

Trans-10,cis-12 Conjugated Linoleic Acid a Novel Inhibitor of Inflammatory Mediators

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Abstract The present study is designed to understand the interaction between trans-10,cis-12 CLA ligand and the inflammatory mediators like COX-2, iNOS by performing molecular docking studies. Probiotic bacteria such as Lactobacilli are capable of converting linoleic acid to conjugated linoleic acid (CLA). CLA is a collective term used to describe a set of 28 distinct positional and geometric isomers of linoleic acid and the most commonly found isomers are cis-9, trans-11 and trans-10, cis-12 both of which possess biological activity. The trans-10,cis-12 CLA was docked into the COX 2 proteins using CDocker CHARMM-based molecular docking algorithm, Accelrys Discovery Studio v 3.5. The binding of COX-2 with CLA isomer revealed that the mode of molecular interaction of trans-10,cis-12 CLA and the standard indomethacin was similar. The *in silico* results suggest that trans-10,cis-12 CLA isomer could be potent anti-inflammatory agents compared to S-ethyl isothiurea and indomethacin.

Keywords Anti-inflammatory, Conjugated linoleic acid, Cyclooxygenase-2, Docking, Indomethacin, Inducible nitric oxide synthase

1. Introduction

Inflammation is an early response to damage instigated by exposure to different pathogens like bacteria, fungi, viruses and a battery of harmful chemical agents which leads to a disrupted tissue homeostasis. Many research reports have demarcated the effect of inflammation in the progression of many diseases such as cancer, atherosclerosis, asthma, and psoriasis. In a majority of these diseases, inflammation has been shown to be a leading cause for pathophysiology of the existing diseases. Chronic inflammation is a long-lasting inflammation that persists for a long time due to continuous stimulus or injury from a persistent pathogen or viral infection. One of the hallmarks of the chronic inflammation is the infiltration of inflammatory cells and fibroblasts leading to prolonged tissue damage [1, 2].

The process of inflammation is mediated by multiple molecular mechanisms; two of the most important are the formation of prostaglandins by cyclooxygenase-2 (prostaglandin synthase) and the production of nitric oxide by the inducible nitric oxide synthase (Figure 1). As per clinical studies and epidemiological studies, ~25% of all cancers are linked to chronic inflammation [3]. Evidence strongly suggests that COX-2 and iNOS play an important role in the development and progression of many tumors [4].

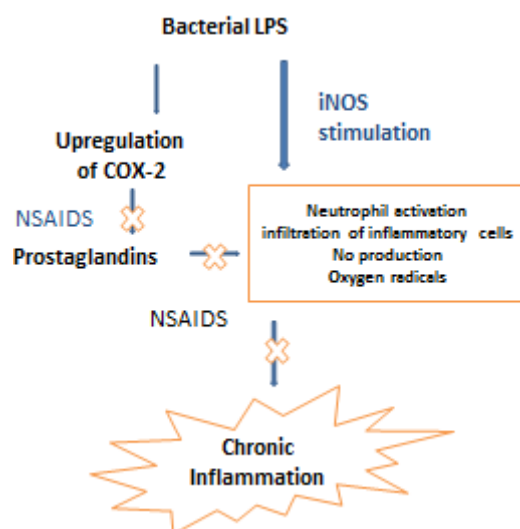


Figure 1. Inflammatory mediators in chronic Inflammation

Probiotics such as *Lactobacillus* and *Bifidobacterium* are ingestible microorganisms and are reported to have numerous health benefits [5, 6]. Additionally these probiotics are capable of producing conjugated linoleic acid (CLA) from linoleic acid [7]. CLAs are heterogeneous geometric isomers derived from linoleic acid. The conjugated double bonds may be of *cis* or *trans* configuration at positions 9 and 11 or 10 and 12 [8, 9]. Among the CLA isoforms, the cis-9, trans-11 and trans-10,cis-12 forms are predominant and found in the greatest abundance in ruminant fatty tissue and. [10] CLA

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has attracted much attention as a novel type of functional lipid due to its biological activities. With this background, the present study is designed to investigate the efficacy of trans-10,cis-12 conjugated linoleic acid as strong anti-inflammatory agent.

