

# Study of Serum Hecpidin, Iron and Ferritin in Chronic Hepatitis C Patients

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**Abstract Background:** Chronic hepatitis C (CHC) is often associated with increased serum iron and ferritin which may cause more liver damage and rapid progression to cirrhosis and hepatocellular carcinoma. Hecpidin, a hepatic hormone which inhibits iron absorption, is considered the main regulator of iron metabolism. Reduced serum hepcidin in CHC patients is thought to be the main cause of these changes. **Aim:** The aim of this study was to assess changes in serum hepcidin, ferritin and iron in chronic HCV patients. **Subjects and Methods:** Serum hepcidin, ferritin and iron were studied in 45 patients with CHC and 25 healthy controls. **Results and Conclusions:** Serum hepcidin was significantly lower while serum iron and ferritin were significantly higher in CHC patients than controls.

**Keywords** Chronic hepatitis C (CHC), Hecpidin, Ferritin, Iron

## 1. Introduction

Hepatitis C virus infection is one of the main causes of chronic liver disease worldwide [1]. Hepatitis C viral infection affects more than 200 million people worldwide, but most of them have no idea about their infection or of their ensuing hepatic disease [2].

The long-term hepatic impact of HCV infection is highly variable from minimal changes to chronic hepatitis, cirrhosis and hepatocellular carcinoma [3]. Patients with CHC frequently develop mild to moderate iron overload [4].

Many experimental and clinical studies, though not all, suggest that excessive iron in CHC is a cofactor promoting the progression of liver damage and increasing the risk of fibrosis, cirrhosis, and HCC [5, 6]. Hepatic iron concentration has been inversely associated with the response to antiviral therapy [7, 8].

Chronic hepatitis C often appears to be associated with disturbances in iron homeostasis, with elevated serum ferritin and hepatic iron stores in approximately 50% of patients [9]. However, little is known about the mechanism of excess iron accumulation during chronic HCV infection [10].

The importance of iron as a co-morbid factor in CHC is

emphasized by several reports of greater fibrosis, and greater risks of HCC development with more hepatic iron [11]. Conversely, iron reduction has consistently been associated with reductions in serum ALT levels, severity of hepatic necro-inflammation, and risk of development of HCC. Iron reduction has also led to improved responses to IFN-based therapy of CHC [7].

Many hypotheses explained the accumulation of iron in CHC, by the local release of iron from necrotic hepatocytes and HCV-induced perturbation of liver iron homeostasis [12].

With the discovery of hepcidin, the liver has emerged as the central organ in the regulation of systemic iron homeostasis [12]. Hecpidin is a 25-aminoacid peptide hormone primarily synthesized by hepatocytes and it negatively controls two critical steps of iron homeostasis: duodenal absorption and the release from macrophages [13, 14].

At the molecular level, hepcidin binds to ferroportin, the membrane iron exporter highly expressed by enterocytes and macrophages [10]. This results in ferroportin internalization and degradation and hence reduction of iron entry in the plasma compartment [14].

Hecpidin expression is modulated by iron stores, so that it decreases in iron deficiency to facilitate iron absorption while it increases in iron repletion to prevent pathological overload. Hecpidin expression is also induced by inflammation and suppressed by hypoxia and anemia [15, 16].

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Disruption of hepcidin regulation has been postulated as a possible mechanism causing iron overload in acquired conditions, including alcoholic liver disease and CHC [8]. Studies in animal and cellular models have suggested that HCV infection may directly modulate hepcidin expression by HCV-induced reactive oxygen species (ROS) [17].

## 2. Subjects and Methods

This study was carried out on 45 chronic HCV patients selected from Internal and Tropical Medicine Departments Clinics and inpatients at Alzhraa University Hospital, Al-Azhar University, Cairo, Egypt and 25 healthy volunteers as controls.

Patients with severe anemia or other causes of chronic liver disease (CLD) were excluded from the study.

Participants were classified into two groups: • Group I: included 45 patients with chronic HCV infection. • Group II: Included 25 healthy controls.

