

# Separation and Purification of Lactic Acid from Sisal Wastes

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**Abstract** The aim of this paper was separation and purification of Lactic acid (LA) produced from Sisal Waste by investigating the influence of modified hybrid short path evaporation (M-HSPE) system on the concentration of LA. A  $2^2$  full factorial design was used to study the influence of temperature and time on the concentration of LA and % mass obtained in distillate and residue flask. Minitab V.17 was used in designing the LA concentration experiments and in calculating the effect of each variable and their interactions. Higher LA concentration of 108 g/L occurred at system temperature of 47°C for 25 minutes. This represented a concentration of 4.5 times the initial concentration (24 g/L) with 100% recovery in residues stream/ flask. Therefore, the M-HSPE system was able to concentrate LA to almost twice the original hybrid short path evaporation (HSPE) system. According to ANOVA, LA concentration presented a correlation coefficient of 0.993, which confirm that, linear model correlated well with the experimental data. The system temperature (Temp.), operation time (time) and the interaction between system temperature and time were statistically significant variables for LA concentration at  $\alpha = 0.05$ .

**Keywords** Lactic acid separation, Sisal boles juice, Modified hybrid short path

## 1. Introduction

Lactic acid (LA) can be produced through chemical synthesis or fermentation route and is used in food and chemical industries. In Fermentation route, microorganisms eat sugars containing materials like sisal boles and excrete LA as product. Sisal bole is a stem part of a sisal plant (Agave). It is reported that in Tanzania, only 2% of sisal plant (sisal fibres) is used to produce fibres [1]. The remaining 98% biomass (sisal poles, sisal boles and stubs of leaves that remain on the boles after every cutting) is discarded as waste [2, 3]. Sisal boles waste contain 26-30%(w/v) sugars inform of inulin [4], that could be fermented to produce LA.

After fermentation, the LA fermentation liquor contains LA, together with various impurities such as unreacted raw materials (e.g. sugars), microbial cells and culture media, i.e. derived saccharides, amino acids, carboxylic acids, proteins and inorganic salts [5-9]. Therefore, the separation and purification of LA from other impurities is mandatory to obtain the quality needed in the polymers synthesis [9, 10]. Purification and separation of LA after fermentation involves removal of biomass and other solids from the broth, acidification of the fermented product with strong acids to

liberate the LA, removal of inorganic salts and concentration of LA [5]. However, the procedure for LA purification is rather difficult due to its chemical behaviour, as it shows strong affinity to water and has low volatility [7]. The presence of two adjacent functional groups (acid and alcohol) in a small molecule of LA gives it, its high reactivity with all bases, as well as its tendency to decompose at high temperatures (227°C at atmospheric pressure) [6, 8].

Purification of LA from fermentation broth has been studied using different techniques, including separation with membranes such as nanofiltration and electrodialysis [11-13], adsorption or ion exchange [14, 15], reactive distillation [16], [17] and hybrid short path evaporation [7, 8, 16]. It is reported that, membrane technologies are very potential and have particular advantage of simultaneous separation and concentration of LA [18]. However, membrane separation performance is compromised by fouling which requires frequent cleaning of the dialyzer [11, 18]. The cost of equipment can be relatively high and the method requires high energy input [18] as such, it was not used in this research as the main separation technique.

On one hand, the major disadvantage of the adsorption or ion exchange method is that it requires regeneration of ion exchange resin and adjustment of feed pH to increase the sorption efficiency thus, requiring large amount of chemicals [8, 9, 14]. On the other hand, distillation requires low vacuum that forces higher molecular weight component such as sugar and protein to leave the system as residue. The formation of oligomers (high boiling esters and dimmers)

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limits an overall distillation yield. HSPE was a preferred alternative method for recovery and concentration of LA [8], [19] as it employs lower temperatures of operation (60–80°C), due to application of vacuum, and hence requires short residence time [7]. This method also reduces the tendency of LA decomposition and it is energy efficient [6, 20]. The description of the method and important parameters to evaluate its performance are described by Komesu *et al.*, [8]. The aim of this paper was separation and purification of LA produced from Sisal Waste by investigating the influence of modified hybrid short path evaporation (M-HSPE) system on the concentration of LA.