2. Methodology

2.1. Generation of Trans-10,cis-12 CLA Ligand Structure for Docking

The 3D structure of trans-10,cis-12 CLA (PubChem ID: 5282800) was retrieved from PubChem database. The structure was cross-checked against Zinc database ([HTTP://ZINC.dock.org](http://ZINC.dock.org)), [11]. The resultant structure was optimized and used for docking using CDOCKER of Discovery studio version v 3.5.

2.2. Construction of COX-1 and COX-2 Homology Models

The amino acid sequence of human COX-2 (P35354.2) with a length of 604 amino acids was obtained from UniProt protein sequence database and it was blasted against Protein Data Bank (PDB) entries to find similar sequences and their structures. The templates for COX-1 and COX-2 were identified as 2OYE (Figure.2A) and 1PXX (Figure.2B) respectively. The COX-1 and COX-2 homology structures were built against the templates structures 2OYE (*Ovis aries*) and 1PXX (*Mus musculus*). The 3D crystal structure of 1PXX bound to indomethacin with resolution of 2.90 [Å] was retrieved from PDB. The 3D structure of COX-2 were verified in Accelrys DS with verify protein tool which calculates the likelihood of each residue to be found in its specific local environment and scores the model confirmation using a statistical potential function. The selected model was further analyzed by VERIFY 3D and the energy minimized structures were superimposed with the model structures using Discovery studio visualize [12].

2.3. Docking of CLA Isomers into the Modeled Structures of COX-1 and COX-2

The trans-10,cis-12 (Figure-3A) was docked into the COX 2 proteins using CDOCKER CHARMM-based molecular docking algorithm, Accelrys Discovery Studio v3.5 (Biosystems Hewlett-Packard, Palo Alto, California, United States). The active site of COX-2 (Figure-3B) protein was defined around the bound inhibitor indomethacin. Inhibitor was removed from the binding site, and chain A was selected for docking. The trans-10,cis-12 CLA isomer was docked against COX-2 and the docking score was compared with standard inhibitor such as indomethacin.

2.4. Docking of CLA Isomer into the Active Site of iNOS

The protein coordinates of iNOS (PDB_ID:4NOS) was retrieved from Protein data bank (www.rcsb.org). The trans-10,cis-12 CLA and Ethyl-isothiourea (S-Ethyl

isothiourea; iNOS-Inhibitor; (PUBCHEM_ID:CID 5139) were docked into the active site of iNOS using CDOCKER [13]. The CHARMM force field was applied along with a grid extension of 8 Å and the partial charges for ligand was set up by Momany-Rone partial charges method. The trans-10,cis-12 CLA isomer along with Ethyl-isothiourea (Figure-3C) was docked with iNOS protein individually into all spheres created around the active sites predicted.

3. Results and Discussion

Molecular docking is a key tool in structural molecular biology and computer-assisted drug design and is useful as it allows a better understanding of molecular events occurring at the binding interface of protein - ligand interaction. It helps in complementing and validating the experimental data. Docking studies of trans-10,cis-12 CLA was carried out to evaluate the most likely target proteins of the inflammatory cascade and to determine the binding mechanism of CLA isomers with target protein using CDOCKER of Accelrys Discovery studio v 3.5.

