

Antinutrients, Mineral and Bioavailability Prediction of Calcium and Iron in Eggplant Fruit (*Solanum* spp.) Varieties Consumed in Côte d'Ivoire

Agnan Marie-Michel Combo^{1,*}, Patrick Aubin Dakia², Obouo Hugues Jonathan Gildas Kobi¹

¹Department of Biochemistry-Microbiology, Laboratory of Agrovalorization, Jean Lorougnon Guédé University, Daloa, Côte d'Ivoire

²Department of Food Science and Technology, Nangui Abrogoua University, Abidjan, Côte d'Ivoire

Abstract Eggplant is one of the most commonly consumed vegetables in Côte d'Ivoire. For this purpose, the mineral bioavailability and antinutrient constituents of eggplant varieties (Niguema, N'drowa, Aliona, Bello, Kotobi, Italy, Tiger, Kalenda and Gnanngnan) were evaluated by standard procedures. The antinutrient composition in dry weight basis was significantly ($P < 0.05$) varied and ranged: oxalate 311.32-129.53 mg/100 g, tannin 370.14-100.98 mg/100 g, flavonoid 47.35-15.82 mg/100 g, and phytate 143.30-41.78 mg/100 g. Gnanngnan had the highest tannin content, Italy had the highest content in flavonoid, while, Tiger presented the highest content in both oxalate and phytate. The results also revealed that eggplant fruits contain some percentage of minerals as follow: Mg (93.90-35.84 mg/100 g), P (19.33-11.70 mg/100 g), Fe (5.37-1.15 mg/100 g), Ca (32.26-13.70 mg/100 g) and K (88.13-41.99 mg/100 g). The calculated molar ratio of phytate/Ca was below the critical value excepted Italy molar ratio, while those of phytate/Fe and oxalate/Ca were above the critical value and this indicate the unavailable of calcium and iron in these studied eggplant varieties. However, this tendency may be reversed if boiling or cooking processes are applied. In view of these results, eggplant antinutritional factors, because of their antioxidant activity being able to have an anti-cancerous property, some authors would recommend to limit their degradation during the technological processes.

Keywords Eggplant, Mineral, Antinutrient, Bioavailability

1. Introduction

African vegetables, long considered secondary, are nowadays taken into account at national and international levels. This is explained by their nutritional value in the diet of rural areas. Diets rich in vegetables providing micronutrients and health-promoting phytochemicals could alleviate both under-nutrition and obesity [1].

Eggplant known as aubergine is a plant belonging to the *Solanaceae* family and to the genus *Solanum* [2]. The family *Solanaceae* is prevalent in tropical, subtropical and Mediterranean areas. Eggplant cultivation is possible in varied climates (temperate, tropical dry or humid). It contains different species and varieties which are distinguished in particular by the color, size and shape of the fruits [3]. It is of great economic importance because several species are cultivated and consumed in the world [4]. World eggplant production is around 50 million tons per year [5].

Eggplant contains a significant amount of phytochemicals

especially phenolic compounds, such as caffeic and chlorogenic acid, and flavonoids, such as nasunin. Eggplant, also contain measurable amounts of oxalates [6]. Some authors reported that eggplant improves digestion, fights constipation, curing cancer, high blood pressure and hepatitis [7,8]. It is effective in the treatment of high blood cholesterol [9].

African eggplants include several species including *Solanum macrocarpon*, *S. anguivi* and *S. aethiopicum*. They are in various varietal forms, and bear different vernacular names according to the ethnic groups and the countries. African eggplants are a valuable source of food, in addition to their use for medicinal purposes [10]. In Côte d'Ivoire, eggplants are an essential component that enters in the diet. Eggplant is known for its good palatability and its uses in different dishes. They are important source of vitamins, minerals, dietary fiber and phytochemicals [11].

Moreover, vegetable sources may contain substances harmful for human health, affecting the bioavailability of nutrients. However, to use eggplant as source of nutrients, it is also necessary to demonstrate the presence of antinutritional factors such as tannins, oxalates, phytates etc. Therefore, an attempt was made in the present study to evaluate the mineral and antinutrient compounds of the

* Corresponding author:

comboagnan@yahoo.fr (Agnan Marie-Michel Combo)

Received: Nov. 20, 2020; Accepted: Dec. 9, 2020; Published: Dec. 15, 2020

Published online at <http://journal.sapub.org/food>

different eggplant varieties consumed in Côte d'Ivoire.