## 2. Material and Methods

### 2.1. Feed Preparation

The raw material used was fermented broth from sisal boles juice. Boles were chopped and pressed using hydraulic pressing machine operated by hydraulic jack to extract juice which was then hydrolyzed to allow the sugar polymers to break into monomer sugar before fermentation. The hydrolysis method by Busairi, [21] was adapted. The microorganism strain utilised was *Lactobacillus delbrueckii* WLP677 from the White Labs Inc. (CA 92126 USA). The bacterial culture was grown in 50 ml of De Man, Rogosa and Sharpe agar (MRS) medium in 250 ml Erlenmeyer flask [22]. After sterilization at 121°C for 15 minutes using portable steam Autoclave (Heuer, 220V and 50 Hz) and cooling to room temperature (30°C ± 2), the medium was inoculated with 10% inoculum level. The medium was then incubated at 37°C for 24 hrs under stationary conditions for microbial growth before fermentation.

The pH of the fermentation medium was regulated before fermentation to 5 or 6, using 10M sodium hydroxide. The broth was then separated after 72 hrs fermentation. The separation of cells and other insoluble proteins was done by centrifugation at 5000rpm for 15 minutes. The broth was then clarified using ultrafiltration system (Amicon 8400). The system used an inorganic membrane with a molecular weight cut-off (MWCO) of 5kDa (Microdyn-Nadir UP005) to remove micro-particles. The clarified broth was acidified to pH that is less than pKa of LA (3.86) with strong acids (H<sub>2</sub>SO<sub>4</sub> or HCl) to liberate the LA from the sodium lactate formed when sodium hydroxide was used to control fermentation pH. The liberated LA was then subjected to ion exchange resin for more purification.

### 2.2. Multivalent Removal

The clarified fermentation broth was then passed through ion exchange resins in order to remove metal ions like potassium, sodium, magnesium and calcium. The experiment was performed in bulk per batch using a glass column of 3.4 cm diameter and 65 cm length, where 450 g of an ion exchange resin was placed. Only 75% of the column was packed with resins to provide room for expansion of the

resins bed. The anion exchanger used was Indion FFIP (Ion Exchange India Ltd), a weak basic anion exchange resin which contains only tertiary amino acid groups. The cation exchanger resin was Indion 225H (Ion Exchange India Ltd), a strongly acidic macro porous cation-exchange resin with uniform size beads.

The ion exchange resin system, Figure 1, operated simultaneously, that is the feed was first introduced to a cation exchange resin column (1) at the bottom and left the column at the top where it was connected to the anion exchange resin column (2). The broth was fed to the anion exchange resin column at the bottom and left the system at the top of the column to the product reservoir. A peristaltic pump Skan-AG (Watson Marlow Ltd) was used to pump the feed. The experiment was concluded when the processed volume was high enough to saturate the column. The regeneration of the resin was conducted from the column top to the bottom to reduce the consumption of reagents. Backwashing was done using distilled water for 45min at a rate of 1 mL/s before regeneration. The regeneration experiment was done at room temperature using 1.1M HCl and 0.75M NaOH for cation and anion exchanger resin, respectively.