All participants were subjected to full history taking; thorough clinical examination; abdominal ultrasound and the following laboratory tests liver function tests (total/direct bilirubin, alanine aminotransferase (ALT), aspartate aminotransferase (AST), total proteins, and albumin), prothrombin time (PT), INR, complete blood count (CBC), Serum iron, transferrin saturation and ferritin. Also; HCVab & PCR, HBsAg & HBcAb were done.

Assessment of serum hepcidin: blood samples were collected from each participant by venipuncture in empty centrifuge tubes: incubated in water bath at 37°C for 15 minutes then centrifuged at 3500 rpm. Sera were separated and stored at – 80°C until measurements of serum hepcidin levels. The quantitative detection of commercially available Human Hepcidin (Hepe) ELISA Kit was performed. The quantitative sandwich enzyme immunoassay technique provided by AMS Biotechnology (Europe) Ltd, UK. Catalogue Number: E01H0051, according to the manufacturer's instructions. Informed oral consent was obtained from all participants before starting the study.

**Statistical Analysis:** Data were expressed as mean±SD and percentages. Continuous variables were compared between the two groups with Student's "t" test. Categorical variables were compared by X2 test, and the Fisher exact test when appropriate. One-sided analysis of variance (ANOVA) was used to assess differences among the two studied groups. P-value of less than 0.05 was considered significant. Data analysis was performed with the SPSS Statistical Package, version 20.

## 3. Results

Patients with CHC(Group I) included 39 males and 6 females while Group II included 16 males and 9 females as controls, their mean ages were 44.9 ± 6.3 in Group I and 27.7 ± 9.4 in Group II (Table 1).

According to modified Child-Pugh score[18] patients in group I included 16 (35.5%) class A, 26 (57.8%) class B and 3(6.7%) class C (Table 2).

A highly significant difference was found between groups I and II as regards total bilirubin, ALT, AST, albumin, PT and INR, while there was no significant difference in Hb or total leucocytic count(TLC) (Table 3).

**Table (1).** Age and sex distribution in the studied groups

	CHC (Group I) (n=45)	Controls (Group II) (n=25)
<b>Age (yr)</b>		
Range	29 – 62	23 – 55
Mean ± SD	44.9 ± 6.3	27.7 ± 9.4
<b>Sex</b>		
Male	39 (87%)	16 (64%)
Female	6 (13%)	9 (36%)

**Table (2).** Modified Child Pugh score in Group I

	(n=45)	%
Child Pugh Class A	16	35.5
Child Pugh Class B	26	57.8
Child Pugh Class C	3	6.7

**Table (3).** The laboratory parameters in the studied groups

Laboratory parameters	CHC (Group I) (n=45) Mean ± SD	Controls (Group II) (n =25) Mean ± SD	P value
Hemoglobin (gm/dl)	12.44 ± 1.15	13.21 ± 1.32	0.034
PLT (×10 <sup>3</sup> /μl)	190.90 ± 60.71	253.81 ± 72.92	≤0.001
T.L.C (×10 <sup>3</sup> /μl)	6.7 ± 1.92	7.1 ± 1.81	0.352
ALT (IU/L)	66.14 ± 21.15	26.88 ± 4.51	≤0.001
AST (IU/L)	52.11 ± 18.28	29.15 ± 7.25	≤0.001
Albumin (gm)	3.5 ± 0.5	4.8 ± 0.7	<0.001
T. Bil (mg/dl)	0.95 ± 0.30	0.69 ± 0.19	≤0.001
PC %	81 ± 7.8	99.5 ± 0.8	<0.001
INR	1.02 ± 0.05	0.9 ± 0.01	≤0.001