In silico approaches were employed to develop potent COX-2 inhibitors that are attractive and potential leads for various human cancers and inflammatory disorders. The crystal structure of COX-2 could be useful to provide a better understanding about the active sites and the protein-inhibitor binding mechanism [14, 15]. Docking studies were performed using crystal structure of COX-2 complexed with indomethacin. For molecular docking, indomethacin bounded with the crystal structure of COX-2 was removed and docked with trans-10, cis-12 CLA isomer along with indomethacin. The computer docking of standard indomethacin against COX-2 exhibited three H-bonds, one between the carboxyl hydrogen of Indomethacin and ARG-120 with bond distance of 2.32 Å and the second between OH of Indomethacin and SER353 with 1.85 Å of distance and the third bond with OH of Indomethacin and SER350 (Figure-4) whereas trans-10, cis-12 CLA isomer showed two hydrogen bonds with ARG120 and SER353 (Figure-5). The CDOCKER energies with COX-2 for the ligand indomethacin and trans-10,cis-12 CLA were -37.03, - and -50.47, (Table-1). The trans-10, cis-12 CLA isomer used in the present study against COX-2 showed higher binding energy than standard anti-inflammatory drugs indomethacin [15]. As per the earlier reports the carboxylate group of NSAIDS interacts with TYR385 and SER530 and stabilizes the negative charge of the tetrahedral intermediate and demonstrated that TYR385 and SER530 have a structural and functional role in the chelation of NSAIDS ligand.

The X-Ray Diffraction structure of human inducible Nitric oxide synthase, PDB_ID 4NOS with 427 amino acid length was considered for docking analysis. Ligands were removed and chain A was considered for docking. Heteroatoms were removed, and polar atoms were added to the structures. The trans-10,cis-12 CLA isomer along with 2-ethyl-isothiourea were docked into the active sites of 4NOS using Discovery

studio. Ethyl isothiurea showed 5 hydrogen bonds with GLU358 (2.46Å), GLN142 (2.44 Å), GLU358 (2.14Å), SER486 (2.01Å) and GLU145 (2.17Å) (Figure-6) whereas trans-10,cis-12 CLA isomer exhibited 2 hydrogen bonds with the residue LYS103(Figure-7). The docking energies of ethyl isothiurea and trans-10,cis-12 CLA isomer with

4NOS were -23.1, and -50.36 kcal/mol respectively (Table 2). However, the binding energies of CLA isomers are higher than ethyl isothiurea.

3.1. Figures and Tables

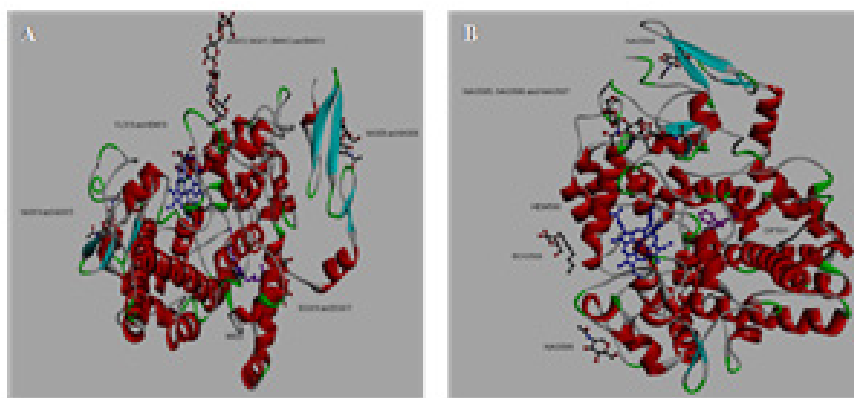


Figure 2. (A). Homology model structure of COX-1 using the template 2OYE; (B). Homology model structure of COX-2 template with IPXX