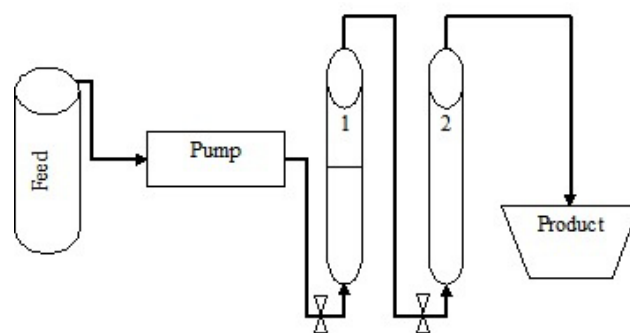


Figure 1. Ion exchange resin system

### 2.3. LA Concentration by M-HSPE

After the removal of ions, cells and other unreacted materials like protein, the LA was concentrated by removing water and other acids like acetic acid. The HSPE system by Komesu *et al.*, [8] that depends on the difference in boiling temperatures (volatility) was adopted as modified hybrid short path evaporation (M-HSPE) system. The LA fermented broth was manually fed into a round bottomed necked flask connected to the system in batches of 200 ml, Figure 2. The system allowed the heated flask with feed connected to another flask (distillate) using the fractionating column to separate and return the dense volatiles back to the flask and the less dense escaped to distillate flask. Digital stirring heating mantle (DSHM 2000 – Azzota Cooperation) with temperature control was used to control the system temperature and provide uniform mixing using magnetic stirrer.

Two condensers were used in the system; one is connected to the distillate flask to condense the distillate that could escape back to the distillate flask. The rotary-vane vacuum

pump two stage (1003317- 3B scientific® Physics) was used to maintain the system with 1 kPa pressure. The second condenser is connected to the pump and third flask (light) was used to freeze the escaped volatiles from distillate flask to collect condensed volatiles that escaped from distillate flask to avoiding their migration to the pump and contaminate the oil. All the condensers were connected to a water bath with temperature controller (serial: 1B1160929-Polystat®) set at 10°C.

A 2<sup>2</sup> full factorial design was used to study the influence of temperature and time on the concentration of LA and % mass (mass of distillate or residue/ sum of residue, distillate and light mass after evaporation) obtained in distillate and residue flask. The real variables and experimental ranges are presented in Table 2. The experiments were performed in random order using two replicates at central points to estimate the pure error. Minitab V.17 was used in designing the concentration experiments and in calculation of the effect of each variable and their interactions. Equation 1 was used to model the relationship between factors and response where  $X_1$  and  $X_2$  are independent variables,  $\beta_0$ ,  $\beta_1$ ,  $\beta_2$  and  $\beta_{12}$  are regression coefficients and  $Y$  is the response function (lactic acid concentration and % mass).