2. Materials and Methods

Eggplant fruits, *Solanum aethiopicum* (Niguema, N'drowa, Kotobi, Aliona and Bello varieties), *Solanum melongena* (Italy, Tiger and Kalenda varieties) and *Solanum anguivi* (Gnangnan) used in this study comes from the Laboratory of crop improvement, agroforestry Unit, Jean Lorougnon Guédé University (Daloa, Côte d'Ivoire). The three names Niguema, N'drowa and Gnangnan are from the Akan ethnic group in Côte d'Ivoire. All reagents used in this study were of analytical grade.

2.1. Preparation of Samples

The eggplant fruits were transported to the laboratory. These fruits were washed with tap water and then with distilled water. The consumable parts were finely cut using a stainless steel knife into rings, oven-dried at 62°C for 24 h, and grounded into fine powder and stored in an airtight container for experimental use. Determinations of minerals and phytochemical constituents were carried out by triplicate. The results were expressed on the basis dry matter (mg/100 g DM).

2.2. Phytochemical Analysis

Phytochemical constituents were carried out to determine the secondary metabolites present in the samples. The metabolites determined include tannins, oxalates, phytates and flavonoids.

Oxalate content was determined using a titration method [12], in which 15 mL of H₂SO₄ (3 M) were added to 1 g of eggplant powder. After 1 h of homogenization, the solution is filtered through Watman paper. The filtrate is hot-titrated with a solution of KMnO₄ (0.05 M) until the pink turn is persistent.

Phytate content was determined using the spectrophotometric method of [13], in which the mixture of eggplant powder and 0.65 N HCl (1 g in 20 mL HCl) was stirring for 12 h at room temperature. The mixture obtained is centrifuged at 12000 rpm for 40 min. A volume of 0.5 mL of supernatant was mixed with 3 mL of wade's reagent. The solution was left to rest for 15 min and the absorbance was measured at 490 nm against blank. A calibration curve was carried out with sodium phytate at 10 mg/mL.

Tannin content was determined using the spectrophotometric method described by [14]. 1 mL of the phenolic extract from polyphenol extraction was mixed with 5 mL of vanillin reagent (0.1 mg/mL of vanillin in sulfuric acid 70%). The sample was left to rest for 20 min in the dark and the absorbance was read at 500 nm against blank. Tannin content was estimated using a calibration curve of tannic acid solution (0.1 mg/mL) as standard.

Flavonoid content was performed spectrophotometrically according to the method described by [15]. 0.5 mL of the phenolic extract obtained from the extraction of the

polyphenols was mixed successively with 0.5 mL of distilled water, 0.5 mL of aluminum chloride (10%, w/v), 5 mL of sodium acetate (1 M) and finally 2 mL of distilled water. The mixture was left to rest for 30 minutes at room temperature and the absorbance was read at 415 nm against a blank. A calibration range was carried from a quercetin solution at 0.1 mg/mL.

2.3. Determination of Mineral Contents

The mineral content was analyzed on aliquots of the solution of the ash. Magnesium (Mg), phosphorus (P) and potassium (K) were determined using an atomic absorption spectrophotometer (Thermo Scientific iCE 3000 Series) described in previous work [16]. Calcium (Ca) content was carried out by the titrimetric method with EDTA and iron (Fe) content was determined by TPTZ (2,4,6 tripyridyl-5 triazine) method [17].

2.4. Determination of Molar Ratio of Antinutrients to Minerals

The molar ratio between antinutrient and mineral was obtained according to the method of [18]. Molar ratio = Moles of antinutrient/Moles of mineral = mg antinutrient/MW of antinutrient/ mg mineral/MW of mineral, where MW = atomic weight.

2.5. Statistical Analysis

The statistical processing of the data consisted of an analysis of variance (ANOVA) with a classification criterion using the Statistica software (Statistica 7.1). Means were compared by the Duncan test at the 5% significance level.