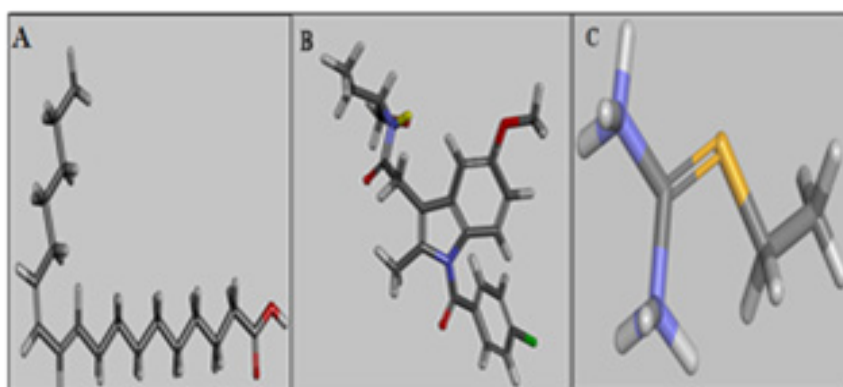


Figure 3. Structures of Ligands (A) trans10,cis-12 CLA (B) Indomethacin and (C) Ethylisothio urea (Grey-Carbon, Red-Oxygen, Blue-Nitrogen, White-Hydrogen and Yellow-Sulfur)

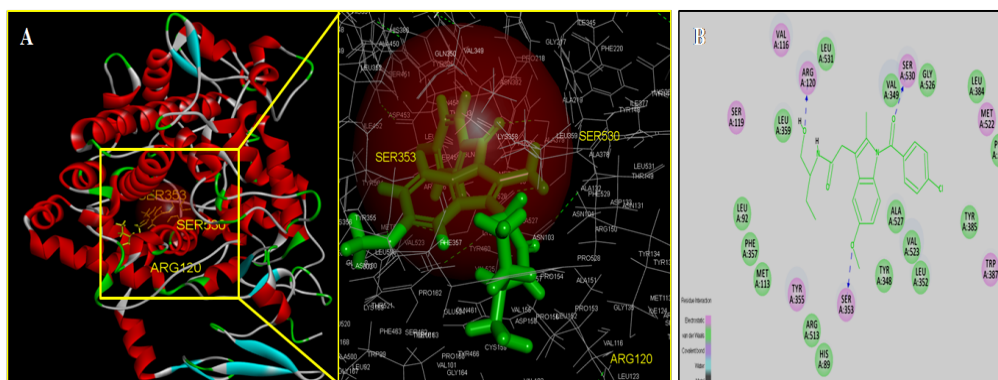


Figure 4. Docking of Indomethacin with COX-2 using Discovery studio v3.5. (A). Indomethacin (green) in the active pocket (red) of COX-2 (ribbon structure); (B). The 2D Hydrogen bonding (blue arrows) between indomethacin and COX-2 residues

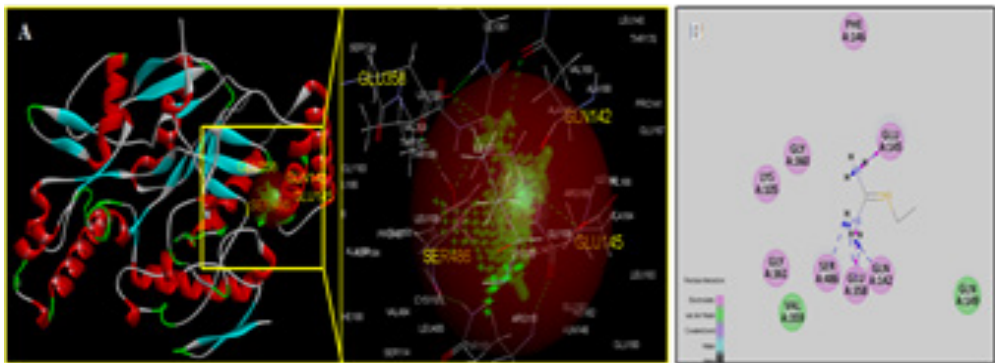


Figure 5. Docked confirmation of trans-10,cis-12 CLA isomer into the binding pocket of COX-2. (A). Ribbon structure of COX-2 with trans-10,cis-12 (green); (B). The 2D Hydrogen bonding between trans-10,cis-12 and COX-2 residues

Table 1. Docking statistics of CLA ligands with COX-2

Receptor	Ligand	No. of Hydrogen Bonds	Amino acids involved in Hydrogen bonding	Bond distance (Å)	CDOCKER interaction energy (kcal/mol)
COX-2	Indomethacin	3	ARG120	2.326	-37.03
			SER353	1.855	
			SER530	1.962	
	trans-10,cis-12 CLA	2	ARG120	2.059	-50.47
			SER353	2.361	