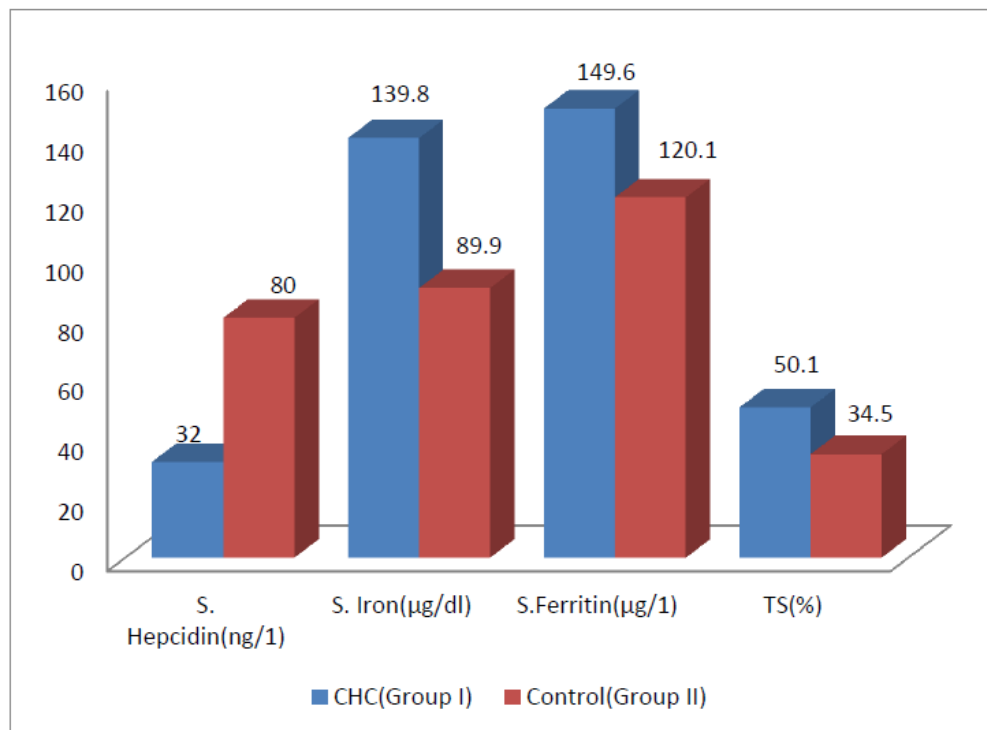
P value < 0.05 = significant

P value ≤0.001 = highly significant

**Table (4).** Serum hepcidin level and iron profile in the studied groups

Laboratory parameters	CHC (Group I) (n=45) Mean ± SD	Controls (Group II) (n=25) Mean ± SD	P value
S. Hepcidin (ng/l)	32 ± 3.9	80 ± 7	≤0.001
S. Iron (μg/dl)	139.8 ± 61.5	89.9 ± 33.9	≤0.001
S. Ferritin (μg/l)	149.6 ± 58.7	120.1 ± 28.2	≤0.001
Transferrin saturation (TS) (%)	50.1 ± 7.6	34.5 ± 11.8	≤0.001

P value ≤0.001 = highly significant



**Figure (1).** Hepcidin and iron, ferritin and transferrin saturation in CHC patients and control group

A highly significant decrease in serum hepcidin was found in group I in comparison to group II, while a highly significant increase was found in serum iron, transferrin saturation (TS) and ferritin in group I compared to group II (Table 4) & (Figure 1).

## 4. Discussion

The hepatic hormone hepcidin is the major regulator of iron metabolism which inhibits iron absorption and recycling from erythrophagocytosis [19].

Patients with CHC often have increased iron, a condition associated with reduced response to antiviral therapy, more rapid progression to cirrhosis, and development of hepatocellular carcinoma [20, 21].

This study demonstrated that hepcidin level was significantly lower in Chronic HCV patients than controls. This goes in agreement with Guo *et al.* [11] and Lange *et al.* [22] they speculated that hepcidin expression in HCV is determined by the opposing effects of hepcidin-suppressive viral factors and the hepcidin stimulation by iron load.

Low level of hepcidin leads to iron overload leading to inflammation and liver fibrosis [21]. However, our results were not in agreement with Fujita *et al.* [23] who found that hepcidin levels did not differ significantly from those in healthy controls, likely because of both methodological imprecision and the very low number of controls enrolled.