$$Y = \beta_0 + \beta_1 X_1 + \beta_2 X_2 + \beta_{12} X_1 X_2 \quad (1)$$

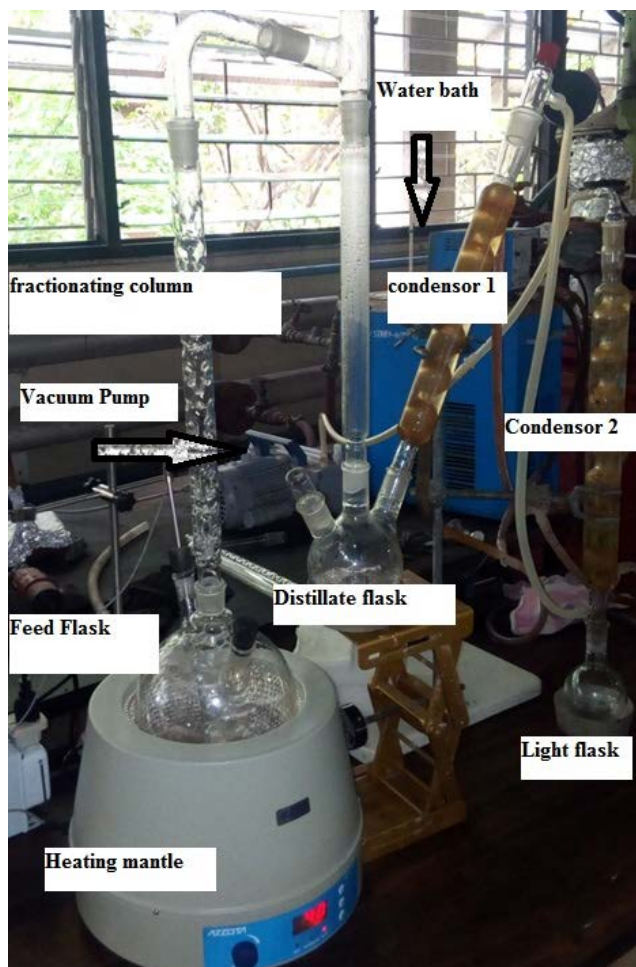


Figure 2. Modified Hybrid Short Path Evaporation System

## 2.4. Analytical Methods

The pH and conductivity of the sample were measured using pH meter (CH-9100 Metrohm) and conductivity meter (B36356 Thermo Scientific), respectively. The amount of metal ions: Potassium, sodium, magnesium and calcium were measured using Atomic Absorption Spectrometer (Varian AA240). Sulphate content was analysed using colometric method [23] to determine total sulphate. Chloride content of the LA was analysed by potentiometric titration as described by APHA, [23]. LA concentration was measured using UV-IV digital spectrophotometer (Labtronics LT-31) following the method by Borshchevskaya *et al.*, [24]. The method was selected since it is relatively cheap and has an error of less than 3% compared to HPLC method. The efficiency of the system on ionic and cationic removal and LA concentration was calculated using the concentration before and after passing the broth in the system. The lactic acid recovery rate was calculated using Equation 2 as per Komesu *et al.*, [8].

$$\text{Rec}(\%) = \frac{m_i x \%LA_i}{\sum_{i=1}^3 m_i x \%LA_i} \quad (2)$$

Where  $i$  = the index for residue, light or distillate,  $m$  is the residue, light or distillate mass (g) and %LA is the fraction of lactic acid in residue, light or distillate.

## 3. Results and Discussion

By means of centrifugation and ultrafiltration the proteins and cells were removed and through ultrafiltration system colour causing compounds like pigments from sisal juice and agar used were removed as the broth was changed from yellow to colourless. The feed to the ion exchange system contained mainly LA, water and inorganic salts. The main cations were sodium, calcium, potassium and magnesium while the main anions were chloride and sulphate. High concentration of Na is due to use of NaOH during fermentation pH regulation. The use of conc. HCL and/or H<sub>2</sub>SO<sub>4</sub> to reduce sample pH to 1 during hydrolysis increase concentration of chloride and sulphate in the product as shown in Table 1. The ionic composition of the fermented broth before and after ion exchange is given in Table 1. The percentage removal was calculated using equation 3:

$$\% \text{ Removal} = 100 * \frac{\Delta C}{C_0} \quad (3)$$

Where  $\Delta C$  = change in concentration (mg/L) and  $C_0$  = initial concentration (mg/L).

From Table 1 it is clearly seen that the ion exchange system was able to remove ions by 99% on dry basis for both cation (K, Mg, Fe, Ca and Na) and anion (Chloride and sulphate). By introducing the crude LA to pre-treatment before concentration using M-HSPE more than 64% of total sugar was removed. The M-HSPE system alone cannot remove all sugars. This is because a total recovery is done in the residue flask and LA has lower molecular weight than residue sugars. Methods like nanofiltration can be used to

remove residue sugars before its concentration by evaporation. The obtained dilute LA was then introduced to concentration system to remove water and other acids that might be present. For this study no other acids were expected since the bacterial used generates only LA in controlled environment [4]. The concentration results of the M-HSPE system are given in Table 2.