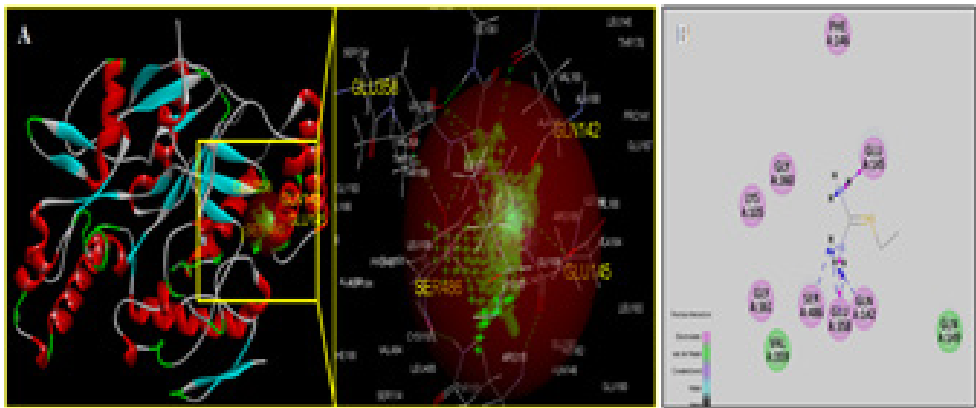


Figure 6. Ligand Ethyl-isothiourea docked into the binding site of iNOS (PDB_ID:4NOS) using Discovery studio v3.5. (A). Ethyl-isothiourea (green) in the active sphere of iNOS; (B). Hydrogen bonding of ethyl-isothiorurea with iNOS residues

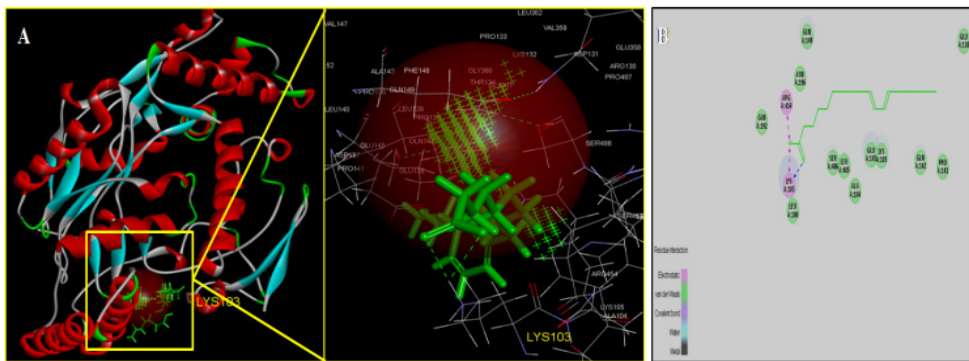


Figure 7. Binding confirmation of trans-10,cis-12 CLA isomer into the active site of iNOS (PDB_ID:4NOS). (A). trans-10,cis-12 (green) in the active sphere of iNOS; (B). Hydrogen bonding of trans-10,cis-12 with iNOS residues

Table 2. Docking energies and the number of hydrogen bond interactions of CLA ligands docked against iNOS using DS CDOCKER

Receptor	Ligand	No. of Hydrogen Bonds	Amino acids involved in Hydrogen bonding.	Bond distance (Å)	CDOCKER interaction energy (kcal/mol)
4NOS	Ethyl-isothiurea	5	GLU358	2.46	-23.1
			GLN142	2.44	
			GLU358	2.14	
			SER486	2.01	
			GLU145	2.17	
	trans-10, cis-12 CLA	2	LYS103	1.904	-50.36