3. Results and Discussion

3.1. Antinutritional Factors of Eggplant Varieties

The nutritional importance of a given food depends on the nutrients and antinutritional constituents. High concentrations of antinutrients have been discovered to cause great effects on mineral bioavailability in foods [19] by forming complexes with them, as a result reducing their absorption and utilization by the body systems [20]. Phytochemical analysis revealed that the bioactive constituents: oxalate, tannin, flavonoid and phytate were present in all the eggplant varieties (Table 1). However, the major constituent found were oxalate and tannin.

The oxalate contents based on dry matter varied from 311.32 to 129.53 mg/100 g. The variety Tiger had the highest content (311.32 mg/100 g) while the variety Gnangnan had the least content (129.53 mg/100 g). A significant difference ($p < 0.05$) was observed in oxalate levels. The values obtained in this study are higher than the values reported earlier on four varieties of eggplant [21].

Tannins are known to bind irreversibly to proteins of forming insoluble complexes but also efficiently bind to Fe and to a lesser extent to Cu, Mn, Al, Zn and Co [22]. The

values obtained from the studied eggplant varieties were between 370.14 and 100.98 mg/100 g. The total acceptable tannic acid daily intake for a man is 560 mg/100 g [23]. Hence, the levels of tannin in eggplant varieties are low compared to its critical toxicity value. Therefore, the toxicity effects of the tannin may not be significant.

Regarding phytate, the values obtained from the studied

eggplant varieties ranged between 143.30 and 41.78 mg/100 g. These values were higher than 22.76 - 38.67 mg/100 g DM obtained by [21] but lower than of eggplant leaves (392.23 mg/100 g DM) mentioned by [24]. On the other hand, a phytate diet of 10-60 mg/100 g has been reported to decrease bioavailability of minerals if consumed over a long period of time [25].

Table 1. Antinutrient Contents of Eggplant Varieties

Varieties	Antinutrients (mg/100 g Dry Matter)			
	Oxalate	Tannin	Flavonoid	Phytate
Niguema	160.91 ^a ±6.50	136.65 ^{ac} ±45.35	19.13 ^{acdf} ±1.25	98.58 ^{abd} ±4.20
N'drowa	210.56 ^b ±1.28	184.43 ^b ±13.79	21.59 ^{acdf} ±1.01	110.81 ^{bc} ±2.38
Aliona	161.11 ^a ±6.03	148.00 ^{ab} ±19.06	15.82 ^{cdf} ±0.70	107.92 ^{abc} ±3.64
Bello	147.07 ^a ±0.42	187.70 ^b ±26.44	16.11 ^{cdf} ±0.78	96.24 ^{abd} ±6.23
Kotobi	152.17 ^{ad} ±10.48	167.42 ^{ab} ±26.35	37.42 ^e ±4.17	87.14 ^d ±8.21
Italy	178.22 ^c ±7.31	100.98 ^c ±13.83	47.35 ^e ±5.65	126.31 ^{cf} ±5.27
Tiger	311.32 ^e ±8.21	125.06 ^{ac} ±6.53	17.06 ^{cdf} ±2.87	143.30 ^f ±8.35
Kalenda	153.98 ^{ad} ±7.71	124.52 ^{ac} ±33.91	40.45 ^e ±7.03	66.61 ^e ±29.15
Gnangnan	129.53 ^f ±0.32	370.14 ^e ±13.99	22.17 ^{acdf} ±2.92	41.78 ^g ±2.41

Values are mean ± Standard Deviation of triplicate samples. Values bearing different letters in the same column differ significantly (p < 0.05).

According to [26], flavonoids isolated from eggplant fruits have potent antioxidant activity and hypolipidemic effects. The flavonoid values obtained vary from 47.35 to 15.82 mg/100 g. Flavonoid content of 39.60 mg/100 g for a cultivar of *S. melongena* (*S. incanum*) has been reported [27]. Others reported that flavonoid levels were in the range of 2.53 to 16.88 mg/100 g DM in four varieties of eggplant [21]. This content is lower than the values obtained in this study.

3.2. Mineral Contents of Eggplant Varieties

Minerals are important components of the human diet because they serve as cofactors for many physiological and metabolic processes [28]. The mineral composition of the studied eggplant varieties is presented in Table 2. Analysis of the minerals revealed that the studied eggplants contained high rate of magnesium (Mg), phosphorus (P), iron (Fe), calcium (Ca) and potassium (K). Magnesium and potassium are the minerals with the highest levels, followed by calcium, phosphorus and iron.