**Table 1.** The Ionic Composition of Broth Before and After Ion Exchange (Mean  $\pm$  SDV, n=2)

Component	Before Ion exchange	After Ion exchange	% Removal
K(mg/L)	218.00 $\pm$ 1.98	1.61 $\pm$ 0.43	99.26
Ca(mg/L)	85.79 $\pm$ 2.19	0.59 $\pm$ 0.28	99.31
Mg(mg/L)	21.64 $\pm$ 2.13	0.18 $\pm$ 0.06	99.17
Na(mg/L)	1452.7 $\pm$ 2.4	2.32 $\pm$ 0.46	99.84
Fe(mg/L)	0.57 $\pm$ 0.09	0.01 $\pm$ 0.00	99.12
Chloride(mg/L)	4649.80 $\pm$ 1.75	45.00 $\pm$ 0.01	99.03
Sulphate(mg/L)	2556.25 $\pm$ 2.55	23.90 $\pm$ 0.28	99.07
Conductivity( $\mu$ S/cm)	3997 $\pm$ 3.54	1.79 $\pm$ 0.62	99.95
Residual sugars (g/L)	106 $\pm$ 3.90	37.7 $\pm$ 1.71	64.47

**Table 2.** Two Level Factorial Design Matrix and Experimental Responses

Runs	Real Variables		% Mass			LA conc.	TRS
	Temp. (°C)	Time (min.)	Distillate	Residue	Light	(g/L)	(g/L)
1	43	25	69.0	26.5	4.5	59	4.2
2	43	25	68.5	27.5	4.0	57	4.1
3	47	25	91.5	5.0	3.5	108	7.1
4	45	20	71.0	21.5	7.5	56	3.9
5	47	25	92.5	6.5	1.0	100	7.0
6	45	20	69.0	26.5	4.5	59	4.1
7	43	15	60.0	32.5	7.5	30	2.0
8	43	15	62.5	33.0	4.5	32	2.2
9	47	15	65.0	31.0	4.0	52	3.8
10	47	15	65.0	30.0	5.0	54	3.8

TRS= Total Residual sugars concentration, Temp. = system temperature  $\pm 3^\circ\text{C}$

The results in Table 2 show the mass percentage of the streams (distillate, light and residue), LA concentrations and residue sugars concentrations after concentration using M-HSPE system. Mass percentages were calculated as the ratio between the stream (distillate, residue or light) and sum of all streams after the process. All the LA was found to remain in the residue stream. The distillate stream and the light stream were found to have no LA which can be attributed to the fact that the temperatures ranges used did not decompose LA, hence the preferred temperature ranges. It was possible to remove low volatiles from residue flask to distillate up to more than 90%.

Higher LA concentration of 108 g/L occurred at system temperature of 47°C for a 25 minutes run. This represents the concentration of 4.5 times the initial concentration (24 g/L) before the process. Since the interest of the study was to obtain richer LA fraction, then concentrating the LA at the

residue flask is advantageous. This is because with short time using low temperatures of less than 50°C under the M-HSPE system, the higher amount of water (distillate) has been removed and LA was concentrated to 4.5 times its initial concentration. This amount is higher compared to concentration using the HSPE system reported by Oleivera *et al*, [20], who was able to concentrate 2.3 times initial concentration with the setback that LA was collected in the distillate flask with water hence needed more separation.

The amount of total residue sugars (TRS) which remained in the residue stream is less than 10 g/L, in all runs, with higher concentration of 7.1 g/L at the same temperature and time where LA concentration was also high. This concentration is almost 6% of the total LA concentration which is small compared to what has been reported by other researchers [8, 20]. The results in Table 2 were used to estimate the effects of time and temperature on LA concentration by fitting the data into polynomial Equation (1). Table 3 shows variable effects on the LA concentration and TRS concentration at 95% confidence level.

**Table 3.** Estimated Effects of Parameters on LA Concentration at 95% Confidence Level

LA concentration				
Term	Regr. Coef	SE Coef	T-value	P-value
Constant	61.50	1.03	59.66	0.000
Temp.	17.00	1.03	16.49	0.000
Time	19.5	1.03	18.92	0.000
Temp.*Time	6.00	1.03	5.82	0.002
TRS Concentration				
Constant	4.275	0.0354	120.92	0.000
Temp.	1.15	0.0354	32.53	0.000
Time	1.325	0.0354	37.48	0.000
Temp.*Time	0.3000	0.0354	8.49	0.000

TRS= Total Residual sugars concentration

Considering a confidence level of 95%, a factor is considered statistically significant if its p-value is lower than 0.05. As per Table 3 System temperature (Temp.), operation time (time) and the interaction between system temperature and time were statistically significant variables for LA concentration. This is also well represented by Figure 3. The interaction plots for time and temperature indicates the potential degree of interaction. It is justified by interaction lines that show great departure from parallel.

