done to avoid early pregnancy loss as PGF_{2α} is an abortifacient.

REFERENCES

- [1] Ajala, MK, Adesehinwa, AOK and Mohammed, AK (2007). Characteristics of smallholder pig production in Southern Kaduna area of Kaduna State, Nigeria, *American- Eurasian Journal of Agriculture and Environmental Science*, 2: 182-188.
- [2] Boma, MH and Bilkei, G (2008). Field experience with early pregnancy diagnosis by progesterone based ELISA in sows. *Onderstepoort Journal of Veterinary Research*, 75: 55-58.
- [3] Brüssow, KP and Wahner, M (2011). Biological and technological background of oestrus synchronization and fixed-time ovulation induction in the pig. *Biotechnology in Animal Husbandry*. 27(3): 533-545.
- [4] De Rensis, F, Saleri, R, Tummaruk, P, Techakumphu, M and Kirkwood, RN (2012). Prostaglandin F_{2α} and control of reproduction in female swine: A Review. *Theriogenology*, 77: 1-11.
- [5] Diaz, FJ and Wiltbank, MC (2004). Acquisition of luteolytic capacity: Changes in Prostaglandin F_{2α} regulation of steroid hormone receptors and oestradiol biosynthesis in pig corpora lutea. *Biology of Reproduction*, 70: 1333-1339.
- [6] Diaz, FJ, Crenshaw, TD and Wiltbank, MC (2000). Prostaglandin F_{2α} induces distinct physiological responses in Porcine corpora lutea after acquisition of luteolytic capacity. *Biology of Reproduction*, 63: 1504-1512.
- [7] Estienne, MJ, Harper, AF, Horsely, BR, Estienne, CE and Knight, JW (2001). Effects of PG600 on the onset of oestrus and ovulation rate in gilts treated with regumate. *Journal of Animal Science*, 79: 2757-2761.
- [8] Estill, CT (2000). Current concepts in oestrus synchronisation in swine. *American Society of Animal Science*, 77:1-9.
- [9] Estill, CT., Britt, JH and Gadsby, JE (1993). Repeated administration of prostaglandin F_{2α} during the early luteal phase causes premature luteolysis in the pig. *Journal of Biology of Reproduction*, 49: 181-185.
- [10] Gadsby, JE, Balapure, AK, Brt, JH and Fitz, TA (1990). Prostaglandin F_{2α} receptors on enzyme-dissociated pig luteal cells throughout the oestrous cycle. *Endocrinology*, 126:787-795.
- [11] GraphPad Prism Version 5.0 (2009). GraphPad Software, San Diego, California, USA (www.graphpad.com).
- [12] Guthrie, HD and Polge, C (1976). Luteal function and oestrus in gilts treated with a synthetic analogue of prostaglandin F-2alpha (ICI 79,939) at various times during the oestrous cycle. *Journal of Reproduction and Fertility*, 48: 423-425.
- [13] Kouamo, J and Kamga-Waladjo, AR (2013). State-of-art in oestrus synchronisation in pigs. *Revue Africaine de Sante et de Productions Animales*, 11: 110 – 115.
- [14] National Research Council (NRC). (1998). *Nutrient Requirements of Swine*. (10th edition). National Academy Press. Washington D.C. U.S.A.
- [15] Przygodzka, E, Witek, KJ, Kaczmarek, MM, Andronowska, A and Ziecik, AJ (2015). Expression of factors associated with apoptosis in the porcine corpus luteum throughout the

luteal phase of the oestrous cycle and early pregnancy: Their possible involvement in acquisition of luteolytic sensitivity. *Theriogenology*, 83: 535–545.

- [16] Tur, I (2013). General reproductive properties in pigs. *Turkish Journal of Veterinary and Animal Sciences*. 37: 1-5.
- [17] Van Leeuwen, J J, Martens, MR, Jourquin, J, Driancourt, MA, Kemp, B and Soede, NM(2011). Effects of altrenogest treatments before and after weaning on follicular development, farrowing rate and litter size in sows. *Journal of Animal Science*, 89: 2397-2406.
- [18] Zannoni, A, Bernardini, C, Rada, T, Ribeiro, LA, Forni, M and Bacci, ML (2007). Prostaglandin F2-alpha receptor (FPr) expression on Porcine *corpus luteum* microvascular endothelial cells (pCL-MVECs). *Reproductive Biology and Endocrinology*, 5: 31-53.