The magnesium content is varied from 93.90 to 35.84 mg/100 g DM in Aliona and Kotobi varieties respectively. These values obtained are lower than those of *Solanum melongena* variety 158.10 mg/100 g DM mentioned by [29].

Phosphorus contents were between 19.33 mg/100 g DM (Niguema variety) and 11.70 mg/100 g DM (Tiger variety). Phosphorus levels are much higher than the values (3.72-5.23 mg/100 g DM) obtained by [30] for the variety *Solanum melongena*. The bioavailability of phosphorus in the diet is variable. High ingestion of phosphorus is harmful to the kidneys, bones and blood vessels [31].

Iron is needed for the production of hemoglobin and the transport of oxygen from the lungs through the blood vessels [32]. The iron contents ranged from 5.37 mg/100 g DM

(Bello variety) to 1.50 mg/100 g DM (Gnangnan variety). These values are close to those (2.02-3.71 mg/100 g DM) obtained by [33] for seven varieties of eggplant.

The calcium contents of the studied eggplant varieties varied from 32.26 mg/100 g in Italy variety to 13.70 mg/100 g DM in Bello variety. The values obtained are close to the levels (15.00 and 30.00 mg/100 g DM) obtained by [21] for four varieties of eggplant. Eggplants could therefore be a good source of calcium and could be used as supplements in diets low in calcium.

Finally, the potassium contents ranged from 88.13 mg/100 g DM in Kotobi variety to 41.99 mg/100 g DM in Bello variety. The potassium contents of the studied eggplant varieties are much lower than the levels (238.10-245.37 mg/100 g DM) obtained by [30] for the varieties of *Solanum melongena*.

3.3. Molar Ratio of Antinutrients and Minerals

To predict the bioavailability of iron and calcium, antinutrient mineral ratios were calculated and compared with the reported critical toxicity values. The calculated molar ratios between antinutrients and minerals of the studied eggplant varieties are showed in Table 3. Results also revealed that there was significantly different in the phytate/Fe, phytate/Ca and oxalate/Ca molar ratio of all the varieties.

Phytic acid is considered an antinutrient as it can reduce bioavailability of minerals such as iron, calcium, copper and zinc rendering them biologically unavailable [34,35]. The phytate/Fe molar ratios obtained in all the studied eggplant (8.04-1.52) were higher than the critical value of 0.4 [18]. This indicates that phytate of these eggplants may hinder iron bioavailability.

Table 2. Mineral contents of Eggplant varieties

Varieties	Minerals (mg/100 g Dry Matter)				
	Mg	P	Fe	Ca	K
Niguema	66.66 ^c ±0.19	19.33 ^g ±0.03	1.92 ^a ±0.06	18.16 ^c ±0.05	80.28 ^f ±0.31
N'drowa	71.39 ^d ±0.43	16.95 ^f ±0.10	3.43 ^c ±0.72	27.92 ^c ±0.17	59.19 ^g ±0.36
Aliona	35.84 ^a ±0.95	17.26 ^f ±0.49	2.54 ^b ±0.36	23.27 ^d ±0.62	53.53 ^e ±2.33
Bello	88.37 ^h ±0.28	15.99 ^e ±0.14	5.37 ^d ±0.12	32.26 ^f ±0.09	41.99 ^a ±0.14
Kotobi	93.90 ⁱ ±0.65	14.28 ^e ±0.11	1.88 ^a ±0.05	14.10 ^a ±0.10	88.13 ^h ±0.22
Italy	78.93 ^f ±0.14	15.85 ^e ±0.08	1.88 ^a ±0.11	13.70 ^a ±0.02	84.17 ^g ±0.26
Tiger	65.31 ^b ±1.04	11.70 ^a ±0.19	1.51 ^a ±0.02	19.25 ^c ±0.31	50.91 ^b ±0.81
Kalenda	81.02 ^g ±0.72	12.82 ^b ±0.09	2.03 ^a ±0.20	15.63 ^b ±2.57	70.67 ^e ±0.71
Gnangnan	76.43 ^e ±0.19	15.22 ^d ±0.05	1.50 ^a ±0.06	22.11 ^d ±0.06	52.22 ^d ±0.21

Values are mean ± Standard Deviation of triplicate samples. Values bearing different letters in the same column differ significantly (p < 0.05).