Serum iron, ferritin and transferrin saturation were significantly higher in CHC patients compared to the control group and these findings were in agreement with Hörl and Schmidt [21] who found a strong correlation between serum

hepcidin, serum ferritin, transferrin and iron levels.

Our findings were also in agreement with Deicher and Hörl [24] who performed a comprehensive analysis of the role of serum ferritin and its genetic determinants in the pathogenesis and treatment of CHC and stated that elevated serum ferritin levels may reflect a systemic inflammatory state as well as increased iron storage, both of which may contribute to an unfavorable outcome of chronic hepatitis C (CHC).

As regards levels of hepcidin, serum iron and ferritin, our results were also in agreement with Girelli *et al.* [25] who demonstrated that CHC hepcidin concentrations were significantly lower than those of controls. Hepcidin down regulation is associated with iron accumulation and increased serum ferritin. Also, our results were in agreement with Vagu *et al.* [26] who found that, Patients with chronic hepatitis C (CHC) often have elevated serum iron markers, which may worsen liver injury.

## 5. Conclusions

Serum hepcidin level in patients with HCV was significantly lower compared to controls. The consequences of this dysregulation of the hepcidin expression by CHC is an important mechanism in increased iron load in those patients and this may have significant implications for the management of chronic HCV infection. Improvement for hepcidin regulation may be beneficial for HCV patients with iron overload. Further studies and also follow up of CHC patients are needed to refine on hepcidin regulation in the full clinical spectrum of HCV.

## 6. Recommendations

Further studies are recommended to understand hepcidin's molecular control and to determine if supplementation of hepcidin is beneficial in CHC patients. Studies also should be carried out on the role of iron depletion in the treatment of hepatitis C in patients with raised iron concentration.