LA concentration model is represented by Equation 4 and TRS concentration model generated is represented by equation 5.

$$\text{LA conc.} = 141.0 - 3.50 \text{ Temp.} - 23.10 \text{ Time} + 0.600 \text{ Temp.*Time} \quad (4)$$

The model fit for equation (4) showed that, the independent variables Temperature and time and their interaction had a significant effect at p-value =0.00. The correlation coefficient of 0.998 represented by the model justified that the model satisfactorily fit the experimental data.

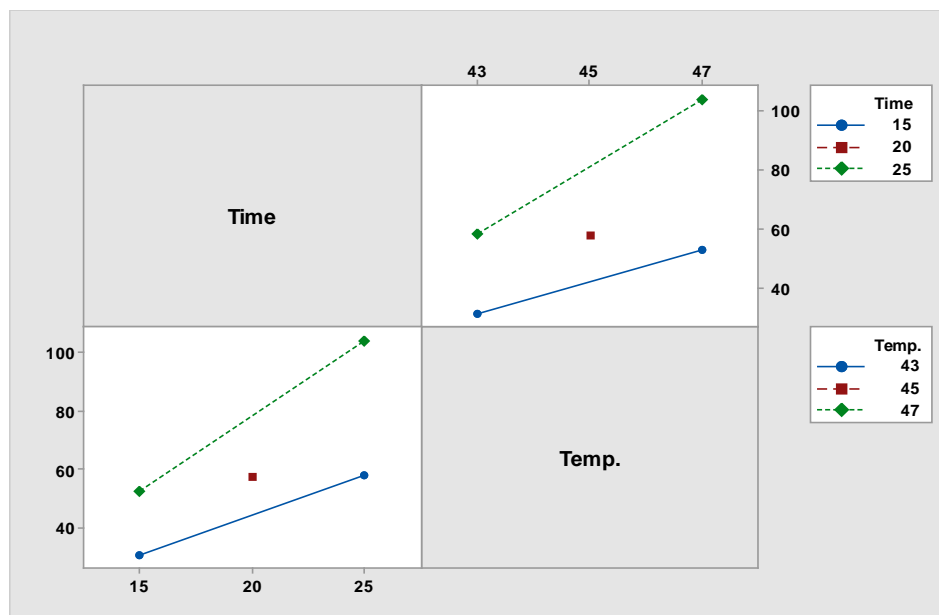


Figure 3. Interaction plot for LA concentration

$$\text{TRS} = 0.10 - 0.0250 \text{ Temp.} - 1.85 \text{ Time} + 0.03000 \text{ Temp.} * \text{Time} \quad (5)$$

Table 4 shows the ANOVA for LA concentration and TRS. In order to evaluate if the models are statistically significant with confidence level of 95% one criterion is to perform an F-Test. According to ANOVA (Table 4), LA concentration presented a correlation coefficient of 0.993, therefore the linear model correlates well with the experimental data. When analysing the results of the ANOVA (Table 4), it is evident that linear and 2- way interactions of the variables temperature and time with p-values that is less than 0.05 significantly affected LA concentration at 95% confidence level.

Table 4. Analysis of Variance (ANOVA) for LA Concentration with 95% Confidence Interval

Source of Variation	DF	Adj SS	Adj MS	F-Value	P-Value
Model	4	5667.60	1416.90	166.69	0.000
Linear	2	5354.00	2677.00	314.94	0.000
Temp.	1	2312.00	2312.00	272.00	0.000
Time	1	3042.00	3042.00	357.88	0.000
2-Way Interactions	1	288.00	288.00	33.88	0.002
Temp.*Time	1	288.00	288.00	33.88	0.002
Pure error	5	42.50	8.50	3.01	
Total	9	5710.10			

DF= Degree of Freedom, Adj SS = adjacent sum of squares, Adj MS =Adjacent Mean squares.