However, the molar ratios of phytate/Ca (0.56-0.11) in all the studied eggplant, excepted Italy variety, were below the critical value of 0.5 known to affect calcium bioavailability [36].

The molar ratios of oxalate/Ca calculated (7.19-2.03) were also higher than the critical level of 1.0 which implies that the high content of oxalate could have adverse effect on bioavailability of calcium in these eggplants [37].

The variations in the calculated molar ratio for the different studied eggplant varieties could be attributed to genetic or varietal differences.

Table 3. Molar Ratio between Antinutrient and Mineral

Varieties	Molar ratio antinutrient/mineral		
	Phytate/Ca	Phytate/Fe	Oxalate/Ca
Niguema	0.33 ^{cd} ±0.01	4.36 ^d ±0.26	3.94 ^{de} ±0.17
N'drowa	0.24 ^{bc} ±0.01	2.82 ^{bc} ±0.61	3.35 ^{cd} ±0.01
Aliona	0.28 ^c ±0.01	3.64 ^{cd} ±0.38	3.08 ^{bc} ±0.12
Bello	0.18 ^{ab} ±0.01	1.52 ^a ±0.08	2.03 ^a ±0.01
Kotobi	0.37 ^{de} ±0.04	3.94 ^d ±0.27	4.80 ^e ±0.30
Italy	0.56 ^f ±0.02	5.73 ^e ±0.56	5.78 ^f ±0.23
Tiger	0.45 ^e ±0.02	8.04 ^f ±0.36	7.19 ^g ±0.17
Kalenda	0.27 ^{bc} ±0.14	2.77 ^{bc} ±1.05	4.47 ^e ±0.82
Gnangnan	0.11 ^a ±0.01	2.36 ^{ab} ±0.05	2.60 ^{ab} ±0.01

Values are mean ± Standard Deviation of triplicate samples. Values bearing different letters in the same column differ significantly (p < 0.05).

4. Conclusions

The present work allows to know the antinutritional, mineral composition and bioavailability of iron and calcium of eggplant fruits consumed in Côte d'Ivoire. The findings obtained shown that eggplants contain high levels of antinutritional factors (oxalate, tannin, flavonoid and phytate) with lower minerals which make iron and calcium unavailable to the human body. However, processing methods such cooking, boiling and other heat treatment would be better to reduce the levels of these antinutrients present in the studied eggplant varieties. Thus, data obtained

in this study provided opportunities for further researches concerning the effect of processing on antinutritional components and the bioavailability of minerals in eggplant.

REFERENCES

- [1] Ezzati, F., Lopez, A.D., Rodgers, A., Hoorn, S.V., and Murray, C.J.L., 2002, Selected major risk factors and global and regional burden of disease, *Lancet*, 360(9343), 1347-1360.
- [2] B. Chérifa, "Etude de l'activité antioxydante des polyphénols extraits de *Solanum melongena* par des techniques électrochimiques," Thesis, Mohamed Khider Biskra University, Algeria, May 2014.
- [3] Furini, A., and Wunder, J., 2004, Analysis of eggplant (*Solanum melongena*) related germplasm: morphological and AFLP data contribute to phylogenetic interpretations and germplasm utilization, *Theoretical and applied genetics*, 108(2), 197-208.
- [4] Sultana, S., Islam, M.N., and Hoque, M.E., 2018, DNA fingerprinting and molecular diversity analysis for the improvement of brinjal (*Solanum melongena* L.) cultivars, *Journal of Advanced Biotechnology and Experimental Therapeutics*, 1(1), 01-06.
- [5] Konan, N.O., Akaffou, M.A., Kouadio L., Akaffou, D.S., and Mergeai, G., 2020, Genetic diversity of exotic and local eggplants (*Solanum* spp.) cultivated in Côte d'Ivoire based on ISSR markers, *Biodiversitas*, 21(8), 3650-3657.
- [6] João, S.D., 2012, Nutritional quality and health benefits of vegetables: A Review, *Food and Nutrition Sciences*, 3, 1354-1374.
- [7] Magioli, C., and Mansur, E., 2005, Eggplant (*Solanum melongena* L.): Tissue culture, genetic transformation and use as an alternative model plant, *Acta Botanica Brasilica*, 19(1), 139-148.
- [8] Silva, M.E., Santos, R.C., O'Leary, M.C., and Santos, R.S., 1999, Effect of aubergine (*Solanum melongena*) on serum and hepatic cholesterol and triglycerides in rats, *Brazilian Archives of Biology and Technology*, 42(3), 339-342.