## REFERENCES

- [1] Lawitz E, Mangia A, Wyles D, Rodriguez-Torres M, Hassanein T, Gordon SC, *et al.* (2013). Sofosbuvir for previously untreated chronic hepatitis C infection. *N Engl J Med*, 368(20): 1878-1887.
- [2] Antonio C, Jean-Michel P and Heiner W (2011). EASL Clinical Practice Guidelines: Management of hepatitis C virus infection *Journal of Hepatology*, 55: 245–264.
- [3] Aoki CA, Rossaro L, Ramsamooj R, Brandhagen D, Burritt MF and Bowlus CL (2005). Liver hepcidin mRNA correlates with iron stores, but not inflammation in patients with chronic hepatitis C. *J Clin Gastroenterol*, 39: 71–74.
- [4] Costa E, Swinkels DW, Laarakkers CM, Rocha-Pereira P, Rocha S, Reis F, *et al.* (2009). Hepcidin serum levels and resistance to recombinant human erythropoietin therapy in hemodialysis patients. *Acta Haematol*, 122(4): 226–229.
- [5] Nicolas G, Chauvet C, Viatte L, Danan JL, Bigard X, Devaux I, *et al.* (2002). The gene encoding the iron regulatory peptide hepcidin is regulated by anemia, hypoxia, and inflammation. *J Clin Invest*, 110(7): 1037–1044.
- [6] Ganz T (2003). Hepcidin, a key regulator of iron metabolism and mediator of anemia of inflammation. *Blood*, 102(3): 783–788.
- [7] Gardenghi S, Ramos P, Marongia MF, Melchiori L, Breda L, Guy E, *et al.* (2010). Hepcidin as a therapeutic tool to limit iron overload and improve anemia in  $\beta$ -thalassemic mice. *J Clin Invest*, 120(12): 4466–4477.
- [8] Hino K, Nishina S and Hara Y (2013). Iron metabolic disorder in chronic hepatitis C: mechanisms and relevance to hepatocarcinogenesis. *J Gastroenterol Hepatol*, 28(4):93-98.
- [9] Jaroszewicz J, Rogalska M, Flisiak I and Flisiak R (2010). Successful antiviral therapy is associated with a decrease of serum prohepcidin in chronic hepatitis C. *World J Gastroenterol*, 16(14): 1747-1752.
- [10] Lavanchy D (2011). Evolving epidemiology of hepatitis C virus. *Clin Microbiol Infect*, 17(2): 107–115.
- [11] Guo P, Cui R, Chang YZ, Wu WS, Qian ZM, Yoshida K, *et al.* (2009). Hepcidin, an antimicrobial peptide is downregulated in ceruloplasmin-deficient mice. *Peptides*, 30(2): 262-266.
- [12] Miyachi H, Kobayashi Y, Relja B, Fujita N, Iwasa M, Gabazza EC and Takei Y (2011). Effect of suppressor of cytokine signaling on hepcidin production in hepatitis C virus replicon cells. *Hepatology Research*, 41(4): 364–374.
- [13] Girelli D, Pasino M, Goodnough JB, Nemeth E, Guido M, Castagna A, *et al.* (2009). Reduced serum hepcidin levels in patients with chronic hepatitis C. *Journal of Hepatology*, 51(5): 845–852.
- [14] Jaroszewicz J, Rogalska M, Flisiak I and Flisiak R (2010). Successful antiviral therapy is associated with a decrease of serum prohepcidin in chronic hepatitis C. *World J Gastroenterol*, 16(14): 1747-1752.
- [15] Anderson DS, Heeney MM, Roth U, Menzel C, Fleming MD and Steen H (2010). A high throughput MALDI-TOF mass spectrometry method for quantification of hepcidin in human urine. *Anal Chem*, 82(4): 1551-1555.
- [16] Cybula M and Szemraj J (2013). The role of hepcidin and polymorphisms in the regulatory region of the IL-28B gene in HCV infections. *Postepy Hig Med Dosw*, 67: 1273-1282.
- [17] Castagna A, Campostrini N, Zaninotto F and Girelli D. (2010). Hepcidin assay in serum by SELDI-TOF-MS and other approaches. *Journal of proteomics*, 73(3):527-536.
- [18] Pugh RN, Murray-Lyon IM, Dawson JL, Pietroni MC and Williams R (1973). Transection of the oesophagus for bleeding oesophageal varices. *Br J Surg*, 60:646–649.
- [19] Piperno A, Mariani R, Trombini P, and Girelli D (2009). Hepcidin modulation in human diseases: from research to clinic. *World J Gastroenterol*, 15(5): 538-551.
- [20] Liu H, Trinh TL, Dong H, Keith R, Nelson D and Liu C (2012). Iron regulator hepcidin exhibits antiviral activity against hepatitis C virus. *PLoS One*, 7(10): e46631.
- [21] Hörl WH and Schmidt A (2013). Low hepcidin triggers hepatic iron accumulation in patients with hepatitis C. *Nephrol Dial Transplant*, 29(6): 1141-1144.
- [22] Lange CM, Kutalik Z, Morikawa K, Bibert S, Cerny A, Dollenmaier G, *et al.* (2012). Serum ferritin levels are associated with a distinct phenotype of chronic hepatitis C poorly responding to pegylated interferon-alpha and ribavirin therapy. *Hepatology*, 55(4): 1038-1047.
- [23] Fujita N, Sugimoto R, Motonishi S, Tomosugi N, Tanaka H, Takeo M, *et al.* (2008). Patients with chronic hepatitis C achieving a sustained virological response to peginterferon and ribavirin therapy recover from impaired hepcidin secretion. *J Hepatol*, 49(5): 702–710.
- [24] Deicher R and Hörl WH (2006). New insights into the regulation of iron homeostasis. *Eur J Clin Invest* 2006; 36: 301-309.
- [25] Girelli D, Pasino M, Goodnough JB, Nemeth E, Guido M, Castagna A, *et al.* (2009). Reduced serum hepcidin levels in patients with chronic hepatitis C. *Journal of Hepatology*, 51(5): 845–852.
- [26] Vagu C, Sultana C and Ruta S (2013). Serum Iron Markers in Patients With Chronic Hepatitis C Infection. *Hepat Mon*, 13(10): e13136.