## 4. Conclusions

The clarified fermentation broth used as feed in the ion exchange system in this work contained mainly LA, water and inorganic salts. The main cations were sodium, calcium, potassium and magnesium while the main anions were chloride and sulphate. By means of centrifugation and

ultrafiltration the proteins cells and more than 60% of the residual sugars were removed. Ultrafiltration also removed the colour causing compounds like pigments from sisal juice and agar. The separation system was able to remove ions by 99% on dry basis for both cations (K, Mg, Fe, Ca and Na) and anions (Chloride and sulphate).

All the LA was found to remain in the residue (feed flask) stream. The distillate stream and the light stream were found to have no LA which means therefore, the temperatures ranges used were sufficiently low to condense LA to remain in the feed flask. It was possible to remove more volatiles from residue flask to distillate up to more than 90%. Higher LA concentration of 108 g/L was obtained at a system temperature of 47°C for 25 minutes. This represented LA concentration of 4.5 times the initial LA concentration (24 g/L) before the purification process with 100% recovery in residues stream/ flask. Therefore the M-HSPE system was able to concentrate LA to almost twice the original HSPE.

According to ANOVA, LA concentration presented a correlation coefficient of 0.993, which confirm that, linear model correlated well with the experimental data. The system temperature (Temp.), operation time (time) and the interaction between system temperature and time were statistically significant variables for LA concentration at alpha = 0.05.