- [9] Jorge, P.A., Neyra, L.C., and Osaki, R.M., 1998, Effect of eggplant on plasma lipid levels, lipidic peroxidation and reversion of endothelial dysfunction in experimental hypercholesterolemia, *Arquivos. Brasileiros de Cardiologia*, 70(2), 87-91.
- [10] Chioma, A., Obiora, A., and Chukwuemeka, U., 2011, Does the African garden egg offer protection against experimentally induced ulcers? *Asian Pacific Journal of Tropical Medicine*, 4(2), 163-166.
- [11] Cao, G., Sofic, E., and Prior, R., 1996, Antioxidant capacity of tea and common vegetables, *Journal of Agriculture and Food Chemistry*, 44(11), 3426-3431.
- [12] Day R.A. & Underwood A.L., 1986, Quantitative analysis, 5th Ed. In Prentice-Hall. 701 p.
- [13] Mohamed, A.I., Perera, A.J.P. and Hafez, Y.S., 1986, New chromophore method for phytic acid determination, *American Association of Cereal Chemists*, 63(6), 475-478.
- [14] Bainbridge, Z., Tomlins, K., Wellings, K., and Westby, A., 1996, Methods for assessing quality characteristic of non-grain starch staples, *Natural Resources Institute*, 112, 27-29.
- [15] Lamien-Meda, A., Lamien, C.E., Romitto, M., Millogo, J.F., and Nacoulma, O.G., 2005, Determination of the total phenolic, flavonoid and proline contents in Burkina Faso Honey, as well as their radical scavenging activity, *Food Chemistry*, 91(3), 571-577.
- [16] Combo, A.M-M., Dakia, P.A., Niaba, K.P.V., Traoré, N., and Beugré, G.A.M., 2020, Assessment of chemical composition and nutritional value of some varieties of okra available in the market of Daloa (Côte d'Ivoire), *Asian Journal of Agriculture and Food Sciences*, 8(3), 18-27.
- [17] Combo, A.M-M., Kouassi, K.A., Niaba, K.P.V., Kouassi, K.C., Konan, A.D.E., Ado, A.J-M.G., and Beugré, G.A.M., 2020, Nutritional value and microbiological quality of potential complementary foods formulated from the combination of fonio, soybean and mango flour, *International Journal of Innovation and Applied Studies*, 30(2), 633-641.
- [18] Woldegiorgis, A.Z., Abate, D., Haki, G.D., and Ziegler, G.R., 2015, Major, minor and toxic minerals and anti-nutrients composition in edible mushrooms collected from Ethiopia, *Food Processing Technology*, 6(3), 1-8.
- [19] Weaver, C.M., and Kannan S., "Phytate and mineral bioavailability," In: Reddy NR, Sathe SK *Food Phytates*. CRC Press: Boca Raton, FL, pp 211-223, 2002.
- [20] Aletor, V.A., 1995, Compositional studies on edible tropical species of mushrooms, *Food Chemistry*, 54 (3), 265-268.
- [21] Ossamulu, I.F., Akanya, H.O., Jigam, A.A., and Egwim E.C., 2014, Evaluation of nutriment and phytochemical constituents of four eggplant cultivars, *Elixir Food Science*, 73, 26424-26428.
- [22] Naumann, H.D., Tedeschi, L.O., Zeller, W.E., and Huntley, N.F., 2017, The role of condensed tannins in ruminant animal production: advances, limitations and future directions, *Revista Brasileira de Zootecnia*, 46(12), 929-949.
- [23] Gemedé, H.F., Haki, G.D., Beyene, F., Woldegiorgis, A.Z., and Rakshit, S.K., 2016, Proximate, mineral, and antinutrient compositions of indigenous Okra (*Abelmoschus esculentus*) pod accessions: implications for mineral bioavailability, *Food Science & Nutrition*, 4(2), 223-233.