4. Conclusions

The CLA isomer was docked into the proteins of inflammatory cascade by using CHARMM-based molecular docking algorithm (CDOCKER), Accelrys Discovery Studio v3.5 (Biosystems Technologies, San Diego, CA, USA) on Z800 workstation. The docking result of COX-2 with CLA isomer revealed that the mode of molecular interaction of trans-10,cis-12 CLA was similar to the standard indomethacin. The docking results deduce that trans-10,cis-12 CLA isomer of CLA docked into the same pocket of indomethacin. The CLA isomer demonstrated stronger interaction than standard NSAID indomethacin as docking score is representative of the binding energy of ligand to the receptor. The trans-10,cis-12 CLA isomer was docked at the predicted active site of 4NOS along with S-ethyl isothiurea, a potent inhibitor of iNOS.

REFERENCES

- [1] Aggarwal, B.B., R.V. Vijayalekshmi, and B. Sung, *Targeting inflammatory pathways for prevention and therapy of cancer: short-term friend, long-term foe*. Clin Cancer Res, 2009. 15(2): p. 425-30.
- [2] Lin, W.W. and M. Karin, *A cytokine-mediated link between innate immunity, inflammation, and cancer*. J Clin Invest, 2007. 117(5): p. 1175-83.
- [3] Balkwill, F. and A. Mantovani, *Cancer and inflammation: implications for pharmacology and therapeutics*. Clin Pharmacol Ther, 2010. 87(4): p. 401-6.
- [4] Kapoor, A., et al., *The histone variant macroH2A suppresses melanoma progression through regulation of CDK8*. Nature, 2010. 468(7327): p. 1105-9.
- [5] Guarner, F., et al., *Should yoghurt cultures be considered probiotic?* Br J Nutr, 2005. 93(6): p. 783-6.
- [6] Hill, C., et al., *Expert consensus document. The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic*. Nat Rev Gastroenterol Hepatol, 2014. 11(8): p. 506-14.
- [7] Ogawa, J., et al., *Production of conjugated fatty acids by lactic acid bacteria*. J Biosci Bioeng, 2005. 100(4): p. 355-64.
- [8] Lawson, R.E., A.R. Moss, and D.I. Givens, *The role of dairy products in supplying conjugated linoleic acid to man's diet: a review*. Nutr Res Rev, 2001. 14(1): p. 153-72.
- [9] Roche, H.M., E. Noone, and A.N. Gibney, *Conjugated linoleic acid: a novel therapeutic nutrient?* Nutr Res Rev, 2001. 14(1): p. 173-88.
- [10] Jensen, R.G., *The composition of bovine milk lipids: January 1995 to December 2000*. J Dairy Sci, 2002. 85(2): p. 295-350.
- [11] Irwin, J.J., et al., *ZINC: a free tool to discover chemistry for biology*. J Chem Inf Model, 2012. 52(7): p. 1757-68.
- [12] Bikadi, Z. and E. Hazai, *Application of the PM6 semi-empirical method to modeling proteins enhances docking accuracy of AutoDock*. J Cheminform, 2009. 1: p. 15.
- [13] Wu, G., et al., *Detailed analysis of grid-based molecular docking: A case study of CDOCKER-A CHARMM-based MD docking algorithm*. J Comput Chem, 2003. 24(13): p. 1549-62.
- [14] Reddy, R.N., et al., *Computer aided drug design approaches to develop cyclooxygenase based novel anti-inflammatory and anti-cancer drugs*. Curr Pharm Des, 2007. 13(34): p. 3505-17.
- [15] Navya Atluri, Rashmi Holur, Vasavi Thirumalanadhuni Uma Maheswari Devi Palempall et al., *Modulation Of Pro-Inflammatory Genes By α -Mangostin From Garcinia mangostana* International Journal of Pharmaceutical Science Invention 2014. 3 (5): p.23-29.
- [16] Kurumbail, R.G., et al., *Structural basis for selective inhibition of cyclooxygenase-2 by anti-inflammatory agents*. Nature, 1996. 384(6610): p. 644-8.