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## REFERENCES

- [1] J. C. Terrapon-Pfaff, M. Fishedick, and H. Monheim, 2012. Energy potentials and sustainability—the case of sisal residues in Tanzania, *Energy for Sustainable Development*, 16, (3), 312–319.
- [2] A. Mshandete, O. Kibazohi, and A. Kivaisi, 2013. Tanzania Sisal Industry: Auditing and Characterization of Sisal Post-Harvest Wastes as a Bio-Resource for Bio-Refining, *International Journal of Pure and Applied Sciences*, 14, (142), 84–94.
- [3] C. J. Ngonyani, 2010. Utilization of Sisal Bole Juice – Inulin As an Alternative Chemical Feedstock in Citric Acid Production, Phd Thesis. University of Dar es Salaam.
- [4] N. Msuya, J. H. Y. Katima, E. Masanja, and A. K. Temu, 2017. Poly(lactic-acid) Production from Monomer to Polymer: A review, *Scied Journal of Polymerscience*, 1, (1), 1–15.
- [5] A. N. Vaidya, R. A. Pandey, S. Mudliar, M. S. Kumar, T. Chakrabarti, and S. Devotta, 2005. Production and Recovery of Lactic Acid for Polylactide—An Overview, *Critical Reviews in Environmental Science and Technology*, 35, (5), 429–467.
- [6] A. Komesu, P. F. M. Martinez, B. H. Lunelli, J. Oliveira, M. R. W. Maciel, and R. M. Filho, 2017. Study of Lactic Acid Thermal Behavior Using Thermoanalytical Techniques, *Journal of Chemistry*, 2017, 1–7.
- [7] A. Komesu, P. F. Martins, J. Oliveira, B. H. Lunelli, R. M. Filho, and M. R. W. Maciel, 2014. Evaluation of lactic acid purification from fermentation broth by hybrid short path evaporation using factorial experimental design, *Separation and Purification Technology*, 136, 233–240.
- [8] A. Komesu, P. Martinsa, B. Lunellia, P. F. Martins, B. H. Lunelli, A. T. Morita, P. L. A. De Coutinho, R. M. Filho, and M. R. W. Maciel, 2013. Lactic acid purification by hybrid short path evaporation, *Chemical Engineering*, 32, 2017–2022.
- [9] K. L. Wasewar, 2005. Separation of Lactic Acid: Recent Advances, *Chemical and Biochemical Engineering Quarterly*, 19, (2), 159–172.
- [10] M. Kazunari and Y. Kimura, 2014. PLA Synthesis. From the Monomer to the Polymer, in *Poly(lactic acid) Science and Technology: Processing, Properties, Additives and Applications*, A. Jimenez, P. Mercedes, and R. Ruseckaite, Eds. Royal Society of Chemistry, 1–36.
- [11] V. Habova, K. Melzoch, and M. Rychtera, 2004. Modern Method of Lactic Acid Recovery from Fermentation Broth, *Czech J. Food Sci*, 22, (3), 87–94.
- [12] X.-L. Jiang, X.-J. Feng, J.-M. Wang, Q.-Z. Li, C. Sun, H.-B. Zhang, M. Xian, and H.-Z. Liu, 2016. Recovery Processes of Organic Acids from Fermentation Broths in the Biomass-Based Industry., *Journal of Microbiology and Biotechnology*, 26, (1), 1–8.
- [13] S. Ramaswamy, H. J. Huang, and B. V. Ramarao, Separation and Purification Technologies in Biorefineries. West Sussex: John Wiley&Sons, Ltd, 2013.
- [14] M. I. González, S. Álvarez, F. A. Riera, and R. Álvarez, 2006. Purification of lactic acid from fermentation broths by ion-exchange resins, *Industrial and Engineering Chemistry Research*, 45, (9), 3243–3247.
- [15] W. Groot, J. V. Krieken, and S. de Vos, Production and Purification of Lactic Acid and Lactide, in *Poly(lactic acid): Synthesis, Structure, Properties, Processing and Application*, R. Auras, L. . Lim, S. E. . Selke, and H. Tsuji, Eds. John Wiley & Son, Inc., 2010.
- [16] A. Komesu, M. R. W. Maciel, J. A. R. Oliveira, R. M. Filho, and L. H. da S. Martins, 2016. Purification of Lactic Acid Produced by Fermentation: Focus on Non-traditional Distillation Processes, *Separation & Purification Reviews*, 0, (0), 1–14.
- [17] A. Komesu, P. F. M. Martinez, B. H. Lunelli, R. M. Filho, and M. R. W. Maciel, 2015. Lactic acid purification by reactive distillation system using design of experiments, *Chemical Engineering and Processing: Process Intensification*, 95, 26–30.
- [18] E. Oztekin and S. Altin, 2016. Wastewater Treatment By Electrodialysis System and Fouling Problems, *The Online Journal of Science and Technology*, 6, (1), 91–99.
- [19] A. Komesu, P. F. Martins, J. Oliveira, B. H. Lunelli, R. M. Filho, and M. R. W. Maciel, 2014. Purification of Lactic Acid Produced from Sugarcane Molasses, *Chemical Engineering Transactions*, 37, 367–372.
- [20] R. A. Oliveira, A. Komesu, C. E. Vaz, M. R. Wolf Maciel, and R. Maciel Filho, 2017. Hybrid Short Path Evaporation as an Option to Lactic Acid Recovery from Fermentation Broth, *Chemical Engineering Transactions*, 57, 1–6.
- [21] A. M. Busairi, 2007. Production of Organic Acid From Raw Local Materials', Phd Thesis. Universiti Teknologi Malaysia.
- [22] P. S. Panesar, J. F. Kennedy, C. J. Knill, and M. Kosseva, 2010. Production of L(+) Lactic Acid using *Lactobacillus casei* from Whey, *Brazilian Archives of Biology and Technology*, 53, (1), 219–226.
- [23] APHA, Standard Methods for the Examination of Water and Wastewater, 20th ed. Washington, D.C., 1998.
- [24] L. N. N. Borshchevskaya, T. L. L. Gordeeva, A. N. N. Kalinina, and S. P. P. Sineokii, 2016. Spectrophotometric Determination of Lactic Acid, *Journal of Analytical Chemistry*, 71, (8), 755–758.