- [24] Oboh, G., Ekperigin, M.M., and Kazeem, M.I., 2005, Nutritional and haemolytic properties of eggplants (*Solanum macrocarpon*) leaves, *Journal of Food Composition and Analysis*, 18, 153-160.
- [25] Elinge, C.M., Muhammad, A., Atiku, F. A., Itodo, A.U., Peni, I.J., and Sanni, O.M., 2012, Proximate, mineral and anti-nutrient composition of pumpkin (*Cucurbita pepo* L) seeds extract, *International Journal of Plant Research*, 2(5), 146-150.
- [26] Chinedu, S.N., Olasumbo, A.C., Eboji, O.K., Emiloju, O.C., Arinola, O.K., and Dania, D.I., 2011, Proximate and phytochemical analyses of *solanum macrocarpon* L. fruits, *Research Journal of Chemical Sciences*, 1(3), 63-71.
- [27] Auta, R., James, S.A., Auta, T., and Sofa, E.M., 2011, Nutritive value and phytochemical composition of processed *solanum incanum* (bitter garden egg), *Science World Journal*, 6(3), 5-6.
- [28] Ibanga, O.I., and Okon, D.E., 2009, Minerals and antinutrients in two varieties of African pear (*Dacryodes edulis*), *Journal of Food Technology*, 7(4), 106-110.
- [29] Scorsatto M., Pimentel A.C., Ribeiro da Silva A.J., Sabally K., Rosa G., Moraes de Oliveira G. M., 2017, Assessment of bioactive compounds, physicochemical composition, and *in vitro* antioxidant activity of eggplant flour, *International Journal of Cardiovascular Sciences*, 30(3), 235-242.
- [30] Agoreyo, B.O., Obansa, E.S., and Obanor, E.O., 2012, Comparative nutritional and phytochemical analyses of two varieties of *solanum melongena*, *Science World Journal*, 7(1), 1597-6343.
- [31] Becker, W., Lyhne, N., Pederson, A.N., Aro, A., Fogelholm, M., Phrdsdotir, I., Alexander, J., Anderssen, S.A., Meltzer, H.M., and Pedersen J.I., 2004, Nordic Nutrition Recommendations: Integrating nutrition and physical activity, *Scandinavian Journal of Nutrition*, 48(4), 178-187.
- [32] Lehninger, A.L., Nelson, D.L., and Cox, M.M, "Lehninger Principles of Biochemistry," (4th ed.) W.H. Freeman and Company, New York, 2005, 1216 pp.
- [33] Ayaz, F.A., Colak, N., Topuz, M., Tarkowski, P., Jaworek, P., Seller, G., and Inceer, H., 2015, Comparison of nutrient content in fruit of commercial cultivars of eggplant (*Solanum melongena* L.), *Polish Journal of Food and Nutrition Sciences*, 65(4), 251-259.
- [34] Food and Agriculture Organization of The United Nations - FAO. Roots, tubers, plantains and bananas in human nutrition. Rome: FAO, (FAO and Food Nutrition series, n. 24). 1990.
- [35] Francis, G., Makkar, H.P.S., and Becker, K., 2001, Anti-nutritional factors present in plant derived alternate fish feed ingredients and their effects in fish, *Aquaculture*, 199(3-4), 197-227.
- [36] Hassan, L.G., Umar, K.J., Dangoggo, S.M., and Maigandi, A.S., 2011, Anti-nutrient Composition and Bioavailability Prediction as Exemplified by Calcium, Iron and Zinc in *Melocia corchorifolia* Leaves., *Pakistan Journal of Nutrition*, 10(1), 23-28.

- [37] Frontela, C., Scarino, M.L., Ferruzza, S., Ros, G., and Martínez, C., 2009, Effect of dephytinization on bioavailability of iron, calcium and from infant cereals assessed in the Caco-2 cell model, *World Journal of Gastroenterology*, 15(16), 1977–1984.